

International Union of Pharmacology. LIII. Nomenclature and Molecular Relationships of Voltage-Gated Potassium Channels

GEORGE A. GUTMAN, K. GEORGE CHANDY, STEPHAN GRISSMER, MICHEL LAZDUNSKI, DAVID MCKINNON, LUIS A. PARDO, GAIL A. ROBERTSON, BERNARDO RUDY, MICHAEL C. SANGUINETTI, WALTER STUHMER, AND XIAOLIANG WANG

Department of Microbiology and Molecular Genetics (G.A.G.) and Physiology and Biophysics (K.G.C.), University of California, Irvine, Irvine, California; Department of Applied Physiology, Universitat Ulm, Ulm, Germany (S.G.); Institut de Pharmacologie Moléculaire et Cellulaire, Centre National de la Recherche Scientifique, Valbonne, France (M.L.); Department of Neurobiology and Behavior, The State University of New York at Stony Brook, Health Sciences Center, Stony Brook, New York (D.M.); Department of Physiology, University of Wisconsin-Madison, Madison, Wisconsin (G.A.R.); Department of Physiology, Neuroscience, and Biochemistry, New York University School of Medicine, New York, New York (B.R.); Eccles Institute of Human Genetics, University of Utah, Salt Lake City, Utah (M.C.S.); Max Planck Institute for Experimental Medicine, Abt. Molekulare Biologie Neuronaler Signale, Gottingen, Germany (L.A.P., W.S.); and Institute of Materia Medica, Chinese Academy of Medical Sciences, and Peking Union Medical College, Beijing, China (X.W.)

Introduction

Potassium-selective channels are the largest and most diverse group of ion channels, represented by some 70 known loci in the mammalian genome. The first cloned potassium channel gene was the *Drosophila* voltage-gated *shaker* channel, and this was rapidly followed by the identification of other voltage- and ligand-gated potassium channel genes in flies, mammals, and many other organisms. The voltage-gated K_v channels, in turn, form the largest family of some 40 genes among the group of human potassium channels, which also includes the Ca^{2+} -activated (K_{Ca}), inward-rectifying (K_{IR}), and two-pore (K_{2P}) families described in the following articles of this compendium. K_v and K_{Ca} channels together constitute the six/seven-transmembrane group of potassium-selective channels, made up of subunits containing six or seven membrane-spanning domains, including the positively charged S4 segment, which confers on some of these channels their voltage sensitivity.

Table 1 lists the International Union of Pharmacology (IUPHAR¹) names assigned to the members of the K_v family of channels, as well as the gene names established by the HUGO Gene Nomenclature Committee (HGNC). Two new sequences, $K_v6.4$ and $K_v8.2$, have been added to this list since the earlier edition of this compendium. Figures 1 and 2 show two phylogenetic tree reconstructions, one for the K_v1-9 families and the other for the K_v10-12 families, based on amino acid sequence alignments of the entire hydrophobic core of the proteins.

Address correspondence to: Dr. George A. Gutman, Department of Microbiology and Molecular Genetics, University of California, Irvine, Irvine, CA. E-mail: gagutman@uci.edu

Article, publication date, and citation information can be found at <http://pharmrev.aspetjournals.org>.
 doi:10.1124/pr.57.4.10.

¹ Abbreviations: IUPHAR, International Union of Pharmacology; HGNC, HUGO Gene Nomenclature Committee.

TABLE 1

K_v channel families

Gene names shown are those assigned by the IUPHAR (Catterall et al., 2002) and HGNC (<http://www.gene.ucl.ac.uk>) in addition to some other commonly used names.

IUPHAR	HGNC	Other
$K_v1.1$	<i>KCNA1</i>	<i>Shaker</i> -related family
$K_v1.2$	<i>KCNA2</i>	
$K_v1.3$	<i>KCNA3</i>	
$K_v1.4$	<i>KCNA4</i>	
$K_v1.5$	<i>KCNA5</i>	
$K_v1.6$	<i>KCNA6</i>	
$K_v1.7$	<i>KCNA7</i>	
$K_v1.8$	<i>KCNA10</i>	
$K_v2.1$	<i>KCNB1</i>	<i>Shab</i> -related family
$K_v2.2$	<i>KCNB2</i>	
$K_v3.1$	<i>KCNC1</i>	<i>Shaw</i> -related family
$K_v3.2$	<i>KCNC2</i>	
$K_v3.3$	<i>KCNC3</i>	
$K_v3.4$	<i>KCNC4</i>	
$K_v4.1$	<i>KCND1</i>	<i>Shal</i> -related family
$K_v4.2$	<i>KCND2</i>	
$K_v4.3$	<i>KCND3</i>	
$K_v5.1$	<i>KCNF1</i>	Modifier
$K_v6.1$	<i>KCNG1</i>	Modifiers
$K_v6.2$	<i>KCNG2</i>	
$K_v6.3$	<i>KCNG3</i>	
$K_v6.4$	<i>KCNG4</i>	
$K_v7.1$	<i>KCNQ1</i>	<i>KVLQT</i> <i>KQT2</i>
$K_v7.2$	<i>KCNQ2</i>	
$K_v7.3$	<i>KCNQ3</i>	Modifiers
$K_v7.4$	<i>KCNQ4</i>	
$K_v7.5$	<i>KCNQ5</i>	
$K_v8.1$	<i>KCNV1</i>	Modifiers
$K_v8.2$	<i>KCNV2</i>	
$K_v9.1$	<i>KCNS1</i>	Modifiers
$K_v9.2$	<i>KCNS2</i>	
$K_v9.3$	<i>KCNS3</i>	
$K_v10.1$	<i>KCNH1</i>	<i>eag1</i>
$K_v10.2$	<i>KCNH5</i>	<i>eag2</i>
$K_v11.1$	<i>KCNH2</i>	<i>erg1</i>
$K_v11.2$	<i>KCNH6</i>	<i>erg2</i>
$K_v11.3$	<i>KCNH7</i>	<i>erg3</i>
$K_v12.1$	<i>KCNH8</i>	<i>elk1, elk3</i>
$K_v12.2$	<i>KCNH3</i>	<i>elk2</i>
$K_v12.3$	<i>KCNH4</i>	<i>elk1</i>

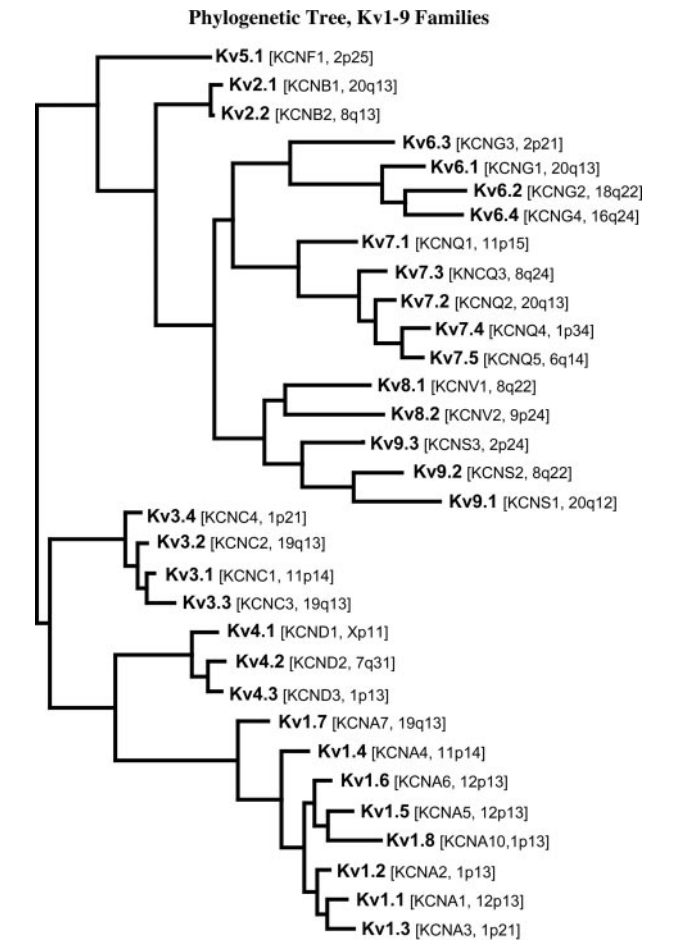


FIG. 1. Phylogenetic tree for the K_v 1–9 families. Amino acid sequence alignments of the human channel K_v proteins were created using CLUSTALW, and analysis by maximum parsimony using PAUP* resulted in unrooted trees comprising the K_v 1– K_v 6 and K_v 8– K_v 9 families that appeared in the previous edition of this compendium. Sequences of K_v 7.1–7.5, K_v 6.4, and K_v 8.2 were added to the existing alignment, and these new sequences were incorporated into the existing tree topology by use of a combination of maximum parsimony and neighbor-joining analysis. Only the hydrophobic cores (S1–S6) were used for analysis. The IUPHAR and HGNC names are shown together with the genes' chromosomal localization and other commonly used names.

K_v channels form an exceedingly diverse group, much more so than one would predict simply based on the number of distinct genes that encode them. This diversity arises from several factors. 1) *Heteromultimerization*. Each K_v gene encodes a peptide subunit, four of which are required to form a functional channel. K_v channels may be homotetramers but may also be heterotetramers formed between different subunits within the same family (in the case of the K_v 1, K_v 7, and K_v 10 families), and these diverse heterotetramers express properties that may be considerably different from those of any of the homotetramers. 2) *“Modifier” subunits*. Four of the K_v families (K_v 5, 6, 8, and 9) encode subunits that act as modifiers. Although these do not produce functional channels on their own, they form heterotetramers with K_v 2 family subunits, increasing the functional diversity within this family. 3) *Accessory proteins*. A variety of other peptides has also been shown to

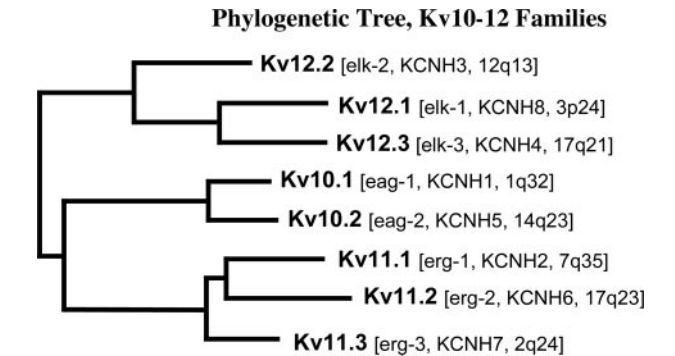


FIG. 2. Phylogenetic tree for the K_v 10–12 families. This unrooted tree was created as described in Fig. 1 and appeared in the previous edition of this compendium. The IUPHAR and HGNC names are shown together with the genes' chromosomal localization and other commonly used names.

associate with K_v tetramers and modify their properties, including several β subunits (which associate with K_v 1 and K_v 2 channels), KCHIP1 (K_v 4), calmodulin (K_v 10), and minK (K_v 11), as well as many others identified in the tables that follow the text of this article. 4) *Alternate mRNA splicing*. A number of K_v channel genes are known to contain intronless coding regions, including all of the K_v 1 family genes (with the sole exception of K_v 1.7) and K_v 9.3. Although alternate splicing of noncoding exons may be important in regulating the expression of these channels, one gene can produce only a single kind of protein subunit. However, various members of the K_v 3, 4, 6, 7, 9, 10, and 11 gene families have coding regions made up of several exons that are alternately spliced, providing yet another significant source of K_v channel functional diversity. 5) *Post-translational modification*. Many K_v channels can be post-translationally modified by phosphorylation (Jereng et al., 2004), ubiquitinylation (Henke et al., 2004), and palmitoylation (Gubitosi-Klug et al., 2005), which in turn modifies channel function.

Our current understanding of the roles of this family of channels is catalogued in Tables 2 through 41, including recent developments in the pharmacology, regulation of expression, and disease associations of its various members (Misonou and Trimmer, 2004; Norton et al., 2004; Wua and Dworetzky, 2005).

REFERENCES

Catterall WA, Chandy KG, and Gutman GA, eds. (2002) *The IUPHAR Compendium of Voltage-gated Ion Channels*. IUPHAR Media, Leeds, UK.
Gubitosi-Klug RA, Mancuso DJ, and Gross RW (2005) The human Kv1.1 channel is palmitoylated, modulating voltage sensing: identification of a palmitoylation consensus sequence. *Proc Natl Acad USA* **102**:5964–5968.
Henke G, Maier G, Wallisch S, Boehmer C, and Lang F (2004) Regulation of the voltage gated K^+ channel Kv1.3 by the ubiquitin ligase Nedd4-2 and the serum and glucocorticoid inducible kinase SGK1. *J Cell Physiol* **199**:194–199.
Jereng HH, Pfaffinger PJ, and Covarrubias M (2004) Molecular physiology and modulation of somatodendritic A-type potassium channels. *Mol Cell Neurosci* **27**: 343–369.
Misonou H and Trimmer JS (2004) Determinants of voltage-gated potassium channel surface expression and localization in mammalian neurons. *Crit Rev Biochem Mol Biol* **39**:125–145.
Norton RS, Pennington MW, and Wulff H (2004) Potassium channel blockade by the sea anemone toxin ShK for the treatment of multiple sclerosis and other autoimmune diseases. *Curr Med Chem* **11**:3041–3052.
Wua YJ and Dworetzky SI (2005) Recent developments on KCNQ potassium channel openers. *Curr Med Chem* **12**:453–456.

TABLE 2
K_V1.1 channels

Channel name	K _V 1.1 ^{1–6}
Description	Voltage-gated potassium channel, delayed rectifier
Other names	HuK (I), MBK1, MK1, RCK1, RBK1, HBK1
Molecular information	Human: 494 aa, NM_000217, chr. 12p13.3, ^{7,8} <i>KCNA1</i> , GeneID: 3736, PMID: 1349297 ³⁵ Mouse: 495aa, NM_010595, chr. 6 Rat: 495aa, NM_173095, chr. 4q42
Associated subunits	K _V β ₁ , K _V β ₂ , PSD95, synapse-associated protein 97 (SAP97), SNAP25 ^{9–19}
Functional assays	Voltage-clamp
Current	Voltage-gated potassium channel in neurons and skeletal muscle
Conductance	10pS ²⁰
Ion selectivity	K ⁺ (1) > Rb ⁺ (0.8) > NH ₄ ⁺ (0.1)
Activation	V _a = −32 mV; k _a = 8.5 mV; τ _n = 5 ms (−32 mV) ^{20,21}
Inactivation	V _h = −51 mV; k _h = 3 mV; τ _h = 11 s (40 mV) ^{20,21}
Activators	None
Gating inhibitors	None
Blockers	Tetraethylammonium (0.3 mM), DTX (20 nM), DTX-K, ShK (16 pM), 10- <i>N</i> -methylcarbamoyl-3,7-bis(dimethylamino)phenothiazine (490 nM), 4-aminopyridine (290 μM), capsaicin (29 μM), resiniferatoxin (9 μM), flecainide (209 μM), nifedipine (96 μM), diltiazem (144 μM), kaliotoxin (41 nM), hongotoxin-1, margatoxin ^{20,22–24}
Radioligands	¹²⁵ I-DTX, ¹²⁵ I-BgK ^{25,26}
Channel distribution	Brain, heart, retina, skeletal muscle, islets ^{27–31}
Physiological functions	Maintaining membrane potential, modulating electrical excitability in neurons and muscle
Mutations and pathophysiology	Episodic ataxia/myokymia syndrome type 1 ^{8,32–34}
Pharmacological significance	Not established
Comments	K _V 1.1 can coassemble with others in the K _V 1 family members in heteromultimers, but not with members of other K _V families; intronless coding region; mammalian <i>Shaker</i> -related family

aa, amino acids; chr., chromosome; DTX, dendrotoxin; ShK, *Styehodactyla helianthus* toxin; BgK, *Bundosoma granulifera* toxin.

1. Tempel BL, Jan YN, and Jan LY (1988) Cloning of a probable potassium channel gene from mouse brain. *Nature (Lond)* **332**:837–839.

2. Baumann A, Grupe A, Ackermann A, and Pongs O (1988) Structure of the voltage-dependent potassium channel is highly conserved from *Drosophila* to vertebrate central nervous systems. *EMBO J* **7**:2457–2463.

3. Kamb A, Weir M, Rudy B, Varmus H, and Kenyon C (1989) Identification of genes from pattern formation, tyrosine kinase, and potassium channel families by DNA amplification. *Proc Natl Acad Sci USA* **86**:4372–4376.

4. McKinnon D (1989) Isolation of a cDNA clone coding for a putative second potassium channel indicates the existence of a gene family. *J Biol Chem* **264**:8230–8236.

5. Christie MJ, Adelman JP, Douglass J, and North RA (1989) Expression of a cloned rat brain potassium channel in *Xenopus* oocytes. *Science (Wash DC)* **244**:221–224.

6. Chandy KG, Williams CB, Spencer RH, Aguilar BA, Ghanshani S, Tempel BL, and Gutman GA (1990) A family of three mouse potassium channel genes with intronless coding regions. *Science (Wash DC)* **247**:973–975.

7. Wymore RS, Korenberg JR, Kinoshita KD, Aiyar J, Coyne C, Chen XN, Hustad CM, Copeland NG, Gutman GA, Jenkins NA, et al. (1994) Genomic organization, nucleotide sequence, biophysical properties, and localization of the voltage-gated K⁺ channel gene KCNA4/Kv1.4 to mouse chromosome 2/human 11p14 and mapping of KCNC1/Kv3.1 to mouse 7/human 11p14.3-p15.2 and KCNA1/Kv1.1 to human 12p13. *Genomics* **20**:191–202.

8. Browne DL, Gancher ST, Nutt JG, Brunt ER, Smith EA, Kramer P, and Litt, M (1994) Episodic ataxia/myokymia syndrome is associated with point mutations in the human potassium channel gene, KCNA1. *Nat Genet* **8**:136–140.

9. Curran ME, Landes GM, and Keating MT (1992) Molecular cloning, characterization, and genomic localization of a human potassium channel gene. *Genomics* **12**:729–737.

10. Scott VE, Rettig J, Parcej DN, Keen JN, Findlay JB, Pongs O, and Dolly JO (1994) Primary structure of a β subunit of α-dendrotoxin-sensitive K⁺ channels from bovine brain. *Proc Natl Acad Sci USA* **91**:1637–1641.

11. Rettig J, Heinemann SH, Wunder F, Lorra C, Parcej DN, Dolly JO, and Pongs O (1994) Inactivation properties of voltage-gated K⁺ channels altered by presence of β-subunit. *Nature (Lond)* **369**:289–294.

12. Wang Z, Kiehn J, Yang Q, Brown AM, and Wible BA (1996) Comparison of binding and block produced by alternatively spliced Kvβ1 subunits. *J Biol Chem* **271**:28311–28317.

13. Stephens GJ, Cockett MI, Nawoschik SP, Schecter LE, and Owen DG (1996) The modulation of the rate of inactivation of the mKv1.1K⁺ channel by the β subunit, Kvβ1 and lack of effect of a Kvβ1 N-terminal peptide. *FEBS Lett* **378**:250–252.

14. Heinemann SH, Rettig J, Graack HR, and Pongs O (1996) Functional characterization of Kv channel β-subunits from rat brain. *J Physiol* **493**:625–633.

15. Jing J, Peretz T, Singer-Lahat D, Chikvashvili D, Thornhill WB, and Lotan, I (1997) Inactivation of a voltage-dependent K⁺ channel by β subunit Modulation by a phosphorylation-dependent inter-action between the distal C terminus of α subunit and cytoskeleton. *J Biol Chem* **272**:14021–14024.

16. Rhodes KJ, Strassle BW, Monaghan MM, Bekele-Arcuri Z, Matos MF, and Trimmer JS (1997) Association and colocalization of the Kvβ1 and Kvβ2 β-subunits with Kv1 α-subunits in mammalian brain K⁺ channel complexes. *J Neurosci* **17**:8246–8258.

17. Kim E, Niethammer M, Rothschild A, Jan YN, and Sheng M (1995) Clustering of Shaker-type K⁺ channels by interaction with a family of membrane-associated guanylate kinases. *Nature (Lond)* **378**:85–88.

18. Doyle DA, Lee A, Lewis J, Kim E, Sheng M, and MacKinnon R (1996) Crystal structures of a complexed and peptide-free membrane protein-binding domain: molecular basis of peptide recognition by PDZ. *Cell* **85**:1067–1076.

19. Tiffany AM, Manganas LN, Kim E, Hsueh YP, Sheng M, and Trimmer JS (2000) PSD-95 and SAP97 exhibit distinct mechanisms for regulating K⁺ channel surface expression and clustering. *J Cell Biol* **148**:147–158.

20. Ji J, Tsuk S, Salapatek AM, Huang X, Chikvashvili D, Pasyk EA, Kang Y, Sheu L, Tsushima R, Diamant NE, et al. (2002) The 25-kDa synaptosome-associated protein (SNAP-25) binds and inhibits delayed rectifier potassium channels in secretory cells. *J Biol Chem* **277**:20195–21204.

21. Grissmer S, Nguyen AN, Aiyar J, Hanson DC, Mather RJ, Gutman GA, Karmilowicz MJ, Auperin DD, and Chandy KG (1994) Pharmacological characterization of five cloned voltage-gated K⁺ channels, types Kv1.1, 1.2, 1.3, 1.5, and 3.1, stably expressed in mammalian cell lines. *Mol Pharmacol* **45**:1227–1234.

22. Stuhmer W, Ruppersberg JP, Schroter KH, Sakmann B, Stocker M, Giese KP, Perschke A, Baumann A, and Pongs O (1989) Molecular basis of functional diversity of voltage-gated potassium channels in mammalian brain. *EMBO J* **8**:3235–3244.

23. Robertson B, Owen D, Stow J, Butler C, and Newland C (1996) Novel effects of dendrotoxin homologues on subtypes of mammalian Kv1 potassium channels expressed in *Xenopus* oocytes. *FEBS Lett* **383**:26–30.

24. Kalman K, Pennington MW, Lanigan MD, Nguyen A, Rauer H, Mahnir V, Paschetto K, Kem WR, Grissmer S, Gutman GA, et al (1998) ShK-Dap²², a potent Kv1.3-specific immunosuppressive polypeptide. *J Biol Chem* **273**:32697–32707.

25. Koch RO, Wanner SG, Koschak A, Hanner M, Schwarzer C, Kaczorowski GJ, Slaughter RS, Garcia ML, and Knaus HG (1997) Complex subunit assembly of neuronal voltage-gated K⁺ channels: basis for high-affinity toxin interactions and pharmacology. *J Biol Chem* **272**:27577–27581.

26. Shamotienko O, Akhtar S, Sidera C, Meunier FA, Ink B, Weir M, and Dolly JO (1999) Recreation of neuronal Kv1 channel oligomers by expression in mammalian cells using Semliki Forest virus. *Biochemistry* **38**:16766–16776.

27. Racape J, Lecoq A, Romi-Lebrun R, Liu J, Kohler M, Garcia ML, Menez A, and Gasparini S (2002) Characterization of a novel radio labeled peptide selective for a subpopulation of voltage-gated potassium channels in mammalian brain. *J Biol Chem* **277**:3886–3893.

28. Beckh S and Pongs O (1990) Members of the RCK potassium channel family are differentially expressed in the rat nervous system. *EMBO J* **9**:777–782.

29. Tsaur ML, Sheng M, Lowenstein DH, Jan YN, and Jan LY (1992) Differential expression of K⁺ channel mRNAs in the rat brain and down-regulation in the hippocampus following seizures. *Neuron* **8**:1055–1067.

30. Roberds SL and Tamkun MM (1991) Cloning and tissue-specific expression of five voltage-gated potassium channel cDNAs expressed in rat heart. *Proc Natl Acad Sci USA* **88**:1798–1802.

31. Klumpp DJ, Farber DB, Bowes C, Song EJ, and Pinto LH (1991) The potassium channel MBK1 (Kv1.1) is expressed in the mouse retina. *Cell Mol Neurobiol* **11**:611–622.

32. Matsubara H, Liman ER, Hess P, and Koren G (1991) Pretranslational mechanisms determine the type of potassium channels expressed in the rat skeletal and cardiac muscles. *J Biol Chem* **266**:13324–13328.

33. Adelman JP, Bond CT, Pessia M, and Maylie J (1995) Episodic ataxia results from voltage-dependent potassium channels with altered functions. *Neuron* **15**:1449–1454.

34. Zerr P, Adelman JP, and Maylie J (1998) Episodic n ataxia mutations in Kv1.1 alter potassium channel function by dominant negative effects or haplo insufficiency. *J Neurosci* **18**:2842–2848.

35. Zerr P, Adelman JP, and Maylie J (1998) Characterization of three episodic ataxia mutations in the human Kv1.1 potassium channel. *FEBS Lett* **431**:461–464.

TABLE 3
K_V1.2 channels

Channel name	K _V 1.2
Description	Voltage-gated potassium channel, delayed rectifier
Other names	HuK (IV), MK2, BK2, RCK5, RAK, BGK5, XSha2, NGK1, HBK5 ^{1–8}
Molecular information	Human: 499aa, NM_004974, chr. 1p13, KCNA2, GeneID: 3737, PMID: 2251283 ³³ Mouse: 499aa, NM_008417, chr. 3 Rat: 499aa, NM_012970 chr. 2q34
Associated subunits	K _V β ₁ , K _V β ₂ , PSD95, synapse-associated protein 97 (SAP97), SNAP25, Caspr2, RhoA ^{9–17}
Functional assays	Voltage-clamp
Current	Delayed rectifier
Conductance	14–18pS ¹⁸
Ion selectivity	K ⁺ -selective
Activation	Voltage-dependent, V _a between 5 and 27 mV; k _a = 13 mV; τ _n = 6 ms (60 mV) ^{6,18}
Inactivation	V _h between –33 and –15 mV; k _h ~ 8 mV ^{6,18}
Activators	None
Gating inhibitors	None
Blockers	4-Aminopyridine (590 μM), capsaicin (45 μM), resiniferatoxin (31 μM), flecainide (217 μM), nifedipine (18 μM), diltiazem (187 μM), 10-N-methylcarbamoyl-3,7-bis(dimethylamino)phenothiazine (0.44 μM), DTX (17 nM), charybdotoxin (14 nM), margatoxin, natrexone (2 nM), tetraethyammonium (560 mM), H37 (18 μM), picrotoxin-Kα (32 pM), OsK2 (97 nM), BgK (25 nM), HgTx (pM), anandamide (2.7 μM) ^{18–23}
Radioligands	¹²⁵ I-DTX, ¹²⁵ I-HgTX1-A19Y/Y37F ²²
Channel distribution	Brain (pons, medulla, cerebellum, inferior colliculus > hippocampus, thalamus, cerebral cortex, superior colliculus > midbrain, corpus striatum, olfactory bulb; neurons associated with mechanoreception and proprioception), spinal cord, Schwann cells, atrium, ventricle, islet, retina, smooth muscle, PC12 cells ^{1–8,24–30}
Physiological functions	Maintaining membrane potential, modulating electrical excitability in neurons and muscle
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	Delayed rectifier potassium channel; can coassemble with other K _V 1 family members in heteromultimers but not with members of other K _V families ^{19,22,25,29,31} ; intronless coding region ⁵ ; T1 domain in N terminus required for multimerization ³² ; mammalian <i>Shaker</i> -related family

aa, amino acids; chr., chromosome; DTX, dendrotoxin; HgTX, hongotoxin.

1. McKinnon D (1989) Isolation of a cDNA clone coding for a putative second potassium channel indicates the existence of a gene family. *J Biol Chem* **264**:8230–8236.

2. Kamb A, Weir M, Rudy B, Varmus H, and Kenyon C (1989) Identification of genes from pattern formation, tyrosine kinase, and potassium channel families by DNA amplification. *Proc Natl Acad Sci USA* **86**:4372–4376.

3. Stuhmer W, Ruppersberg JP, Schroter KH, Sakmann B, Stocker M, Giese KP, Perschke A, Baumann A, and Pongs O (1989) Molecular basis of functional diversity of voltage-gated potassium channels in mammalian brain. *EMBO J* **8**:3235–3244.

4. Yokoyama S, Imoto K, Kawamura T, Higashida H, Iwabe N, Miyata T, and Numa S (1989) Potassium channels from NG108–15 neuroblastoma-glioma hybrid cells: primary structure and functional expression from cDNAs. *FEBS Lett* **259**:37–42.

5. Chandy KG, Williams CB, Spencer RH, Aguilar BA, Ghanshani S, Tempel BL, and Gutman GA (1990) A family of three mouse potassium channel genes with intronless coding regions. *Science (Wash DC)* **247**:973–975.

6. Ramashwami M, Gautam M, Kamb AA, Rudy B, Tanouye MA, and Mathew MK (1990) Human potassium channel genes: molecular cloning and functional expression *Mol Cell Neurosci* **1**:214–223.

7. Roberds SL and Tamkun MM (1991) Cloning and tissue-specific expression of five voltage-gated potassium channel cDNAs expressed in rat heart. *Proc Natl Acad Sci USA* **88**:1798–1802.

8. Paulmichl M, Nasmith P, Hellmiss R, Reed K, Boyle WA, Nerbonne JM, Peralta EG, and Clapham DE (1991) Cloning and expression of a rat cardiac delayed rectifier potassium channel. *Proc Natl Acad Sci USA* **88**:7892–7895.

9. Grissmer S, Dethlefs B, Wasmuth JJ, Goldin AL, Gutman GA, Cahalan MD, and Chandy KG (1990) Expression and chromosomal localization of a lymphocyte K⁺ channel gene. *Proc Natl Acad Sci USA* **87**:9411–9415.

10. Scott VE, Rettig J, Parcej DN, Keen JN, Findlay JB, Pongs O, and Dolly JO (1994) Primary structure of a β subunit of α-dendrotoxin-sensitive K⁺ channels from bovine brain. *Proc Natl Acad Sci USA* **91**:1637–1641.

11. Rettig J, Heinemann SH, Wunder F, Lorra C, Parcej DN, Dolly JO, and Pongs O (1994) Inactivation properties of voltage-gated K⁺ channels altered by presence of β-subunit. *Nature (Lond)* **369**:289–294.

12. Kim E, Niethammer M, Rothschild A, Jan YN, and Sheng M (1995) Clustering of Shaker-type K⁺ channels by interaction with a family of membrane-associated guanylate kinases. *Nature (Lond)* **378**:85–88.

13. Rhodes KJ, Keilbaugh SA, Barrezueta NX, Lopez KL, and Trimmer JS (1995) Association and colocalization of K⁺ channel α- and β-subunit polypeptides in rat brain. *J Neurosci* **15**:5360–5371.

14. Shi G, Nakahira K, Hammond S, Rhodes KJ, Schechter LE, and Trimmer JS (1996) Beta subunits promote K⁺ channel surface expression through effects early in biosynthesis. *Neuron* **16**:843–852.

15. Rhodes KJ, Strassle BW, Monaghan MM, Bekele-Arcuri Z, Matos MF, and Trimmer JS (1997) Association and colocalization of the Kvβ1 and Kvβ2 β-subunits with Kv1 α-subunits in mammalian brain K⁺ channel complexes. *J Neurosci* **17**:8246–8258.

16. Cachero TG, Morielli AD, and Peralta EG (1998) The small GTP-binding protein RhoA regulates a delayed rectifier potassium channel. *Cell* **93**:1077–1085.

17. Poliak S, Gollan L, Martinez R, Custer A, Einheber S, Salzer JL, Trimmer JS, Shrager P, and Peles E (1999) Caspr2, a new member of the neurexin superfamily, is localized at the juxtaparanodes of myelinated axons and associates with K⁺ channels. *Neuron* **24**:1037–1047.
18. Tiffany AM, Manganas LN, Kim E, Hsueh YP, Sheng M, and Trimmer JS (2000) PSD-95 and SAP97 exhibit distinct mechanisms for regulating K⁺ channel surface expression and clustering. *J Cell Biol* **148**:147–158.
19. Grissmer S, Nguyen AN, Aiyar J, Hanson DC, Mather RJ, Gutman GA, Karmilowicz MJ, Auperin DD, and Chandy KG (1994) Pharmacological characterization of five cloned voltage-gated K⁺ channels, types Kv1.1, 1.2, 1.3, 1.5, and 3.1, stably expressed in mammalian cell lines. *Mol Pharmacol* **45**:1227–1234.
20. Koch RO, Wanner SG, Koschak A, Hanner M, Schwarzer C, Kaczorowski GJ, Slaughter RS, Garcia ML, and Knaus HG (1997) Complex subunit assembly of neuronal voltage-gated K⁺ channels: basis for high-affinity toxin interactions and pharmacology. *J Biol Chem* **272**:27577–27581.
21. Wulff H, Rauer H, Doring T, Hanselmann C, Ruff K, Wrisch A, Grissmer S, and Haensel W (1998) Alkoxypsoralens, novel nonpeptide blockers of Shaker-type K⁺ channels: synthesis and photoreactivity. *J Med Chem* **41**:4542–4529.
22. Rogowski RS, Collins JH, O'Neill TJ, Gustafson TA, Werkman TR, Rogawski MA, Tenenholz TC, Weber DJ, and Blaustein MP (1996) Three new toxins from the scorpion *Pandinus imperator* selectively block certain voltage-gated K⁺ channels. *Mol Pharmacol* **50**:1167–1177.
23. Koschak A, Bugianesi RM, Mitterdorfer J, Kaczorowski GJ, Garcia ML, and Knaus HG (1998) Subunit composition of brain voltage-gated potassium channels determined by hongotoxin-1, a novel peptide derived from *Centruroides limbatus* venom. *J Biol Chem* **273**:2639–2644.
24. Poling JS, Rogawski MA, Salem NJ, and Vicini S (1996) Anandamide, an endogenous cannabinoid, inhibits Shaker-related voltage-gated K⁺ channels. *Neuropharmacology* **35**:983–991.
25. UniGene Cluster Hs0.248139; Online Mendelian Inheritance in Man (OMIM) no. 176262.
26. Sheng M, Liao YJ, Jan YN, and Jan LY (1993) Presynaptic A-current based on heteromultimeric K⁺ channels detected in vivo. *Nature (Lond)* **365**:72–75.
27. Sheng M, Tsaur ML, Jan YN, and Jan LY (1994) Contrasting subcellular localization of the Kv1.2 K⁺ channel subunit indifferent neurons of rat brain. *J Neurosci* **14**:2408–2417.
28. Wang H, Kunkel DD, Schwartzkroin PA, and Tempel BL (1994) Localization of Kv1.1 and Kv1.2, two K channel proteins, to synaptic terminals, somata, and dendrites in the mouse brain. *J Neurosci* **14**:4588–4599.
29. Klumpp DJ, Song EJ, Ito S, Sheng MH, Jan LY, and Pinto LH (1995) The Shaker-like potassium channels of the mouse rod bipolar cell and their contributions to the membrane current. *J Neurosci* **15**:5004–5013.
30. Sobko A, Peretz A, Shirihai O, Etkin S, Cherepanova V, Dagan D, and Attali B (1998) Heteromultimeric delayed-rectifier K⁺ channels in Schwann cells: developmental expression and role in cell proliferation. *J Neurosci* **18**:10398–10408.
31. Coleman SK, Newcombe J, Pryke J, and Dolly JO (1999) Subunit composition of Kv1 channels in human CNS. *J Neurochem* **73**:849–858.
32. Rasband MN, Park EW, Vanderah TW, Lai J, Porreca F, and Trimmer JS (2001) Distinct potassium channels on pain-sensing neurons. *Proc Natl Acad Sci USA* **98**:13373–13378.
33. Minor DL, Lin YF, Mobley BC, Avelar A, Jan YN, Jan LY, and Berger JM (2000) The polar T1 interface is linked to conformational changes that open the voltage-gated potassium channel. *Cell* **102**:657–670.

TABLE 4
K_v1.3 channels

Channel name	K _v 1.3 ^{1–8}
Description	Voltage-gated potassium channel, delayed rectifier
Other names	MK3, MBK3, RCK3, hPCN3, HuK (III), HLK3, RKG5, KV3, HGK5, <i>n</i> -channel
Molecular information	Human: 523aa, NM_002232, chr. 1p13.3, ^{7,9} KCNA3, GeneID: 3738, PMID: 22512834 Mouse: 528aa, NM_008418, chr. 3 Rat: 525aa, NM_019270, chr. 2q34 K _v β, hDlg, β ₁ integrin, KChAP ^{10–12}
Associated subunits	
Functional assays	Voltage-clamp
Current	Type N voltage-gated potassium channel in lymphocytes ^{3,4}
Conductance	13pS ⁴
Ion selectivity	K ⁺ (1) > Rb ⁺ (0.77) > NH ₄ ⁺ (0.1) > Cs ⁺ (0.02) > Na ⁺ (<0.01) ¹³
Activation	Voltage, V _a = –35 mV; k _a = 6 mV; τ _n = 3 ms at 40 mV ^{4,13}
Inactivation	C-type inactivation, V _h = –63 mV; k _h = 7.7 mV; τ _h = 250 ms (40 mV) ⁴
Activators	None
Gating inhibitors	None
Blockers	4-Aminopyridine (195 μM), tetraethylammonium (10 mM), charybdotoxin (3 nM), naltrexone (1 nM), MgTX (110 pM), kaliotoxin (650 pM), AgTX2 (200 pM), Pi1 (11 nM), Pi2 (50 pM), Pi3 (500 pM), HsTx1 (12 pM), ShK (11 pM), BgK (39 nM), ShK-Dap22 (52 pM), quinine (14 μM), diltiazem (60 μM), verapamil (6 μM), CP339818 (150 nM), UK78282 (200 nM), correolide (90 nM), sulfamid-benzamidoindane (100 nM), capsaicin (26 μM), resiniferatoxin (3 μM), nifedipine (5 μM), H37 (23 μM) ^{14,15}
Radioligands	¹²⁵ I-HgTx1-A19Y/Y37F mutant (0.1–0.25 pM); ¹²⁵ I-MgTx (0.3 pM) ^{16,17}
Channel distribution	Brain (inferior colliculus > olfactory bulb, pons/medulla > midbrain, superior colliculus, corpus striatum, hippocampus, cerebral cortex), lung, islets, thymus, spleen, lymph node, fibroblasts, B lymphocytes, T lymphocytes, pre-B cells, tonsils, macrophages, microglia, oligodendrocytes, osteoclasts, platelets, testis ^{1–8,18–21}
Physiological functions	Regulation of membrane potential and calcium signaling in lymphocytes and oligodendrocytes ^{14,21–23}
Mutations and pathophysiology	Not established
Pharmacological significance	Therapeutic target for immunosuppressants; K _v 1.3 inhibitors suppress T-cell activation in vitro and delayed type hypersensitivity in vivo and have proven effective for multiple sclerosis in an animal ^{24,25} ; K _v 1.3 expression is dramatically and exclusively increased in effector memory T cells
Comments	Can coassemble with other K _v 1 family members in heteromultimers but not with members of other K _v families; intronless coding region; mammalian <i>Shaker</i> -related family

aa, amino acids; chr., chromosome; MgTX, margatoxin; HgTX, hongotoxin; CP339818, *N*-[1-(phenylmethyl)-4(1*H*)-quinolinylidene]-1-pentamine monohydrochloride; UK78282, 4-[(diphenylmethoxy)methyl]-1-[3-(4-methoxyphenyl)propyl]-piperidine.

1. Stuhmer W, Ruppersberg JP, Schroter KH, Sakmann B, Stocker M, Giese KP, Perschke A, Baumann A, and Pongs O (1989) Molecular basis of functional diversity of voltage-gated potassium channels in mammalian brain. *EMBO J* **8**:3235–3244.

2. Chandy KG, Williams CB, Spencer RH, Aguilar BA, Ghanshani S, Tempel BL, and Gutman GA (1990) A family of three mouse potassium channel genes with intronless coding regions. *Science (Wash DC)* **247**:973–975.

3. Douglass J, Osborne PB, Cai YC, Wilkinson M, Christie MJ, and Adelman JP (1990) Characterization and functional expression of a rat genomic DNA clone encoding a lymphocyte potassium channel. *J Immunol* **144**:4841–4850.

4. Grissmer S, Dethlefs B, Wasmuth JJ, Goldin AL, Gutman GA, Cahalan MD, and Chandy KG (1990) Expression and chromosomal localization of a lymphocyte K⁺ channel gene. *Proc Natl Acad Sci USA* **87**:9411–9415.

5. Philipson LH, Hice RE, Schaefer K, LaMendola J, Bell GI, Nelson DJ, and Steiner, DF (1991) Sequence and functional expression in *Xenopus* oocytes of a human insulinoma and islet potassium channel. *Proc Natl Acad Sci USA* **88**:53–57.

6. Swanson R, Marshall J, Smith JS, Williams JB, Boyle MB, Folander K, Luneau CJ, Antanavage J, Oliva C, Buhrow SA, et al. (1990) Cloning and expression of cDNA, and genomic clones encoding three delayed rectifier potassium channels in rat brain. *Neuron* **4**:929–939.

7. Attali B, Romey G, Honore E, Schmid-Alliana A, Mattei MG, Lesage F, Ricard P, Barhanin J, and Lazdunski M (1992) Cloning, functional expression, and regulation of two K⁺ channels in human T lymphocytes. *J Biol Chem* **267**:8650–8657.

8. Cai YC, Osborne PB, North RA, Dooley DC, and Douglass J (1992) Characterization and functional expression of genomic DNA encoding the human lymphocyte type n potassium channel. *DNA Cell Biol* **11**:163–172.

9. Folander K, Douglass J, and Swanson R (1994) Confirmation of the assignment of the gene encoding Kv1.3, a voltage-gated potassium channel (KCNA3) to the proximal short arm of human chromosome 1. *Genomics* **23**:295–296.

10. McCormack T, McCormack K, Nadal MS, Vieira E, Ozaita A, and Rudy B (1999) The effects of Shaker β -subunits on the human lymphocyte K⁺ channel Kv1.3. *J Biol Chem* **274**:20123–21126.

11. Hanada T, Lin L Chandy KG, Oh SS, and Chishti AH (1997) Human homologue of the *Drosophila* discs large tumor suppressor binds to p56lck tyrosine kinase and Shaker type Kv1.3 potassium channel in T lymphocytes. *J Biol Chem* **272**:26899–26904.

12. Levite M, Cahalon L, Peretz A, Hershkovitz R, Sobko A, Ariel A, Desai R, Attali B, and Lider O (2000) Extracellular K⁺ and opening of voltage-gated potassium channels activate T cell integrin function: physical and functional association between Kv1.3 channels and β 1 integrins. *J Exp Med* **191**:1167–1176.

13. Cahalan MD, Chandy KG, DeCoursey TE, and Gupta S (1985) A voltage-gated potassium channel in human T lymphocytes. *J Physiol* **358**:197–237.

14. Grissmer S, Nguyen AN, Aiyar J, Hanson DC, Mather RJ, Gutman GA, Karmilowicz MJ, Auperin DD, and Chandy KG (1994) Pharmacological characterization of five cloned voltage-gated K⁺ channels, types Kv1.1, 1.2, 1.3, 1.5, and 3.1, stably expressed in mammalian cell lines. *Mol Pharmacol* **45**:1227–1234.

15. Chandy KG, Cahalan MD, Pennington M, Norton RS, Wulff H, and Gutman GA (2001) Potassium channels in T lymphocytes: toxins to therapeutic immunosuppressants. *Toxicon* **39**:1269–1276.

16. Koschak A, Bugianesi RM, Mitterdorfer J, Kaczorowski GJ, Garcia ML, and Knaus HG (1998) Subunit composition of brain voltage-gated potassium channels determined by hongoxin-1, a novel peptide derived from *Centruroides limbatus* venom. *J Biol Chem* **273**:2639–2644.

17. Koch RO, Wanner SG, Koschak A, Hanner M, Schwarzer C, Kaczorowski GJ, Slaughter RS, Garcia ML, and Knaus HG (1997) Complex subunit assembly of neuronal voltage-gated K⁺ channels: basis for high-affinity toxin interactions and pharmacology. *J Biol Chem* **272**:27577–27581.

18. UniGene Cluster Hs0.169948 *Homo sapiens*.

19. Arkett SA, Dixon J, Yang JN, Sakai DD, Minkin C, and Sims SM (1994) Mammalian osteoclasts express a transient potassium channel with properties of Kv1.3. *Receptors Channels* **2**:281–293.

20. DeCoursey TE, Kim SY, Silver MR, and Quandt FN (1996) Ion channel expression in PMA-differentiated human THP-1 macrophages. *J Membr Biol* **152**:141–157.

21. Chittajallu R, Chen Y, Wang H, Yuan X, Ghiani CA, Heckman T, McBain CJ, and Gallo V (2002) Regulation of Kv1 subunit expression in oligodendrocyte progenitor cells and their role in G1/S phase progression of the cell cycle. *Proc Natl Acad Sci USA* **99**:2350–2355.

22. Cahalan MD and Chandy KG (1997) Ion channels in the immune system as targets for immuno suppression. *Curr Opin Biotechnol* **8**:749–756.

23. Cahalan MD, Wulff H, and Chandy KG (2001) Molecular properties and physiological roles of ion channels in the immune system. *J Clin Immunol* **21**:235–252.

24. Koo GC, Blake JT, Talento A, Nguyen M, Lin S, Sirotina A, Shah K, Mulvany K, Hora D Jr, Cunningham P, et al (1997) Blockade of the voltage-gated potassium channel Kv1.3 inhibits immune responses in vivo. *J Immunol* **158**:5120–5128.

25. Beeton C, Wulff H, Barbaria J, Clot-Faybesse O, Pennington M, Bernard D, Cahalan MD, Chandy KG, and Beraud E (2001) Selective blockade of T lymphocyte K⁺ channels ameliorates experimental autoimmune encephalomyelitis, a model for multiple sclerosis. *Proc Natl Acad Sci USA* **98**:13942–13947.

TABLE 5
K_V1.4 channels

Channel name	K _V 1.4 ^{1–7}
Description	Voltage-gated potassium channel, A-type, fast-inactivating
Other names	HuK (II), hPCN2, HK1, RCK4, RHK1, RK4, RK8, MK4
Molecular information	Human: 653aa, NM_002233, chr. 11p14.3–15.2, ⁷ KCNA4, GeneID: 3739, PMID: 2263489 ³² Mouse: 654aa, NM_021275, chr. 2 Rat: 654aa, NM_012971, chr. 3q33
Associated subunits	K _V β , PSD95, synapse-associated protein 97 (SAP97), SAP90, α -actinin-2, KChAP, σ receptor ^{8–18}
Functional assays	Voltage-clamp
Current	K _V 1.4/K _V 1.2 heteromultimers may underlie the presynaptic A-type K ⁺ channel ¹⁹
Conductance	5pS ¹
Ion selectivity	K ⁺ -selective (50 times more selective for K ⁺ than Na ⁺) ²⁰
Activation	Voltage, V _a = –22 mV ¹ ; –34 mV ²⁰ ; K _a = 5 ²¹
Inactivation	N-type inactivation, V _h = –62 mV ²⁰ ; τ_h = 47 ms (0 mV) ²⁰
Activators	CaMKII/calcineurin regulation through phosphorylation/dephosphorylation makes inactivation Ca ²⁺ -dependent ²²
Gating inhibitors	None
Blockers	4-Aminopyridine (13 μ M), ¹ tetraethylammonium (>100 mM), ³ UK78282 (170 nM), ²³ riluzole (70 μ M), ²⁴ quinidine (10 μ M–1 mM), ²⁵ nifedipine (0.8 μ M) ²⁶
Radioligands	None
Channel distribution	Brain (olfactory bulb, corpus striatum > hippocampus, superior and inferior colliculus > cerebral cortex, midbrain basal ganglia > pons/medulla), lung-carcinoid, skeletal muscle, heart, pancreatic islet ^{1,6,27–29}
Physiological functions	Neuronal afterhyperpolarization
Mutations and pathophysiology	K _V 1.4 expression increases in rat ventricular myocytes after myocardial infarction and induction of diabetes ^{30,31}
Pharmacological significance	Not established
Comments	Can coassemble with other K _v 1 family members in heteromultimers but not with members of other K _v families; intronless coding region; mouse K _v 1.4 mRNA contains an internal ribosome entry site in its 5′-noncoding region and may be translated by cap-independent mechanisms ^{33,34} , mammalian <i>Shaker</i> -related family

aa, amino acids; chr., chromosome.

1. Stuhmer W, Ruppersberg JP, Schroter KH, Sakmann B, Stocker M, Giese KP, Perschke A, Baumann A, and Pongs O (1989) Molecular basis of functional diversity of voltage-gated potassium channels in mammalian brain. *EMBO J* **8**:3235–3244.

2. Kamb A, Weir M, Rudy B, Varmus H, and Kenyon C (1989) Identification of genes from pattern formation, tyrosine kinase, and potassium channel families by DNA amplification. *Proc Natl Acad Sci USA* **86**:4372–4376.

3. Philipson LH, Hice RE, Schaefer K, LaMendola J, Bell GI, Nelson DJ, and Steiner DF (1991) Sequence and functional expression in *Xenopus* oocytes of a human insulinoma and islet potassium channel. *Proc Natl Acad Sci USA* **88**:53–57.

4. Tseng-Crank JC, Tseng GN, Schwartz A, and Tanouye MA (1990) Molecular cloning and functional expression of a potassium channel cDNA isolated from a rat cardiac library. *FEBS Lett* **268**:63–68.
5. Roberds SL and Tamkun MM (1991) Cloning and tissue-specific expression of five voltage-gated potassium channel cDNAs expressed in rat heart. *Proc Natl Acad Sci USA* **88**:1798–1802.
6. Tamkun MM, Knoth KM, Walbridge JA, Kroemer H, Roden DM, and Glover DM (1991) Molecular cloning and characterization of two voltage-gated K⁺ channel cDNAs from human ventricle. *FASEB J* **5**:331–337.
7. Wymore RS, Korenberg JR, Kinoshita KD, Aiyar J, Coyne C, Chen XN, Hustad CM, Copeland NG, Gutman GA, Jenkins NA, et al. (1994) Genomic organization, nucleotide sequence, biophysical properties, and localization of the voltage-gated K⁺ channel gene KCNA4/Kv1.4 to mouse chromosome 2/human 11p14 and mapping of KCNC1/Kv3.1 to mouse 7/human11p14.3-p15.2 and KCNA1/Kv1.1 to human 12p13. *Genomics* **20**:191–202.
8. Morales MJ, Castellino RC, Crews AL, Rasmusson RL, and Strauss HC (1995) A novel β subunit increases rate of inactivation of specific voltage-gated potassium channel α subunits. *J Biol Chem* **270**:6272–6277.
9. Rhodes KJ, Keilbaugh SA, Barrezueta NX, Lopez KL, and Trimmer JS (1995) Association and colocalization of K⁺ channel α - and β -subunit polypeptides in rat brain. *J Neurosci* **15**:5360–5371.
10. McCormack K, McCormack T, Tanouye M, Rudy B, and Stuhmer W (1995) Alternative splicing of the human Shaker K⁺ channel β 1 gene and functional expression of the β 2 gene product. *FEBS Lett* **370**:32–36.
11. Heinemann SH, Rettig J, Graack HR, and Pongs O (1996) Functional characterization of Kv channel β -subunits from rat brain. *J Physiol* **493**:625–633.
12. Rhodes KJ, Strassle BW, Monaghan MM, Bekele-Arcuri Z, Matos MF, and Trimmer JS (1997) Association and colocalization of the Kv β 1 and Kv β 2 β -subunits with Kv1 α -subunits in mammalian brain K⁺ channel complexes. *J Neurosci* **17**:8246–8258.
13. Kim E and Sheng M (1996) Differential K⁺ channel clustering activity of PSD-95 and SAP97, two related membrane-associated putative guanylate kinases. *Neuropharmacology* **35**:993–1000.
14. Topinka JR and Brecht DS (1998) N-terminal palmitoylation of PSD-95 regulates association with cell membranes and interaction with K⁺ channel Kv1.4. *Neuron* **20**:125–134.
15. Cukovic D, Lu GW, Wible B, Steele DF, and Fedida D (2001) A discrete amino terminal domain of Kv1.5 and Kv1.4 potassium channels interacts with the spectrin repeats of α -actinin-2. *FEBS Lett* **498**:87–92.
16. Kuryshv YA, Wible BA, Gudiz TI, Ramirez AN, and Brown AM (2001) KChAP/Kv β 1.2 interactions and their effects on cardiac Kv channel expression. *Am J Physiol Cell Physiol* **281**:C290–C299.
17. Piserchio A, Pellegrini M, Mehta S, Blackman SM, Garcia EP, Marshall J, and Mierke DF (2002) The PDZ1 domain of SAP90: characterization of structure and binding. *J Biol Chem* **277**:6967–6973.
18. Aydar E, Palmer CP, Klyachko VA, and Jackson MB (2002) The sigma receptor as a ligand-regulated auxiliary potassium channel subunit. *Neuron* **34**:399–410.
19. Sheng M, Liao YJ, Jan YN, and Jan LY (1993) Presynaptic A-current based on heteromultimeric K⁺ channels detected in vivo. *Nature (Lond)* **365**:72–75.
20. Po S, Snyder DJ, Baker R, Tamkun MM, and Bennett PB (1992) Functional expression of an inactivating potassium channel cloned from human heart. *Circ Res* **71**:732–736.
21. Elliott A and Elliott JR (1997) Channel-specific effects of n-alkyl sulphate anions on three Shaker-related potassium channels expressed in *Xenopus* oocytes. *Pflug Arch Eur J Physiol* **434**:132–136.
22. Roeper J, Lorra C, and Pongs O (1997) Frequency-dependent inactivation of mammalian A-type K⁺ channel KV1.4 regulated by Ca²⁺/calmodulin-dependent protein kinase. *J Neurosci* **17**:3379–3391.
23. Nguyen A, Kath JC, Hanson DC, Biggers MS, Canniff PC, Donovan CB, Mather RJ, Bruns MJ, Rauer H, Aiyar J, et al (1996) Novel non peptide agents potently block the C-type inactivated conformation of Kv1.3 and suppress T cell activation. *Mol Pharmacol* **50**:1672–1679.
24. Xu L, Enyeart JA, and Enyeart JJ (2001) Neuro protective agent riluzole dramatically slows in-activation of Kv1.4 potassium channels by a voltage-dependent oxidative mechanism. *J Pharmacol Exp Ther* **299**:227–237.
25. Yamagishi T, Ishii K, and Taira N (1995) Antiarrhythmic and bradycardic drugs inhibit currents of cloned K⁺ channels, Kv1.2 and Kv1.4. *Eur J Pharmacol* **281**:151–159.
26. Hatano N, Ohya S, Muraki K, Giles W, and Imaizumi Y (2003) Dihydropyridine Ca²⁺ channel antagonists and agonists block Kv4.2, Kv4.3 and Kv1.4 K⁺ channels expressed in HEK293 cells. *Br J Pharmacol* **139**:533–544.
27. Beckh S and Pongs O (1990) Members of the RCK potassium channel family are differentially expressed in the rat nervous system. *EMBO J* **9**:777–782.
28. Sheng M, Tsaur ML, Jan YN, and Jan LY (1992) Subcellular segregation of two A-type K⁺ channel proteins in rat central neurons. *Neuron* **9**:271–284.
29. Blair TA, Roberds SL, Tamkun MM, and Hartshorne RP (1991) Functional characterization of RK5, a voltage-gated K⁺ channel cloned from the rat cardiovascular system. *FEBS Lett* **295**:211–213.
30. Kaprielian R, Wickenden AD, Kassiri Z, Parker TG, Liu PP, and Backx, PH (1999) Relationship between K⁺ channel down-regulation and [Ca²⁺]_i in rat ventricular myocytes following myocardial infarction. *J Physiol (Lond)* **517**:229–245.
31. Nishiyama A, Ishii DN, Backx PH, Pulford BE, Birks BR, and Tamkun MM (2001) Altered K⁺ channel gene expression in diabetic rat ventricle: isoform switching between Kv4.2 and Kv1.4. *Am J Physiol* **281**:H1800–H1807.
32. Philipson LH, Schaefer K, LaMendola J, Bell GI, and Steiner DF (1990) Sequence of a human fetal skeletal muscle potassium channel cDNA related to RCK4. *Nucleic Acids Res* **18**:7160.
33. Negulescu D, Long LE, Chandy KG, Semler B, and Gutman GA (1998) Translation initiation of a cardiac voltage-gated potassium channel by internal ribosome entry. *J Biol Chem* **273**:20109–20113.
34. Jang GM, Leong LE, Hoang LT, Wang PH, Gutman GA, Semler BL (2004) Structurally distinct elements mediate internal ribosome entry within the 5' noncoding region of a voltage-gated potassium channel mRNA. *J Biol Chem* **279**:47419–47430.

TABLE 6
K_v1.5 channels

Channel name	K _v 1.5
Description	Voltage-gated potassium channel, delayed rectifier
Other names	HpCN1, HK2, HCK1, KV1, fHK, RK3, RMK2, HuK (II) ^{1–8}
Molecular information	Human: 613aa, NM_002234, chr. 12p13.3, ^{8–10} <i>KCNA5</i> , GeneID: 3741, PMID: 1986382 ³ Mouse: 602aa, NM_002234, chr. 6 Rat: 602aa, NM_012972, chr. 4q42–44
Associated subunits	K _v β ₁ , K _v β ₂ , KCNA3B, Src tyrosine kinase, fyn, KChAP, α-actinin-2, caveolin, synapse-associated protein 97 (SAP97) ^{11–21}
Functional assays	Voltage-clamp
Current	Ultrarapid-activating K ⁺ current in heart (IK _{ur}) ^{22,23}
Conductance	8pS ²⁴
Ion selectivity	K ⁺
Activation	Voltage, V _a = −14 mV; k _a = 6–12 mV ^{22,24}
Inactivation	V _h = −25 to −10 mV; k _h = 3–5 mV; τ _{h1} = 460 ms; τ _{h2} = 5 s (40 mV) ^{22,24}
Activators	None
Gating inhibitors	None
Blockers	S9947 (420 nM), 4-aminopyridine (270 μM),capsaicin (23 μM), resiniferatoxin (26 μM), flecainide (101 μM), nifedipine (81 μM), diltiazem (115 μM), tetraethylammonium (330 mM), clofilium inside (140 nM), bupivacaine (4.1 μM), propafenone (4.4 μM), ^{24–26} quinidine (0.6 μM) ²⁷
Radioligands	None
Channel distribution	Aorta, colon, kidney, pooled colon, kidney, stomach, smooth muscle, whole embryo, hippocampus and cortex (oligodendrocytes, microglia, Schwann cells), pituitary, pulmonary artery ^{1–7,28–33}
Physiological functions	K _v 1.5 has properties similar to the ultrarapidly activating IK _{ur} current in the heart, and antisense-targeting K _v 1.5 suppresses IK _{ur} currents almost 50% ^{22,23} ; maintains membrane potential that modulates electrical excitability in neurons
Mutations and pathophysiology	Not established
Pharmacological significance	Potential use in management of atrial fibrillation via blockade of IK _{ur} ^{34,35}
Comments	Can coassemble with other K _v 1 family members in heteromultimers but not with members of other K _v families; intronless coding region; mammalian <i>Shaker</i> -related family.

aa, amino acids; chr., chromosome.

1. Swanson R, Marshall J, Smith JS, Williams JB, Boyle MB, Folander K, Luneau CJ, Antanavage J, Oliva C, Buhrow SA, et al. (1990) Cloning and expression of cDNA, and genomic clones encoding three delayed rectifier potassium channels in rat brain. *Neuron* **4**:929–939.

2. Ramashwami M, Gautam M, Kamb AA, Rudy B, Tanouye MA, and Mathew MK (1990) Human potassium channel genes: molecular cloning and functional expression *Mol Cell Neurosci* **1**:214–223.

3. Philipson LH, Hice RE, Schaefer K, LaMendola J, Bell GI, Nelson DJ, and Steiner DF (1991) Sequence and functional expression in *Xenopus* oocytes of a human insulinoma and islet potassium channel. *Proc Natl Acad Sci USA* **88**:53–57.

4. Kamb A, Weir M, Rudy B, Varmus H, and Kenyon C (1989) Identification of genes from pattern formation, tyrosine kinase, and potassium channel families by DNA amplification. *Proc Natl Acad Sci USA* **86**:4372–4376.

5. Roberds SL and Tamkun MM (1991) Cloning and tissue-specific expression of five voltage-gated potassium channel cDNAs expressed in rat heart. *Proc Natl Acad Sci USA* **88**:1798–1802.

6. Tamkun MM, Knoth KM, Walbridge JA, Kroemer H, Roden DM, and Glover DM (1991) Molecular cloning and characterization of two voltage-gated K⁺ channel cDNAs from human ventricle. *FASEB J* **5**:331–337.

7. Matsubara H, Liman ER, Hess P, and Koren G (1991) Pretranslational mechanisms determine the type of potassium channels expressed in the rat skeletal and cardiac muscles. *J Biol Chem* **266**:13324–13328.

8. Curran ME, Landes GM, and Keating MT (1992) Molecular cloning, characterization, and genomic localization of a human potassium channel gene. *Genomics* **12**:729–737.

9. Phromchotikul T, Browne DL, Curran ME, Keating MT, and Litt M (1993) Dinucleotide repeat polymorphism at the KCNA5 locus. *Hum Mol Genet* **2**:1512.

10. Albrecht B, Weber K, and Pongs O (1995) Characterization of a voltage-activated K-channel gene cluster on human chromosome 12p13. *Receptors Channels* **3**:213–220.

11. Sewing S, Roeper J, and Pongs O (1996) Kv β 1 subunit binding specific for shaker-related potassium channel α subunits. *Neuron* **16**:455–463.

12. Uebele VN, England SK, Chaudhary A, Tamkun MM, and Snyders DJ (1996) Functional differences in Kv1.5 currents expressed in mammalian cell lines are due to the presence of endogenous Kv β 2.1 subunits. *J Biol Chem* **271**:2406–2412.

13. Heinemann SH, Rettig J, Graack HR, and Pongs O (1996) Functional characterization of Kv channel β-subunits from rat brain. *J Physiol* **493**:625–633.

14. Wang Z, Kiehn J, Yang Q, Brown AM, and Wible BA (1996) Comparison of binding and block produced by alternatively spliced Kvβ1 subunits. *J Biol Chem* **271**:28311–28317.

15. Holmes TC, Fadool DA, Ren R, and Levitan IB (1996) Association of Src tyrosine kinase with a human potassium channel mediated by SH3 domain. *Science (Wash DC)* **274**:2089–2091.

16. Sobko A, Peretz A, and Attali B (1998) Constitutive activation of delayed-rectifier potassium channels by a src family tyrosine kinase in Schwann cells. *EMBO J* **17**:4723–4734.

17. Wible BA, Yang Q, Kuryshv YA, Accili EA, and Brown AM (1998) Cloning and expression of a novel K⁺ channel regulatory protein, KchAP. *J Biol Chem* **273**:11745–11751.

18. Leicher T, Bahringer R, Isbrandt D, and Pongs O (1998) Co expression of the KCNA3B gene product with Kv1.5 leads to a novel A-type potassium channel. *J Biol Chem* **273**:35095–35101.

19. Maruoka ND, Steele DF, Au BP, Dan P, Zhang X, Moore ED, and Fedida D (2000) α-Actinin-2 couples to cardiac Kv1.5 channels, regulating current density and channel localization in HEK cells. *FEBS Lett* **473**:188–194.

20. Martens JR, Sakamoto N, Sullivan SA, Grobaski TD, and Tamkun MM (2001) Isoform-specific localization of voltage-gated K⁺ channels to distinct lipid raft populations: targeting of Kv1.5 to caveolae. *J Biol Chem* **276**:8409–8414.

21. Murata M, Buckett PD, Zhou J, Brunner M, Folco E, and Koren G (2001) SAP97 interacts with Kv1.5 in heterologous expression systems. *Am J Physiol Heart Circ Physiol* **281**:H2575–H2584.

22. Snyders DJ, Tamkun MM, and Bennett PB (1993) A rapidly activating and slowly inactivating potassium channel cloned from human heart. Functional analysis after stable mammalian cell culture expression. *J Gen Physiol* **101**:513–543.

23. Feng J, Wible B, Li GR, Wang Z, and Nattall S (1997) Antisense oligodeoxynucleotides directed against Kv1.5 mRNA specifically inhibit ultrarapid delayed rectifier K⁺ current in cultured adult human atrial myocytes. *Circ Res* **80**:572–579.

24. Grissmer S, Nguyen AN, Aiyar J, Hanson DC, Mather RJ, Gutman GA, Karmilowicz MJ, Auperin DD, and Chandy KG (1994) Pharmacological characterization of five cloned voltage-gated K⁺ channels, types Kv1.1, 1.2, 1.3, 1.5, and 3.1, stably expressed in mammalian cell lines. *Mol Pharmacol* **45**:1227–1234.

25. Malayev AA, Nelson DJ, and Philipson LH (1995) Mechanism of clofilium block of the human Kv1.5 delayed rectifier potassium channel. *Mol Pharmacol* **47**:198–205.

26. Franquez L, Longobardo M, Vicente J, Delpon E, Tamkun MM, Tamargo J, Snyders DJ, and Valenzuela C (1997) Molecular determinants of stereo selective bupivacaine block of hKv1.5 channels. *Circ Res* **81**:1053–1064.

27. Snyder J, Knoth KM, Roberds SL, and Tamkun M.M (1992) Time-, voltage-, and state-dependent block by quinidine of a cloned human cardiac potassium channel. *Mol Pharmacol* **41**:322–330.
28. UniGene Cluster Hs0.150208; OMIM no. 176267.
29. Kotecha SA and Schlichter LC (1999) A Kv1.5 to Kv1.3 switch in endogenous hippocampal microglia and a role in proliferation. *J Neurosci* **19**:10680–10693.
30. Sobko A, Peretz A, Shirihai O, Etkin S, Cherepanova V, Dagan D, and Attali B (1998) Heteromultimeric delayed-rectifier K⁺ channels in Schwann cells: developmental expression and role in cell proliferation. *J Neurosci* **18**:10398–10408.
31. Chittajallu R, Chen Y, Wang H, Yuan X, Ghiani CA, Heckman T, McBain CJ, and Gallo V (2002) Regulation of Kv1 subunit expression in oligodendrocyte progenitor cells and their role in G1/S phase progression of the cell cycle. *Proc Natl Acad Sci USA* **99**:2350–2355.
32. Takimoto K, Fomina AF, Gealy R, Trimmer JS, and Levitan ES (1993) Dexamethasone rapidly induces Kv1.5 K⁺ channel gene transcription and expression in clonal pituitary cells. *Neuron* **11**:359–369.
33. Wang J, Juhaszova M, Rubin LJ, and Yuan XJ (1997) Hypoxia inhibits gene expression of voltage-gated K⁺ channel α subunits in pulmonary artery smooth muscle cells. *J Clin Invest* **100**:2347–2353.
34. Van Wagoner DR, Pond AL, McCarthy PM, Trimmer JS, and Nerbonne JM (1997) Outward K⁺ current densities and Kv1.5 expression are reduced in chronic human atrial fibrillation. *Circ Res* **80**:772–781.
35. Brundel BJ, Van Gelder IC, Henning RH, Tuinenburg AE, Wietes M, Grandjean JG, Wilde AA, Van Gilst WH, and Crijns HJ (2001) Alterations in potassium channel gene expression in atria of patients with persistent and paroxysmal atrial fibrillation: differential regulation of protein and mRNA levels for K⁺ channels. *J Am Coll Cardiol* **37**:926–932.

TABLE 7
K_v1.6 channels

Channel name	K _v 1.6 ^{1–5}
Description	Voltage-gated potassium channel, delayed rectifier
Other names	HBK2, MK1.6, RCK2, KV2
Molecular information	Human: 528aa, NM_002235, chr. 12p13.3, ⁶ KCNA6, GeneID: 3742, PMID:2347305 ¹ Mouse: 529aa, NM_013568, chr. 6 Rat: 530 aa, XM_575671 (predicted), chr. 4q42
Associated subunits	K _v β ₁ , K _v β ₂ , ^{7,8} Caspr2 ¹⁸
Functional assays	Voltage-clamp
Current	Delayed rectifier
Conductance	9pS ¹
Ion selectivity	K ⁺ -selective
Activation	V _a = −20 mV; k _a = 8 mV ¹
Inactivation	K _h = −43 ² ; τ_h = > 3 s ¹
Activators	None
Gating inhibitors	None
Blockers	α -Dendrotoxin (20 nM), ¹ 10-N-methylcarbamoyl-3,7-bis(dimethylamino)phenothiazine (10 nM, ¹ 200 nM ³), 4-aminopyridine (1.5 mM), ^{1,3} tetraethylammonium (7 mM), ^{1,3} ShK (160 pM), ⁹ HgTx (9.6 pM), ¹⁰ BgK (W5Y/F6A/Y26F) ¹¹ ¹²⁵ I-BgK (W5Y/F6A/Y26F), ¹¹ ¹²⁵ I-HgTx
Radioligands	
Channel distribution	Brain, colon, germ cell, heart, lung, ovary, testis, astrocytes, pulmonary artery smooth muscle cells, oligodendrocytes ^{1,3–5,8,12–16}
Physiological functions	Regulator of membrane potential in neurons
Mutations and pathophysiology	No K ⁺ channel clustering in optic nerves of hypomyelinating Shiverer mice
Pharmacological significance	Not established
Comments	Can coassemble with other K _v 1 family members in heteromultimers but not with members of other K _v families; intronless coding region; N terminus contains an N terminus inactivation prevention (NIP) domain; ¹⁷ mammalian <i>Shaker</i> -related family

aa, amino acids; chr., chromosome; HgTx, hongotoxin.

- Grupe A, Schroter KH, Ruppersberg JP, Stocker M, Drewes T, Beckh S, and Pongs O (1990) Cloning and expression of a human voltage-gated potassium channel: a novel member of the RCK potassium channel family. *EMBO J* **9**:1749–1756.
- Guihard G, Bellocq C, Grelet E, and Escande D (2003) Human Kv1.6 current displays a C-type-like inactivation when re-expressed in cos-7 cells. *Biochem Biophys Res Commun* **311**:883–89.
- Swanson R, Marshall J, Smith JS, Williams JB, Boyle MB, Folander K, Luneau CJ, Antanavage J, Oliva C, Buhrow SA, et al. (1990) Cloning and expression of cDNA, and genomic clones encoding three delayed rectifier potassium channels in rat brain. *Neuron* **4**:929–939.
- Migeon MB, Street VA, Demas VP, and Tempel BL (1992) Cloning, sequence and chromosomal localization of MK6, a murine potassium channel gene. *Epilepsy Res Suppl* **9**:173–180.
- Ramashwami M, Gautam M, Kamb AA, Rudy B, Tanouye MA, and Mathew MK (1990) Human potassium channel genes: molecular cloning and functional expression. *Mol Cell Neurosci* **1**:214–223.
- Albrecht B, Weber K, and Pongs O (1995) Characterization of a voltage-activated K-channel gene cluster on human chromosome 12p13. *Receptors Channels* **3**:213–220.
- Rhodes KJ, Strassle BW, Monaghan MM, Bekele-Arcuri Z, Matos MF, and Trimmer JS (1997) Association and colocalization of the Kv β 1 and Kv β 2 β -subunits with Kv1 α -subunits in mammalian brain K⁺ channel complexes. *J Neurosci* **17**:8246–8258.
- Rasband MN, Trimmer JS, Peles E, Levinson SR, and Shrager P (1999) K⁺ channel distribution and clustering in developing and hypomyelinated axons of the optic nerve. *J Neurocytol* **28**:319–331.
- Kalman K, Pennington MW, Lanigan MD, Nguyen A, Rauer H, Mahnir V, Paschetto K, Kern WR, Grissmer S, Gutman GA, et al. (1998) ShK-Dap²², a potent Kv1.3-specific immunosuppressive polypeptide. *J Biol Chem* **273**:32697–32707.
- Koschak A, Bugianesi RM, Mitterdorfer J, Kaczorowski GJ, Garcia ML, and Knaus HG (1998) Subunit composition of brain voltage-gated potassium channels determined by hongotoxin-1, a novel peptide derived from *Centruroides limbatus* venom. *J Biol Chem* **273**:2639–2644.
- Racape J, Lecoq A, Romi-Lebrun R, Liu J, Kohler M, Garcia ML, Menez A, and Gasparini S (2002) Characterization of a novel radio labeled peptide selective for a subpopulation of voltage-gated potassium channels in mammalian brain. *J Biol Chem* **277**:3886–3893.
- UniGene Cluster Hs0.301306.
- Scott VE, Muniz ZM, Sewing S, Lichtinghagen R, Parcej DN, Pongs O, Dolly JO (1994) Antibodies specific for distinct Kv subunits unveil a heterooligomeric basis for subtypes of α -dendrotoxin-sensitive K⁺ channels in bovine brain. *Biochemistry* **33**:1617–1623.
- Smart SL, Bosma MM, and Tempel BL (1997) Identification of the delayed rectifier potassium channel, Kv1.6, in cultured astrocytes. *Glia* **20**:127–134.
- Yuan XJ, Wang J, Juhaszova M, Golovina VA, and Rubin LJ (1998) Molecular basis and function of voltage-gated K⁺ channels in pulmonary arterial smooth muscle cells. *Am J Physiol* **274**:L621–L635.
- Koh SD, Ward SM, Dick GM, Epperson A, Bonner HP, Sanders KM, Horowitz B, and Kenyon JL (1999) Contribution of delayed rectifier potassium currents to the electrical activity of murine colonic smooth muscle. *J Physiol* **515**:475–487.
- Roeper J, Sewing S, Zhang Y, Sommer T, Wanner SG, and Pongs O (1998) NIP domain prevents N-type inactivation in voltage-gated potassium channels. *Nature (Lond)* **391**:390–393.
- Polliak S, Gollan L, Martinez R, Custer A, Einheber S, Salzer JL, Trimmer JS, Shrager P, and Peles E (1999) Caspr2, a new member of the neurexin superfamily, is localized at the juxtaparanodes of myelinated axons and associates with K⁺ channels. *Neuron* **24**:1037–1047.

TABLE 8
K_v1.7 channels

Channel name	K _v 1.7 ¹⁻³
Description	Voltage-gated potassium channel, delayed rectifier
Other names	None
Molecular information	Human: 456aa, NM_031886, chr. 19q13.3 ¹⁻³ , <i>KCNA7</i> , GeneID: 3743, PMID: 11368907 ⁶ Mouse: 532aa, NM_010596, chr. 7 Rat: 457, XM_344889 (predicted), chr. 1q22
Associated subunits	None identified
Functional assays	Voltage-clamp
Current	Possibly a component of I _{K_{ur}} in the heart ³
Conductance	21pS ¹
Ion Selectivity	K ⁺
Activation	Voltage, V _a = -8 mV; τ _n = 6 ms (30 mV) ³
Inactivation	Very slow inactivation
Activators	None
Gating inhibitors	None
Blockers	Flecainide (8 μM), quinidine (15 μM), verapamil (16 μM), amiodarone (35 μM), 4-aminopyridine (150 μM), tetraethyammonium (150 mM) ³
Radioligands	None
Channel distribution	Placenta, amnion, islets (mouse), skeletal muscle, heart, pulmonary arteries ^{4,5}
Physiological functions	K _v 1.7 has properties similar to the ultrarapidly activating I _{K_{ur}} current in the heart ³
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	Can coassemble with other K _v 1 family members in heteromultimers but not with members of other K _v families; only member of this family that has an intron in the coding region ¹⁻³ ; mammalian <i>Shaker</i> -related family

aa, amino acids; chr., chromosome.

1. Kalman K, Nguyen A, Tseng-Crank J, Dukes ID, Chandy G, Hustad CM, Copeland NG, Jenkins NA, Mohrenweiser H, Brandriff B, et al (1998) Genomic organization, chromosomal localization, tissue distribution, and biophysical characterization of a novel mammalian Shaker-related voltage-gated potassium channel, Kv1.7. *J Biol Chem* **273**:5851-5857.

2. Kashuba VI, Kvasha SM, Protopopov AI, Gizatullin RZ, Rynditch AV, Wahlstedt C, Wasserman WW, Zabarovsky ER (2001) Initial isolation and analysis of the human Kv1.7 (KCNA7) gene, a member of the voltage-gated potassium channel gene family. *Gene* **268**:115-122.

3. Bardien-Kruger S, Wulff H, Arieff Z, Brink P, Chandy KG, and Corfield V (2002) Characterization of the human voltage-gated potassium channel gene, KCNA7, a candidate gene for inherited cardiac disorders, and its exclusion as a cause of progressive familial heart block I (PFHBI). *Eur J Human Genet* **10**:36-43.

4. UniGene Cluster Hs0.306973; OMIM no. 176268.

5. Davies AR and Kozlowski RZ (2001) Kv channel subunit expression in rat pulmonary arteries. *Lung* **179**:147-161.

6. Kashuba VI, Kvasha SM, Protopopov AI, Gizatullin RZ, Rynditch AV, Wahlstedt C, Wasserman WW, and Zabarovsky ER (2001) Initial isolation and analysis of the human Kv1.7 (KCNA7) gene, a member of the voltage-gated potassium channel gene family. *Gene* **268**:115-122.

TABLE 9
K_v1.8 channels

Channel name	K _v 1.8
Description	Voltage-gated potassium channel, delayed rectifier
Other names	K _v 1.10, Kcn1 ¹⁻⁵
Molecular information	Human: 511aa, NM_005549, chr. 1p13.1, <i>KCNA10</i> , GeneID: 3744, PMID: 9177773 ¹ Mouse: 503aa, XM_143471 (predicted), chr. 3 Rat: 511aa, XM_227577 (predicted), chr. 2q34
Associated subunits	KCNA4B
Functional assays	Voltage-clamp
Current	Possibly a component of I _{K_{ur}} in the heart ²
Conductance	10-12pS ²
Ion selectivity	K ⁺ /Na ⁺ > 70:1 ²
Activation	V _a = 3.6 mV (oocytes); τ _a = 18 ms at +60 mV (oocytes) ²
Inactivation	τ _h = 10 s
Activators	cGMP
Gating inhibitors	None
Blockers	Barium (5 mM), tetraethyammonium (50 mM), 4-aminopyridine (1.5 mM), charybdotoxin (100 nM), ketoconazole (500 nM), pimoziide (300 nM), verapamil (45 μM) ²
Radioligands	None
Channel distribution	Kidney (cortex > medulla), brain, heart, skeletal muscle, adrenal gland ^{1-3,6}
Physiological functions	Regulation of membrane potential in renal proximal tubule
Mutations and pathophysiology	None
Pharmacological significance	Not established
Comments	Can coassemble with other K _v 1 family members in heteromultimers but not with members of other K _v families; intronless coding region; mammalian <i>Shaker</i> -related family

aa, amino acids; chr., chromosome.

1. Orias M, Bray-Ward P, Curran ME, Keating MT, and Desir GV (1997) Genomic localization of the human gene for KCNA10, a cGMP-activated K channel. *Genomics* **42**:33-37.

2. Lang R, Lee G, Liu W, Tian S, Rafi H, OriasM, Segal AS, and Desir GV (2000) KCNA10: a novel ion channel functionally related to both voltage-gated potassium and CNG cation channels. *Am J Physiol* **278**:F1013-F1021.

3. Yao X, Segal AS, Welling P, Zhang X, McNicholas CM, Engel D, Boulpaep EL, and Desir GV (1995) Primary structure and functional expression of a cGMP-gated potassium channel. *Proc Natl Acad Sci USA* **92**:11711-11715.

4. Yao X, Liu Y, Tung F, and Desir GV (1996) Genomic structure and regulation of Kcn1, a cGMP-gated potassium channel. *Am J Physiol* **271**:F37-F41.

5. Iwasa H, Kurabayashi M, Nagai R, Nakamura Y, and Tanaka T (2001) Genetic variations in five genes involved in the excitement of cardiomyocytes. *J Hum Genet* **46**:549-552.

6. UniGene Cluster Hs0.306973; OMIM no. 176268.

TABLE 10
K_v2.1 channels

Channel name	K _v 2.1 ^{1–3}
Description	Voltage-gated potassium channel, delayed rectifier
Other names	hDRK1, DRK1
Molecular information	Human: 858aa, NM_004975, chr. 20q13.2, ^{4,5} <i>KCNB1</i> , GeneID: 3745, PMID: 8081723 ³⁵ Mouse: 857aa, NM_008420, chr. 2 Rat: 853aa, NM_013186, chr. 3q42
Associated subunits	K _v 5.1, K _v 6.1–K _v 6.3, K _v 8.1, K _v 9.1–K _v 9.3, KChAP (binds to N terminus of K _v 2.1), Fyn SH2 domain ^{6–15}
Functional assays	Voltage-clamp
Current	K _v 2.1/K _v 9.3 (delayed rectifier in oxygen-sensitive pulmonary artery), ⁹ delayed rectifier current in hippocampal and globus pallidus neurons ^{16,17}
Conductance	8pS; on removal of K ⁺ , K _v 2.1 displays a large Na ⁺ conductance that is inhibited by low concentrations of K ⁺ ^{2,18}
Ion selectivity	K ⁺ > Rb ⁺
Activation	Voltage, V _a = 12 mV; k _a = 3 mV ³
Inactivation	Noninactivating
Activators	Linoleic acid ¹⁹
Gating inhibitors	Hanatoxin (42 nM) ^{20,21}
Blockers	Internal tetraethylammonium and tetrapentylammonium, internal Ba ²⁺ (13 μM), external Ba ²⁺ (30 mM), internal Mg ²⁺ , 4-AP (18 mM), halothane ^{22–25}
Radioligands	None
Channel distribution	Brain (cerebral cortex > hippocampus > cerebellum > olfactory bulb; restricted to neurons, where staining is present on dendrites and cell bodies but not on axons; Schwann cells), atria, ventricle, skeletal muscle, retina, cochlea, eye, germ cell, lung, PC12 cells, pulmonary arteries, insulinomas ^{1,3,9,14,16,17,26–33}
Physiological functions	Maintaining membrane potential and modulating electrical excitability in neurons and muscle ^{9,16,17}
Mutations and pathophysiology	K _v 2.1 expression is reduced in chronic hypoxic pulmonary hypertension. ^{30,32}
Pharmacological significance	Not established
Comments	Ser857Asn polymorphism in 0–3% in different ethnic populations ⁵ ; two other single nucleotide polymorphisms have been identified ³⁴ ; the 4-AP binding site is in the S6 inner vestibule. ²³ Mammalian <i>Shab</i> -related family.

aa, amino acids; chr., chromosome; 4-AP, 4-aminopyridine.

1. Frech GC, Van Dongen AM, Schuster G, Brown AM, and Joho RH (1989) A novel potassium channel with delayed rectifier properties isolated from rat brain by expression cloning. *Nature (Lond)* **340**:642–645.

2. Hartmann HA, Kirsch GE, Drewe JA, Tagliatela M, Joho RH, and Brown AM (1991) Exchange of conduction pathways between two related K⁺ channels. *Science (Wash DC)* **251**:942–944.

3. Albrecht B, Lorra C, Stocker M, and Pongs O (1993) Cloning and characterization of a human delayed rectifier potassium channel gene. *Receptors Channels* **1**:99–110.

4. Melis R, Stauffer D, Zhao X, Zhu XL, Albrecht B, Pongs O, Brothman A, and Leppert M (1995) Physical and genetic localization of a Shab subfamily potassium channel (KCNB1) gene to chromosomal region 20q13.2. *Genomics* **25**:285–287.

5. Mazzanti CM, Bergen A, Enoch MA, Michelini S, and Goldman D (1996) Identification of a Ser857-Asn857 substitution in DRK1 (KCNB1), population frequencies and lack of association to the low voltage α EEG trait. *Hum Genet* **98**:134–137.

6. Post MA, Kirsch GE, and Brown AM (1996) Kv2.1 and electrically silent Kv6.1 potassium channel subunits combine and express a novel current. *FEBS Lett* **399**:177–182.

7. Salinas M, de Weille J, Guillemare E, Lazdunski M, and Hugnot JP (1997) Modes of regulation of *Shab* K⁺ channel activity by the Kv8.1 subunit. *J Biol Chem* **272**:8774–8780.

8. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian *Shab* K⁺ channels. *J Biol Chem* **272**:24371–24379.

9. Patel AJ, Lazdunski M, and Honore E (1997) Kv2.1/Kv9.3, a novel ATP-dependent delayed-rectifier K⁺ channel in oxygen-sensitive pulmonary artery myocytes. *EMBO J* **16**:6615–6625.

10. Sobko A, Peretz A, and Attali B (1998) Constitutive activation of delayed-rectifier potassium channels by a src family tyrosine kinase in Schwann cells. *EMBO J* **17**:4723–4734.

11. Wible BA, Yang Q, Kuryshv YA, Accili EA, and Brown AM (1998) Cloning and expression of a novel K⁺ channel regulatory protein, KChAP. *J Biol Chem* **273**:11745–11751.

12. Kramer JW, Post MA, Brown AM, and Kirsch GE (1998) Modulation of potassium channel gating by co expression of Kv2.1 with regulatory Kv5.1 or Kv6.1 α-subunits. *Am J Physiol* **274**:C1501–C1510.

13. Chiara MD, Monje F, Castellano A, and Lopez-Barneo J (1999) A small domain in the N terminus of the regulatory α-subunit Kv2.3 modulates Kv2.1 potassium channel gating. *J Neurosci* **19**:6865–6873.

14. Kuryshv YA, Wible BA, Gudzi TI, Ramirez AN, and Brown AM (2001) KChAP/Kvβ1.2 interactions and their effects on cardiac Kv channel expression. *Am J Physiol Cell Physiol* **281**:C290–C299.

15. Sano Y, Mochizuki S, Miyake A, Kitada C, Inamura K, Yokoi H, Nozawa K, Matsushime H, and Furuichi, K (2002) Molecular cloning and characterization of Kv6.3, a novel modulatory subunit for voltage-gated K⁺ channel Kv2.1. *FEBS Lett* **512**:230–234.

16. Murakoshi H, and Trimmer JS (1999) Identification of the Kv2.1 K⁺ channel as a major component of the delayed rectifier K⁺ current in rat hippocampal neurons. *J Neurosci* **19**:1728–1735.

17. Baranauskas G, Tkatch T, and Surmeier DJ (1999) Delayed rectifier currents in rat globus pallidus neurons are attributable to Kv2.1 and Kv3.1/3.2 K⁺ channels. *J Neurosci* **19**:6394–6404.

18. Korn SJ and Ikeda SR (1995) Permeation selectivity by competition in a delayed rectifier potassium channel. *Science (Wash DC)* **269**:410–412.

19. McKay MC and Worley JF 3rd (2001) Linoleic acid both enhances activation and blocks Kv1.5 and Kv2.1 channels by two separate mechanisms. *Am J Physiol Cell Physiol* **281**:C1277–C1284.

20. Swartz KJ and MacKinnon R (1995) An inhibitor of the Kv2.1 potassium channel isolated from the venom of a Chilean tarantula. *Neuron* **15**:941–949.

21. Swartz KJ and MacKinnon R (1997) Hanatoxin modifies the gating of a voltage-dependent K⁺ channel through multiple binding sites. *Neuron* **18**:665–673.

22. Tagliatela M, Vandongen AM, Drewe JA, Joho RH, Brown AM, and Kirsch GE (1991) Patterns of internal and external tetraethylammonium block in four homologous K⁺ channels. *Mol Pharmacol* **40**:299–307.

23. Kirsch GE, Shieh CC, Drewe JA, Vener DF, and Brown AM (1993) Segmental exchanges define 4-aminopyridine binding and the inner mouth of K⁺ pores. *Neuron* **11**:503–512.

24. Tagliatela M, Drewe JA, and Brown AM (1993) Barium blockade of a clonal potassium channel and its regulation by a critical pore residue. *Mol Pharmacol* **44**:180–190.

25. Kulkarni RS, Zorn LJ, Anantharam V, Bayley H, and Treistman SN (1996) Inhibitory effects of ketamine and halothane on recombinant potassium channels from mammalian brain. *Anesthesiology* **84**:900–909.

26. Trimmer JS (1991) Immunological identification and characterization of a delayed rectifier K⁺ channel polypeptide in rat brain. *Proc Natl Acad Sci USA* **88**:10764–10768.

27. Hwang PM, Fotuhi M, Brecht DS, Cunningham AM, and Snyder SH (1993) Contrasting immunohistochemical localizations in rat brain of two novel K⁺ channels of the Shab subfamily. *J Neurosci* **13**:1569–1576.

28. Sharma N, D'Arcangelo G, Kleinlaus A, Halegoua S, and Trimmer JS (1993) Nerve growth factor regulates the abundance and distribution of K⁺ channels in PC12 cells. *J Cell Biol* **123**:1835–1843.

29. Scannevin RH, Murakoshi H, Rhodes KJ, and Trimmer JS (1996) Identification of a cytoplasmic domain important in the polarized expression and clustering of the Kv2.1 K⁺ channel. *J Cell Biol* **135**:1619–1632.

30. Archer SL, Souil E, Dinh-Xuan AT, Schremmer B, Mercier JC, El Yaagoubi A, Nguyen-Huu L, Reeve HL, and Hampl V (1998) Molecular identification of the role of voltage-gated K⁺ channels, Kv1.5 and Kv2.1, in hypoxic pulmonary vasoconstriction and control of resting membrane potential in rat pulmonary artery myocytes. *J Clin Invest* **101**:2319–2330.

31. MacDonald PE, Ha XF, Wang J, Smukler SR, Sun AM, Gaisano HY, Salapatek AM, Backx PH, and Wheeler MB (2001) Members of the Kv1 and Kv2 voltage-dependent K⁺ channel families regulate insulin secretion. *Mol Endocrinol* **15**:1423–1435.

32. Michelakis ED, McMurtry MS, Wu XC, Dyck JR, Moudgil R, Hopkins TA, Lopaschuk GD, Puttagunta L, Waite R, and Archer SL (2002) Dichloroacetate, a metabolic modulator, prevents and reverses chronic hypoxic pulmonary hypertension in rats: role of increased expression and activity of voltage-gated potassium channels. *Circulation* **105**:244–250.

33. UniGene Cluster Hs0.84244; OMIM no. 600397.

34. Iwasa H, Kurabayashi M, Nagai R, Nakamura Y, and Tanaka, T (2001) Multiple single-nucleotide polymorphisms (SNPs) in the Japanese population in six candidate genes for long QT syndrome. *J Hum Genet* **46**:158–162.

35. Albrecht B, Lorra C, Stocker M, and Pongs O (1993) Cloning and characterization of a human delayed rectifier potassium channel gene. *Receptor Channels* **1**:99–110.

TABLE 11
K_v2.2 channels

Channel name	K _v 2.2 ^{1–3}
Description	Voltage-gated potassium channel, delayed rectifier
Other names	CDRK
Molecular information	Human: 911 aa, NM_004770, chr. 8q13.2, <i>KCNB2</i> , GeneID: 9312, PMID: 9612272 ¹⁵ Mouse: 758 aa, XM_136482 (predicted), chr. 1 Rat: 802 aa, NM_054000, chr. 5q11
Associated subunits	Mouse K _{vβ} 4 associates with K _v 2.2 and enhances expression level, K _v 8.1, K _v 9, KChAP ^{4–7}
Functional assays	Voltage-clamp
Current	None determined
Conductance	15pS ⁸
Ion selectivity	K ⁺ -selective
Activation	Voltage
Inactivation	Noninactivating
Activators	None
Gating inhibitors	None
Blockers	Quinine (13.7 μM), tetraethylammonium (2.6 mM), 4-aminopyridine (1.5 mM), phencyclidine (μM) ^{8,9}
Radioligands	None
Channel distribution	Brain [olfactory bulb (granule cell layer > olfactory tubercle) > cortex > hippocampus > cerebellum; hypothalamus], ventricle, tongue, sympathetic neurons, gastrointestinal smooth muscle, mesenteric artery smooth muscle ^{1–3,10–14}
Physiological functions	Maintaining membrane potential, modulating electrical excitability in neurons
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	The angiotensin II type 1 receptor mediates inhibition of K _v 2.2 in brainstem and hypothalamic neurons ¹² ; mammalian <i>Shab</i> -related family

aa, amino acids; chr., chromosome.

1. Hwang PM, Glatt CE, Brecht DS, Yellen G, and Snyder SH. (1992). A novel K⁺ channel with unique localizations in mammalian brain: molecular cloning and characterization. *Neuron* **8**:473–481.

2. Hwang PM, Fotuhi M, Brecht DS, Cunningham AM, and Snyder SH (1993) Contrasting immunohistochemical localizations in rat brain of two novel K⁺ channels of the Shab subfamily. *J Neurosci* **13**:1569–1576.

3. Hwang PM, Cunningham AM, Peng YW, and Snyder SH (1993) CDRK and DRK1 K⁺ channels have contrasting localizations in sensory systems. *Neuroscience* **55**:613–620.

4. Fink M, Duprat F, Lesage F, Heurteaux C, Romey G, Barhanin J, and Lazdunski M (1996) A new K⁺ channel β subunit to specifically enhance Kv2.2 (CDRK) expression. *J Biol Chem* **271**:26341–26348.

5. Salinas M, de Weille J, Guillemare E, Lazdunski M, and Hugnot JP (1997) Modes of regulation of *Shab* K⁺ channel activity by the Kv8.1 subunit. *J Biol Chem* **272**:8774–8780.

6. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian *Shab* K⁺ channels. *J Biol Chem* **272**:24371–24379.

7. Wible BA, Yang Q, Kuryshv YA, Accili EA, and Brown AM (1998) Cloning and expression of a novel K⁺ channel regulatory protein, KChAP. *J Biol Chem* **273**:11745–11751.

8. Schmalz F, Kinsella J, Koh SD, Vogalis F, Schneider A, Flynn ER, Kenyon JL, and Horowitz B (1998) Molecular identification of a component of delayed rectifier current in gastrointestinal smooth muscles. *Am J Physiol* **274**:G901–G911.

9. Frey BW, Lynch FT, Kinsella JM, Horowitz B, Sanders KM, and Carl A (2000) Blocking of cloned and native delayed rectifier K channels from visceral smooth muscles by phencyclidine. *Neurogastroenterol Motil* **12**:509–516.

10. Maletic-Savatic M, Lenn NJ, and Trimmer JS (1995) Differential spatiotemporal expression of K⁺ channel polypeptides in rat hippocampal neurons developing in situ and in vitro. *J Neurosci* **15**:3840–3851.

11. Dixon JE and McKinnon D (1996) Potassium channel mRNA expression in prevertebral and para-vertebral sympathetic neurons. *Eur J Neurosci* **8**:183–191.

12. Gelband CH, Warth JD, Mason HS, Zhu M, Moore JM, Kenyon JL, Horowitz B, and Sumners C (1999) Angiotensin II type 1 receptor-mediated inhibition of K⁺ channel subunit Kv2.2 in brain stem and hypothalamic neurons. *Circ Res* **84**:352–359.

13. Xu C, Lu Y, Tang G, and Wang R (1999) Expression of voltage-dependent K⁺ channel genes in mesenteric artery smooth muscle cells. *Am. J. Physiol* **277**:G1055–G1063.

14. Lim ST, Antonucci DE, Scannevin RH, and Trimmer JS (2000) A novel targeting signal for proximal clustering of the Kv2.1 K⁺ channel in hippocampal neurons. *Neuron* **25**:385–397.

15. Schmalz F, Kinsella J, Koh SD, Vogalis F, Schneider A, Flynn ER, Kenyon JL, and Horowitz B (1998) Molecular identification of a component of delayed rectifier current in gastrointestinal smooth muscles. *Am J Physiol* **274**:G901–G911.

TABLE 12
K_v3.1 channels

Channel name	K _v 3.1
Description	Voltage-gated potassium channel, delayed rectifier
Other names	Kv3.1, ¹ NGK2, ² KV4, ³ KShIIIB, ¹⁵ Raw2, ⁴ type <i>l</i> channel in T cells ⁵
Molecular information	Human: 511aa, NM_004976, chr. 11p15, ^{1-4,16} <i>KCNC1</i> , GeneID: 3746, PMID: 1400413 ¹ Mouse: 511aa, NM_008421, chr. 7 Rat: 585aa, NM_012856, chr. 1q22
Associated subunits	Not established
Functional assays	Electrophysiology
Current	Delayed rectifier
Conductance	27pS ^{1,5}
Ion selectivity	K ⁺ (1) > Rb ⁺ (0.76) > NH ₄ ⁺ (0.12) = Cs ⁺ (0.12) > Na ⁺ (0.004) ⁶
Activation	V _a = 16 mV; k _a = 10 mV; τ _a = 2 ms (40 mV) ⁷
Inactivation	τ _h = 630 ms (40 mV) ¹
Activators	None
Gating inhibitors	None
Blockers	4-Aminopyridine (29 μM), capsaicin (158 μM), resiniferatoxin (46 μM), flecainide (108 μM), nifedipine (131 μM), diltiazem (97 μM), cromakalim (237 μM), tetraethylammonium (0.2 mM) ⁸
Radioligands	None
Channel distribution	Brain (cerebellum > globus pallidus, subthalamic nucleus, substantia nigra > reticular thalamic nuclei, cortical and hippocampal interneurons > inferior colliculi, cochlear and vestibular nuclei), skeletal muscle, human Louckes B cells, germ cell, lung, testis, AtT20 cell line ^{9-13,19,20}
Physiological functions	Important for the high-firing frequency of auditory ⁸ and fast-spiking GABAergic interneurons ^{11,21} ; regulation of action potential duration in presynaptic terminals ^{17,18}
Mutations and pathophysiology	Kv3.1-/- mice exhibit impaired motor skills and reduced muscle contraction force ¹³ ; Kv3.1/Kv3.3 double knockout mice display severe ataxia, myoclonus, and hypersensitivity to ethanol ¹⁴
Pharmacological significance	Not established
Comments	H-ras oncogene switches anterior pituitary-derived cells (AtT20) to a more neuron-like phenotype in parallel with the induction of expression of K _v 3.1 ¹² ; mammalian <i>Shaw</i> -related family

aa, amino acids; chr., chromosome.

1. Grissmer S, Ghanshani S, Dethlefs B, McPherson JD, Wasmuth JJ, Gutman GA, Cahalan MD, and Chandy KG (1992) The Shaw-related potassium channel gene, Kv3.1, on human chromosome 11, encodes the type I K⁺ channel in T cells. *J Biol Chem* **267**:20971–20979.
2. Yokoyama S, Imoto K, Kawamura T, Higashida H, Iwabe N, Miyata T, and Numa S (1989) Potassium channels from NG108–15 neuroblastoma-glioma hybrid cells: primary structure and functional expression from cDNAs. *FEBS Lett* **259**:37–42.
3. Luneau CJ, Williams JB, Marshall J, Levitan ES, Oliva C, Smith JS, Antanavage J, Folander K, Stein RB, and Swanson R (1991) Alternative splicing contributes to K⁺ channel diversity in the mammalian central nervous system. *Proc Natl Acad Sci USA* **88**:3932–3936.
4. Rettig J, Wunder F, Stocker M, Lichtenhagen R, Mastiaux F, Beckh S, Kues W, Pedarzani P, Schroter KH, Ruppersberg JP, et al. (1992) Characterization of a Shaw-related potassium channel family in rat brain. *EMBO J* **11**:2473–2486.
5. Decoursey TE, Chandy KG, Gupta S, and Cahalan MD (1987) Two types of potassium channels in murine T lymphocytes. *J Gen Physiol* **89**:379–404.
6. Shapiro MS and DeCoursey TE (1991) Selectivity and gating of the type I potassium channel in mouse lymphocytes. *J Gen Physiol* **97**:1227–1250.
7. Shapiro MS and DeCoursey TE (1991) Permeant ion effects on gating kinetics of the type I potassium channel in mouse lymphocytes. *J Gen Physiol* **97**:1251–1278.
8. Grissmer S, Nguyen AN, Aiyar J, Hanson DC, Mather RJ, Gutman GA, Karmilowicz MJ, Auferin DD, and Chandy KG (1994) Pharmacological characterization of five cloned voltage-gated K⁺ channels, types Kv1.1, 1.2, 1.3, 1.5, and 3.1, stably expressed in mammalian cell lines. *Mol Pharmacol* **45**:1227–1234.
9. UniGene ClusterHs0.181768; OMIM no. 176258.
10. Wang LY, Gan L, Forsythe ID, and Kaczmarek LK (1998) Contribution of the Kv3.1 potassium channel to high-frequency firing in mouse auditory Neurons. *J Physiol* **509**:183–194.
11. Massengill JL, Smith MA, Son DI, and O'Dowd DK (1997) Differential expression of K4-AP current and Kv3.1 potassium channel transcripts in cortical neurons that develop distinct firing phenotypes. *J Neurosci* **17**:3136–3147.
12. Hemmick LM, Perney TM, Flamm RE, Kaczmarek LK, and Birnberg NC (1992) Expression of the H-ras oncogene induces potassium conductance and neuron-specific potassium channel mRNAs in the AtT20 cell line. *J Neurosci* **12**:2007–2014.
13. Ho CS, Grange RW, and Joho RH (1997) Pleiotropic effects of a disrupted K⁺ channel gene: reduced body weight, impaired motor skill and muscle contraction, but no seizures. *Proc Natl Acad Sci USA* **94**:1533–1558.
14. Espinosa F, McMahon A, Chan E, Wang S, Ho CS, Heintz N, and Joho RH (2001) Alcohol hyper-sensitivity, increased locomotion, and spontaneous myoclonus in mice lacking the potassium channels Kv3.1 and Kv3.3. *J Neurosci* **21**:6657–6665.
15. Haas M, Ward DC, Lee J, Roses AD, Clarke V, D'Eustachio P, Lau D, Vega-Saenz de Miera E, and Rudy B (1993) Localization of Shaw-related K⁺ channel genes on mouse and human chromosomes. *Mamm Genome* **4**:711–715.
16. Ried T, Rudy B, Vega-Saenz de Miera E, Lau D, Ward DC, and Sen K (1993) Localization of a highly conserved potassium channel gene (NGK2-KV4; KCNC1) to chromosome 11p15. *Genomics* **15**:405–411.
17. Ishikawa T, Nakamura Y, Saitoh N, Li WB, Iwasaki S, and Takahashi T (2003) Distinct roles of Kv1 and Kv3 potassium channels at the calyx of Held presynaptic terminal. *J Neurosci* **23**:10445–10453.
18. Devaux J, Alcaraz G, Grinspan J, Bennett V, Joho R, Crest M, and Scherer SS (2003) Kv3.1b is a novel component of CNS nodes. *J Neurosci* **23**:4509–4518.
19. Weiser M, Vega-Saenz de Miera E, Kentros C, Moreno H, Franzen L, Hillman D, Baker H, and Rudy B (1994) Differential expression of Shaw-related K⁺ channels in the rat central nervous system. *J Neurosci* **14**:949–972.
20. Weiser M, Bueno E, Sekirnjak C, Martone ME, Baker H, Hillman D, Thornhill W, Ellisman M, and Rudy B (1995) The potassium channel subunit KV3.1b is localized to somatic and axonal membranes of specific populations of CNS neurons. *J Neurosci* **15**:4298–4314.
21. Erisir A, Lau D, Rudy B, and Leonard CS (1999) The function of specific K⁺ channels in sustained high frequency firing of fast-spiking neocortical interneurons. *J Neurophysiol* **82**:2476–2489.

TABLE 13
K_v3.2 channels

Channel name	K _v 3.2
Description	Voltage-gated potassium channel, delayed rectifier
Other names	RKShIIIa, ¹ Raw1, ² Kv3.2a, ³ rKv3.2b and rKv3.2c ⁴
Molecular information	Human: 613aa, NM_139136 (transcript variant 1), chr. 12q14.1, ⁵ <i>KCNC2</i> , GeneID: 3747, PMID: 8111118 ²¹ Mouse: AC121610 (genomic), chr. 10 Rat: 613aa, NM_139216 (transcript variant a), chr. 7q12–22
Associated subunits	None
Functional assays	Electrophysiology
Current	Delayed rectifier
Conductance	16–20pS ¹⁶
Ion selectivity	K ⁺
Activation	V _a = 13 mV; k _a = 7–7.5 mV ¹ ; t _{on} = 10–90% (40 mV) 4 ms; τ _{off} 2.9 ms (–60 mV) ¹⁶
Inactivation	Very slow ¹⁶
Activators	None
Gating inhibitors	None
Blockers	Tetraethylammonium (0.1 mM), ⁶ 4-aminopyridine (0.1 mM), ⁶ 8-bromo-cGMP, ⁷ 3-isobutyl-1-methylxanthine, ⁶ D-NONOate, ⁷ verapamil (11 μM), ⁸ ShK ¹⁹
Radioligands	None
Channel distribution	Brain (fast-spiking GABAergic interneurons of the neocortex, hippocampus, and caudate; terminal fields of thalamocortical projections), ^{9–12} islets, ¹³ mesenteric artery, Schwann cells ¹⁴
Physiological functions	Probably in heteromeric complexes with K _v 3.1; important for the high-frequency firing of fast spiking GABAergic interneurons ¹⁷ and GABA release via regulation of action potential duration in presynaptic terminals ¹⁸ ; modulated by protein kinase A in vitro and in vivo ^{10,20}
Mutations and pathophysiology	See “Comments”
Pharmacological significance	Not established
Comments	Fast deactivation; knockout mice show specific alterations in their cortical electroencephalographic patterns and an increased susceptibility to epileptic seizures consistent with an impairment of a cortical inhibitory mechanism ¹⁵ ; mammalian <i>Shaw</i> -related family

aa, amino acids; chr., chromosome; D-NONOate, 1,1-diethyl-2-hydroxy-2-nitrosodiazine; ShK, *Stichodactyla helianthus* toxin.

1. McCormack T, Vega-Saenz de Miera EC, and Rudy B (1991) Molecular cloning of a member of a third class of Shaker-family K⁺ channel genes in mammals. *Proc Natl Acad Sci USA* **87**:5227–5231.

2. Rettig J, Wunder F, Stocker M, Lichtinghagen R, Mastiaux F, Beckh S, Kues W, Pedarzani P, Schroter KH, Ruppersberg JP, et al. (1992) Characterization of a Shaw-related potassium channel family in rat brain. *EMBO J* **11**:2473–2486.

3. Ponce A, Vega-Saenz de Miera E, Kentros C, Moreno H, Thornhill B, and Rudy B (1997) K⁺ channel subunit isoforms with divergent carboxy-terminal sequences carry distinct membrane targeting signals. *J Membr Biol* **159**:149–159.

4. Luneau C, Wiedmann R, Smith JS, and Williams JB (1991) Shaw-like rat brain potassium channel cDNA's with divergent 3' ends. *FEBS Lett* **288**:163–167.

5. Haas M, Ward DC, Lee J, Roses AD, Clarke V, D'Eustachio P, Lau D, Vega-Saenz de Miera E, and Rudy B (1993) Localization of Shaw-related K⁺ channel genes on mouse and human chromosomes. *Mamm Genome* **4**:711–715.

6. Lien CC, Martina M, Schultz JH, Ehmke H, and Jonas P (2002) Gating, modulation and subunit composition of voltage-gated K⁺ channels in dendritic inhibitory interneurons of rat hippocampus. *J Physiol* **538**:405–419.

7. Moreno H, Vega-Saenz de Miera E, Nadal MS, Amarillo Y, and Rudy B (2001) Modulation of Kv3 potassium channels expressed in CHO cells by a nitric oxide-activated phosphatase. *J Physiol* **530**:345–358.

8. Madeja M, Muller V, Musshoff U, and Speckmann EJ (2001) Sensitivity of native and cloned hippocampal delayed-rectifier potassium channels to verapamil. *Neuropharmacology* **39**:202–210.

9. Weiser M, Vega-Saenz de Miera E, Kentros C, Moreno H, Franzen L, Hillman D, Baker H, and Rudy B (1994) Differential expression of Shaw-related K⁺ channels in the rat central nervous system. *J Neurosci* **14**:949–972.

10. Moreno H, Kentros C, Bueno E, Weiser M, Hernandez A, Vega-Saenz de Miera E, Ponce A, Thornhill W, and Rudy B (1995) Thalamocortical projections have a K⁺ channel that is phosphorylated and modulated by cAMP-dependent protein kinase. *J Neurosci* **15**:5486–5501.

11. Chow A, Erisir A, Farb C, Nadal MS, Ozaita A, Lau D, Welker E, and Rudy B (1999) K⁺ channel expression distinguishes subpopulations of parvalbumin- and somatostatin-containing neocortical interneurons. *J Neurosci* **19**:9332–9345.

12. Tansey EP, Chow A, Rudy B, and McBain CJ (2002) Developmental expression of potassium-channel subunit Kv3.2 within subpopulations of mouse hippocampal inhibitory interneurons. *Hippocampus* **12**:137–148.

13. Roe MW, Worley JF 3rd, Mittal AA, Kuznetsov A, Dasgupta S, Mertz RJ, Witherspoon SM 3rd, Blair N, Lancaster ME, McIntyre MS, et al. (1996) Expression and function of pancreatic β-cell delayed rectifier K⁺ channels: role in stimulus-secretion coupling. *J Biol Chem* **271**:32241–32246.

14. Sobko A, Peretz A, Shiriha O, Etkin S, Cherepanova V, Dagan D, and Attali B (1998) Heteromultimeric delayed-rectifier K⁺ channels in Schwann cells: developmental expression and role in cell proliferation. *J Neurosci* **18**:10398–10408.

15. Lau D, Vega-Saenz de Miera EC, Contreras D, Ozaita A, Harvey M, Chow A, Noebels JL, Paylor R, Morgan JJ, Leonard CS, and Rudy B (2000) Impaired fast-spiking, suppressed cortical inhibition, and increased susceptibility to seizures in mice lacking Kv3.2 K⁺ channel proteins. *J Neurosci* **20**:9071–9085.

16. Rudy B, Chow A, Lau D, Amarillo Y, Ozaita A, Saganich M, Moreno H, Nadal MS, Hernandez-Pineda R, Hernandez-Cruz A, et al. (1999) Contributions of Kv3 channels to neuronal excitability. *Ann NY Acad Sci* **868**:304–343.

17. Erisir A, Lau D, Rudy B, and Leonard CS (1999) The function of specific K⁺ channels in sustained high frequency firing of fast-spiking neocortical interneurons. *J Neurophysiol* **82**:2476–2489.

18. Goldberg EM, Watanabe S, Chang SY, Joho RH, Huang ZJ, Leonard CS, and Rudy B (2005) Specific functions of synaptically localized potassium channels in synaptic transmission at the neocortical GABAergic fast-spiking cell synapse. *J Neurosci* **25**:5230–5235.

19. Yan L, Herrington J, Goldberg E, Dulski PM, Bugianesi RM, Slaughter RS, Banerjee P, Brochu RM, Priest BT, Kaczorowski GJ, et al. (2005) *Stichodactyla helianthus* peptide, a pharmacological tool for studying Kv3.2 channels. *Mol Pharmacol* **67**:1513–1521.

20. Atzori M, Lau D, Tansey EP, Chow A, Ozaita A, Rudy B, and McBain CJ (2000) H₂ histamine receptor-phosphorylation of Kv3.2 modulates interneuron fast spiking. *Nat Neurosci* **3**:791–798.

21. Haas M, Ward DC, Lee J, Roses AD, Clarke V, D'Eustachio P, Lau D, Vega-Saenz de Miera E, and Rudy B (1993) Localization of Shaw-related K⁺ channel genes on mouse and human chromosomes. *Mamm Genome* **4**:711–715.

TABLE 14
K_v3.3 channels

Channel name	K _v 3.3 ¹⁻⁴
Description	Voltage-gated A-type potassium channel ²
Other names	hKv3.3, mKv3.3, ¹ RKShIIID, ³ Kv3.3b ⁴
Molecular information	Human: 757aa, NM_004977, chr. 19q13.3-4, ¹⁻³ <i>KCNK3</i> , GeneID: 3748, PMID: 1740329 ¹ Mouse: 679aa, NM_008422, chr. 7 Rat: 770aa, NM_053997, chr. 1q22
Associated subunits	None
Functional assays	Electrophysiology
Current	A-type
Conductance	Not established
Ion selectivity	K ⁺
Activation	$V_a = 7$ mV; $k_a = 6$ mV ²
Inactivation	$\tau_h \sim 200$ ms (40 mV) ²
Activators	None
Gating inhibitors	None
Blockers	Tetraethylammonium (0.14 mM), ² 4-aminopyridine (1.2 mM) ² ; blocked by hypoxia ⁵
Radioligands	None
Channel distribution	Brain, Purkinje cells, central nervous system motoneurons; auditory brainstem ¹² ; electrosensory, cerebellar neurons, central auditory nuclei ⁶⁻⁸ ; mesenteric artery ⁹ ; lens and corneal epithelium ¹⁰
Physiological functions	Not established
Mutations and pathophysiology	See "Comments"
Pharmacological significance	Not established
Comments	Alcohol hypersensitivity, ataxia, increased locomotion and myoclonus occur in mice lacking K _v 3.3 and K _v 3.1 ¹¹ ; mammalian <i>Shaw</i> -related family

aa, amino acids; chr., chromosome.

1. Ghanshani S, Pak M, McPherson JD, Strong M, Dethlefs B, Wasmuth JJ, Salkoff L, Gutman GA, and Chandy KG (1992) Genomic organization, nucleotide sequence, and cellular distribution of a Shaw-related potassium channel gene, Kv3.3, and mapping of Kv3.3 and Kv3.4 to human chromosomes 19 and 1. *Genomics* **12**:190–196.

2. Vega-Saenz de Miera E, Moreno H, Fruhling D, Kentros C, and Rudy B (1992) Cloning of ShIII (Shaw-like) cDNAs encoding a novel high-voltage-activating, TEA-sensitive, type-A K⁺ channel. *Proc R Soc Lond B Biol Sci* **248**:9–18.

3. Haas M, Ward DC, Lee J, Roses AD, Clarke V, D'Eustachio P, Lau D, Vega-Saenz de Miera E, and Rudy B (1993) Localization of Shaw-related K⁺ channel genes on mouse and human chromosomes. *Mamm Genome* **4**:711–715.

4. Goldman-Wohl DS, Chan E, Baird D, and Heintz N (1994) Kv3.3b: a novel Shaw type potassium channel expressed in terminally differentiated cerebellar Purkinje cells and deep cerebellar nuclei. *J Neurosci* **14**:511–522.

5. Patel AJ and Honore E (2001) Molecular physiology of oxygen-sensitive potassium channels. *Eur Respir J* **18**:221–227.

6. Rashid AJ, Dunn RJ, and Turner RW (2001) A prominent soma-dendritic distribution of Kv3.3 K⁺ channels in electrosensory and cerebellar neurons. *J Comp Neurol* **441**:234–247.

7. Grigg JJ, Brew HM, and Tempel BL (2000) Differential expression of voltage-gated potassium channel genes in auditory nuclei of the mouse brainstem. *Hear Res* **140**:77–90.

8. Li W, Kaczmarek LK, and Perney TM (2001) Localization of two high-threshold potassium channel subunits in the rat central auditory system. *J Comp Neurol* **437**:196–218.

9. Xu C, Lu Y, Tang G, and Wang R (1999) Expression of voltage-dependent K⁺ channel genes in mesenteric artery smooth muscle cells. *Am J Physiol* **277**:G1055–G1063.

10. Rae JL and Shepard AR (2000) Kv3.3 potassium channels in lens epithelium and corneal endothelium. *Exp Eye Res* **70**:339–348.

11. Espinosa F, McMahon A, Chan E, Wang S, Ho CS, Heintz N, and Joho RH (2001) Alcohol hyper-sensitivity, increased locomotion, and spontaneous myoclonus in mice lacking the potassium channels Kv3.1 and Kv3.3. *J Neurosci* **21**:6657–6665.

12. Weiser M, Bueno E, Sekirnjak C, Martone ME, Baker H, Hillman D, Chen S, Thornhill W, Ellisman M, and Rudy B (1995) The potassium channel subunit KV3.1b is localized to somatic and axonal membranes of specific populations of CNS neurons. *J Neurosci* **15**:4298–4314.

TABLE 15
K_v3.4 channels

Channel name	K _v 3.4
Description	Voltage-gated potassium channel, A-type, fast-inactivating
Other names	Raw3, ¹ HKShIIIC, ² mKv3.4 ³
Molecular information	Human: 635 aa, NM_004978 (transcript variant 1), chr. 1p21 ^{1,2} , <i>KCNC4</i> , GeneID: 3749, PMID: 1920536 ² Mouse: 628 aa, NM_145922, chr. 3 Rat:
Associated subunits	MiRP2 forms potassium channels in skeletal muscle with K _v 3.4 ⁴
Functional assays	Electrophysiology
Current	A-type
Conductance	14pS ^{1,5}
Ion selectivity	K ⁺
Activation	V _a = 3.4 mV ⁵ , +14 mV ¹ ; k _a = 8.4 mV ⁵
Inactivation	N-type inactivation, V _h = 53 mV; k _h = 7.4 mV; τ _h = 15.9 ms (50 mV) ^{1,2,5}
Activators	None
Gating inhibitors	None
Blockers	BDS-I (47 nM), ⁶ tetraethylammonium (0.3 mM) ^{1,5} ; the specificity of BDS-I for K _v 3.4 has been questioned ¹²
Radioligands	None
Channel distribution	Parathyroid, prostate, brain ⁷ (brainstem, hippocampal granule cells), ⁸ skeletal muscle, ^{4,8,9} pancreatic acinar cells ^{10,11}
Physiological functions	Together with MiRP2 forms low-voltage-ctivating potassium channels that regulate skeletal muscle resting potential ⁴
Mutations and pathophysiology	Mutations of MiRP2, which associates with K _v 3.4 in skeletal muscle, are associated with periodic paralysis ⁴
Pharmacological significance	Not established
Comments	Mammalian <i>Shaw</i> -related family

aa, amino acids; chr., chromosome.

1. Schroter KH, Ruppersberg J, Wunder F, Rettig J, Stocker M, and Pongs O (1991) Cloning and functional expression of a TEA-sensitive A-type potassium channel from rat brain. *FEBS Lett* **278**:211–216.

2. Rudy B, Sen K, Vega-Saenz de Miera E, Lau D, Ried T, and Ward DC (1991) Cloning of a human cDNA expressing a high voltage-activating, TEA-sensitive, type-A K⁺ channel which maps to chromosome 1 band p21. *J Neurosci Res* **29**:401–412.

3. Ghanshani S, Pak M, McPherson JD, Strong M, Dethlefs B, Wasmuth JJ, Salkoff L, Gutman GA, and Chandy KG (1992) Genomic organization, nucleotide sequence, and cellular distribution of a Shaw-related potassium channel gene, Kv3.3, and mapping of Kv3.3 and Kv3.4 to human chromosomes 19 and 1. *Genomics* **12**:190–196.

4. Abbott GW, Butler MH, Bendahhou S, Dalakas MC, Ptacek LJ, and Goldstein SA (2001) MiRP2 forms potassium channels in skeletal muscle with Kv3.4 and is associated with periodic paralysis. *Cell* **104**:217–231.

5. Rettig J, Wunder F, Stocker M, Lichtinghagen R, Mastiaux F, Beckh S, Kues W, Pedarzani P, Schroter KH, Ruppersberg JP, et al. (1992) Characterization of a Shaw-related potassium channel family in rat brain. *EMBO J* **11**:2473–2486.

6. Diochot S, Schweitz H, Beress L, and Lazdunski M (1998) Sea anemone peptides with a specific blocking activity against the fast inactivating potassium channel Kv3.4. *J Biol Chem* **273**:6744–6749.

7. Riazanski V, Becker A, Chen J, Sochivko D, Lie A, Wiestler OD, Elger CE, and Beck H (2001) Functional and molecular analysis of transient voltage-dependent K⁺ currents in rat hippocampal granule cells. *J Physiol* **537**:391–406.

8. Weiser M, Vega-Saenz de Miera E, Kentros C, Moreno H, Franzen L, Hillman D, Baker H, and Rudy B (1994) Differential expression of Shaw-related K⁺ channels in the rat central nervous system. *J Neurosci* **14**:949–972.

9. Vullhorst D, Klocke R, Bartsch JW, and Jockusch H (1998) Expression of the potassium channel KV3.4 in mouse skeletal muscle parallels fiber type maturation and depends on excitation pattern. *FEBS Lett* **421**:259–262.

10. Kalman K, Nguyen A, Tseng-Crank J, Dukes ID, Chandy G, Hustad CM, Copeland NG, Jenkins NA, Mohrenweiser H, Brandriff B, et al. (1998) Genomic organization, chromosomal localization, tissue distribution, and biophysical characterization of a novel mammalian Shaker-related voltage-gated potassium channel, Kv1.7. *J Biol Chem* **273**:5851–5857.

11. Gopel SO, Kanno T, Barg S, and Rorsman P (2000) Patch-clamp characterisation of somatostatin-secreting-cells in intact mouse pancreatic islets. *J Physiol* **528**:497–507.

12. Yeung SYM, Thompson D, Wang Z, Fedida D, and Robertson B (2005) Modulation of Kv3 subfamily potassium currents by the sea anemone toxin BDS: significance for CNS and biophysical studies. *J Neurosci* **25**:8735–8745.

TABLE 16
K_v4.1 channels

Channel name	K _v 4.1
Description	Voltage-gated potassium channel, A-type potassium current
Other names	mShal ¹
Molecular information	Human: 647aa, NM_004979, chr. Xp11.23, ² <i>KCND1</i> (see 'Comments'), GeneID: 3750, PMID: 10729221 ¹² Mouse: 651aa, NM_008423, chr. X Rat: 650aa, XM_217601 (predicted), chr. Xq13
Associated subunits	KChIP1 increases K _v 4.1 current densities, accelerates inactivation time course and recovery from inactivation, and shifts steady-state inactivation to more depolarized potentials ^{3,4}
Functional assays	Patch-clamp, two-electrode voltage-clamp
Current	Somatodendritic depolarization-activated potassium currents in rat neostriatal cholinergic interneurons are predominantly of the A-type and attributable to coexpression of K _v 4.2 and K _v 4.1 subunits ⁵ ; subthreshold transient A currents in rat brain ⁶
Conductance	~6pS (main unitary conductance under physiological conditions) ^{4,7}
Ion selectivity	P _{Na} /P _K < 0.01
Activation	Voltage, V _a = -47.9 mV; k _a = 24.2 mV (assuming a fourth-order Boltzmann function) ⁷
Inactivation	V _h = -69 mV; k _h = 4.8 mV; τ _{h1} = 22 ms (20 mV); τ _{h2} = 86 ms (20 mV); τ _{h3} = 368 ms (20 mV) ⁷ (see "Comments")
Activators	None
Gating inhibitors	None
Blockers	4-Aminopyridine (9 mM) ^{1,7} , tetraethylammonium (>10 mM) ¹
Radioligands	None
Channel distribution	Fetal, infant, and adult brain; colon, heart, lung, stomach, testis, liver, kidney, thyroid gland, pancreas, pulmonary artery ⁸⁻¹⁰
Physiological functions	Not established
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	The <i>K_v4.1</i> (<i>KCND1</i>) gene is encoded by at least 6 exons ² —the first exon encodes the protein from the N terminus through S5 into the P-region, whereas the remainder of the protein is encoded by exons 2–6; kinetic properties depend on the expression system, recording configuration, and the presence of auxiliary subunits (KChIPs) ^{4,11} ; mammalian <i>Shal</i> -related family

aa, amino acids; chr., chromosome.

1. Pak MD, Baker K, Covarrubias M, Butler A, Ratcliffe A, and Salkoff L (1991) mShal, a subfamily of A-type K⁺ channel cloned from mammalian brain. *Proc Natl Acad Sci USA* **88**:4386–4390.2. Isbrandt D, Leicher T, Waldschutz R, Zhu X, Luhmann U, Michel U, Sauter K, and Pongs O (2000) Gene structures and expression profiles of three human KCND (Kv4) potassium channels mediating A-type currents I_{TO} and I_{SA}. *Genomics* **64**:144–154.3. Nakamura TY, Nandi S, Pountney DJ, Artman M, Rudy B, and Coetzee WA (2001) Different effects of the Ca²⁺-binding protein, KChIP1, on two Kv4 sub-family members, Kv4.1 and Kv4.2. *FEBS Lett* **499**:205–209.4. Beck EJ, Bowlby M, An WF, Rhodes KJ, and Covarrubias M (2002) Remodelling inactivation gating of Kv4 channels by KChIP1, a small-molecular-weight calcium-binding protein. *J Physiol* **538**:691–706.5. Song WJ, Tkatch T, Baranauskas G, Ichinohe N, Kitai ST, and Surmeier DJ (1998) Somatodendritic depolarization-activated potassium currents in rat neo-striatal cholinergic interneurons are predominantly of the A type and attributable to coexpression of Kv4.2 and Kv4.1 subunits. *J Neurosci* **18**:3124–3137.6. Serodio P and Rudy B (1998) Differential expression of Kv4 K⁺ channel subunits mediating sub-threshold transient K⁺ (A-type) currents in rat brain. *J Neurophysiol* **79**:1081–1091.7. Jerng HH, Shahidullah M, and Covarrubias M (1999) Inactivation gating of Kv4 potassium channels: molecular interactions involving the inner vestibule of the pore. *J Gen Physiol* **113**:641–660.

8. UniGeneCluster Hs0.55276; OMIM no. 300281.

9. Davies AR and Kozlowski RZ (2001) Kv channel subunit expression in rat pulmonary arteries. *Lung* **179**:147–161.10. Brahmajothi MV, Morales MJ, Liu S, Rasmuson RL, Campbell DL, and Strauss HC (1996) In situ hybridization reveals extensive diversity of K⁺ channel mRNA in isolated ferret cardiac myocytes. *Circ Res* **78**:1083–1089.11. Beck E, and Covarrubias M (2001) Preferential modulation of closed-state inactivation in Kv4 K⁺ channels. *Biophys J* **81**:867–883.12. Isbrandt D, Leicher T, Waldschutz R, Zhu X, Luhmann U, Michel U, Sauter K, and Pongs O (2000) Gene structures and expression profiles of three human KCND (Kv4) potassium channels mediating A-type currents I_{TO} and I_{SA}. *Genomics* **64**:144–154.

TABLE 17
K_v4.2 channels

Channel name	K _v 4.2
Description	Voltage-gated potassium channel, A-type potassium current
Other names	Shal1, RK5 ^{1–3}
Molecular information	Human: 630aa, NM_012281, chr. 7q31, <i>KCND2</i> (see “Comments”), GeneID: 3751, PMID: 10551270 ²⁴ Mouse: 630aa, NM_019697, chr. 6 Rat: 490aa, NM_031730, chr. 4q22
Associated subunits	Coexpression of KChIP1 results in increased current densities, slowed onset of inactivation, and accelerated recovery from inactivation ⁴ ; KChIP4/CALP interacts with K _v 4.2 and presenilin 2 ⁵ , frequenin, a calcium-binding protein, enhances K _v 4.2 current amplitudes, slows inactivation time course and accelerates recovery from inactivation ⁶ ; PSD95, a PDZ domain protein, associates with K _v 4.2 and is involved in trafficking of the channel ⁷ ; a number of proteins have been shown to interact and modify K _v 4 proteins, including KChIPs, DPPX, DPP10, frequenin, PSD95, and filamin—most of these studies have used K _v 4.2 and sometimes K _v 4.3 proteins, but it is likely that these interactions also occur with Kv4.1; the physiological role of these proteins in native channels remains to be studied in most cases
Functional assays	Patch-clamp, two-electrode voltage-clamp
Current	I _{to} current in the heart is a heteromultimer of K _v 4.2 and K _v 4.3 subunits and KChIP2 ⁸ ; I _{SA} current in somatic recordings from neurons ⁹
Conductance	Not established
Ion selectivity	P _{Na} /P _K < 0.01
Activation	Midpoint of activation = 1 mV ²
Inactivation	Rapid inactivation with time constants of 15 and 60 ms ²
Activators	None
Gating inhibitors	None
Blockers	4-Aminopyridine (5 mM), ^{1,10} heteropodatoxins, ¹¹ PaTX1,2 (2–70 nM), arachidonic acid (2 μM) ¹²
Radioligands	None
Channel distribution	Brain [cerebellum (granular cells) > hippocampus, thalamus, medial habenular nucleus > cerebral cortex; basal ganglia and forebrain ¹³ ; concentrated in dendrites and soma ¹⁴], cochlear nucleus, ¹⁵ atrium, ventricle ^{1–3,16} ; in situ hybridization has shown that many neuronal populations preferentially express K _v 4.2 or K _v 4.3 ²³ —for example, CA1 hippocampal neurons express K _v 4.2 but not K _v 4.3—on the other hand, Purkinje cells and cortical interneurons express K _v 4.3 preferentially; in cerebellar granule cells, there is a reciprocal anterior-posterior gradient of expression
Physiological functions	Repolarization of the cardiac action potential (notch phase), dampening back-propagating action potentials in CA1 hippocampal neurons
Mutations and pathophysiology	KChIP2–/– mice lack the I _{to} current and are susceptible to ventricular tachycardia ¹⁷ ; seizure activity reduces K _v 4.2 expression in the dentate granule cells of the hippocampus ¹⁸
Pharmacological significance	Not established
Comments	The <i>K_v4.2</i> (<i>KCND2</i>) gene, like <i>KCND1</i> and <i>KCND3</i> , contains six exons— however, the introns are significantly longer ¹⁹ ; kinetic properties depend on the expression system, recording configuration, and the presence of auxiliary subunits (KChIPs) ^{20,21} ; K _v 4.2 currents expressed in <i>Xenopus</i> oocytes are suppressed in response to protein kinase C activation ²² ; mammalian <i>Shal</i> -related family

aa, amino acids; chr., chromosome.

1. Roberds SL and Tamkun MM (1991) Cloning and tissue-specific expression of five voltage-gated potassium channel cDNAs expressed in rat heart. *Proc Natl Acad Sci USA* **88**:1798–1802.

2. Blair TA, Roberds SL, Tamkun MM, and Hartshorne RP (1991) Functional characterization of RK5, a voltage-gated K⁺ channel cloned from the rat cardiovascular system. *FEBS Lett* **295**:211–213.

4. Roberds SL and Tamkun MM (1991) Develo pMental expression of cloned cardiac potassium channels. *FEBS Lett* **284**:152–154.

5. Nakamura TY, Nandi S, Pountney DJ, Artman M, Rudy B, and Coetzee WA (2001) Different effects of the Ca²⁺-binding protein, KChIP1, on two Kv4 subfamily members, Kv4.1 and Kv4.2. *FEBS Lett* **499**:205–209.

5. Morohashi Y, Hatano N, Ohya S, Takikawa R, Watabiki T, Takasugi N, Imaizumi Y, Tomita T, and Iwatsubo T (2002) Molecular cloning and characterization of CALP/KChIP4, a novel EF-hand protein interacting with presenilin 2 and voltage-gated potassium channel subunit Kv4. *J Biol Chem* **277**:14965–14975.

6. Nakamura TY, Pountney DJ, Ozaita A, Nandi S, Ueda S, Rudy B, and Coetzee WA (2001) A role for frequenin, a Ca²⁺-binding protein, as a regulator of Kv4 K⁺-currents. *Proc Natl Acad Sci USA* **98**:12808–12813.

7. Wong W, Newell E, Jugloff DG, Jones OT, and Schlichter LC (2002) Cell-surface targeting and clustering interactions between heterologously expressed PSD-95 and the Shal voltage-gated potassium channel, Kv4.2. *J Biol Chem* **277**:20423–20430.

8. Guo W, Li H, Aimond F, Johns DC, Rhodes KJ, Trimmer JS, and Nerbonne JM (2002) Role of heteromultimers in the generation of myocardial transient outward K⁺ currents. *Circ Res* **90**:586–593.

9. Serodio P, Kentros C, and Rudy B (1994) Identification of molecular components of A-type channels activating at subthreshold potentials. *J Neurophysiol* **72**:1516–1529.

10. Tseng GN, Jiang M, and Yao JA (1996) Reverse use dependence of Kv4.2 blockade by 4-aminopyridine. *J Pharmacol Exp Ther* **279**:865–876.

11. Sanguinetti MC, Johnson JH, Hammerland LG, Kelbaugh PR, Volkmann RA, Saccamano NA, and Mueller AL (1997) Heteropodatoxins: peptides isolated from spider venom that block Kv4.2 potassium channels. *Mol Pharmacol* **51**:491–498.

12. Villarreal A and Schwarz TL (1996) Inhibition of the Kv4 (Shal) family of transient K⁺ currents by arachidonic acid. *J Neurosci* **16**:2522–2532.

13. Tkatch T, Baranauskas G, and Surmeier DJ (2000) Kv4.2 mRNA abundance and A-type K⁺ current amplitude are linearly related in basal ganglia and basal forebrain neurons. *J Neurosci* **20**:579–588.

14. Sheng M, Tsaur ML, Jan YN, and Jan LY (1992) Subcellular segregation of two A-type K⁺ channel proteins in rat central neurons. *Neuron* **9**:271–284.

15. Fitzakerley JL, Star KV, Rinn JL, and Elmquist BJ (2000) Expression of Shal potassium channel subunits in the adult and developing cochlear nucleus of the mouse. *Hear Res* **147**:31–45.

16. Dixon JE and McKinnon D (1994) Quantitative analysis of potassium channel mRNA expression in atrial and ventricular muscle of rats. *Circ Res* **75**:252–260.

17. Kuo HC, Cheng CF, Clark RB, Lin JJ, Lin JL, Hoshijima M, Nguyen-Tran VT, Gu Y, Ikeda Y, Chu PH, et al. (2001) A defect in the Kv channel-interacting protein 2 (KChIP2) gene leads to a complete loss of I_{to} and confers susceptibility to ventricular tachycardia. *Cell* **107**:801–813.

18. Tsaur ML, Sheng M, Lowenstein DH, Jan YN, and Jan LY (1992) Differential expression of K⁺ channel mRNAs in the rat brain and down-regulation in the hippocampus following seizures. *Neuron* **8**:1055–1067.

19. Isbrandt D, Leicher T, Waldschutz R, Zhu X, Luhmann U, Michel U, Sauter K and Pongs O (2000) Gene structures and expression profiles of three human KCND (Kv4) potassium channels mediating A-type currents I_{TO} and I_{SA}. *Genomics* **64**:144–154.
20. An WF, Bowlby MR, Betty M, Cao J, Ling HP, Mendoza G, Hinson JW, Mattsson KI, Strassle JS, Trimmer BW, et al. (2000) Modulation of A-type potassium channels by a family of calcium sensors. *Nature (Lond)* **403**:553–556.
21. Beck E and Covarrubias M (2001) Preferential modulation of closed-state inactivation in Kv4 K⁺ channels. *Biophys J* **81**:867–883.
22. Nakamura T, Coetzee WA, Vega-Saenz de Miera E, Artman M, and Rudy B (1997) Modulation of Kv4 channels, key components of rat ventricular transient K⁺ current, by PKC. *Am J Physiol* **273**:H1775–H1786.
23. Serodio P and Rudy B (1998) Differential expression of Kv4 K⁺ channel subunits mediating subthreshold transient K⁺ (A-type) currents in rat brain. *J Neurophysiol* **79**:1081–1091.
24. Zhu XR, Wulf A, Schwarz M, Isbrandt D, and Pongs O (1999) Characterization of human Kv4.2 mediating a rapidly-inactivating transient voltage-sensitive K⁺ current. *Receptors Channels* **6**:387–400.

TABLE 18
K_v4.3 channels

Channel name	K _v 4.3 ^{1–6}
Description	Voltage-gated potassium channel, A-type potassium current
Other names	None
Molecular information	Human: 655aa, NM_004980 (transcript variant 1), chr. 1p13.3, <i>KCND3</i> (see “Comments”), GeneID: 3752, PMID: 8734615 ² Mouse: 655aa, NM_019931, chr. 3 Rat: 636aa, NM_031739, chr. 2q34
Associated subunits	KChIP1 increases K _v 4.3 current densities, accelerates inactivation time course and recovery from inactivation, and shifts steady-state inactivation to more depolarized potentials; KChIP4a abolishes fast inactivation ⁷ ; expression of K _{vβ} 2 in brain increases current density and protein expression ⁸ ; KChAP acts as a chaperone for K _v 4.3 ⁹ ; K _v 4.3 may associate preferentially with DPP10 in native neurons that predominantly express this subunit ²⁰
Functional assays	Patch-clamp, two-electrode voltage-clamp
Current	I _{to} current in the heart is a heteromultimer of K _v 4.2 and K _v 4.3 subunits and KChIP2 ¹⁰
Conductance	~5pS (main unitary conductance under physiological conditions) ⁷ ; association with DPPX increases single channel conductance ²¹
Ion selectivity	P _{Na} /P _K < 0.01
Activation	Threshold for activation –30 mV, time course for activation 1.71 ms at 60 mV ¹¹
Inactivation	Time course for inactivation fit by a biexponential function; τ _{h1} = 27 ms at 60 mV, τ _{h2} = 142 ms at 60 mV ¹¹ (see “Comments”)
Activators	None
Gating inhibitors	None
Blockers	4-Aminopyridine, bupivacaine (31 μM), ¹¹ PaTX1,2, (2–70 nM), nicotine (40 nM) ¹²
Radioligands	None
Channel distribution	Heart, brain, smooth muscle ^{1–6,13,14}
Physiological functions	Repolarization of the cardiac action potential (notch phase)
Mutations and pathophysiology	K _v 4.3 mRNA levels are decreased in patients with paroxysmal atrial fibrillation ¹⁵
Pharmacological significance	Not established
Comments	The K _v 4.3 (<i>KCND3</i>) gene contains six exons analogous to those found in <i>KCND1</i> and <i>KCND2</i> and an additional exon L between exons 4 and 5—relative to <i>KCND1</i> , the introns are significantly longer; kinetic properties depend on the expression system, recording configuration, and the presence of auxiliary subunits (KChIPs) ^{16–18} ; K _v 4.3 currents expressed in <i>Xenopus</i> oocytes are suppressed in response to protein kinase C activation ¹⁹ ; mammalian <i>Shal</i> -related family

aa, amino acids; chr., chromosome.

1. Serodio P, Kentros C, and Rudy B (1994) Identification of molecular components of A-type channels activating at subthreshold potentials. *J Neurophysiol* **72**:1516–1529.
2. Serodio P, Vega-Saenz de Miera E, and Rudy B (1996) Cloning of a novel component of A-type K⁺ channels operating at subthreshold potentials with unique expression in heart and brain. *J Neurophysiol* **75**:2174–2179.
3. Dixon JE, Shi W, Wang HS, McDonald C, Yu H, Wymore RS, Cohen IS, and McKinnon D (1996) Role of the Kv4.3 K⁺ channel in ventricular muscle: a molecular correlate for the transient outward current. *Circ Res* **79**:659–668.
4. Tsaour ML, Chou CC, Shih YH, and Wang HL (1997) Cloning, expression and CNS distribution of Kv4.3, an A-type K⁺ channel α subunit. *FEBS Lett* **400**:215–220.
5. Ohya S, Tanaka M, Oku T, Asai Y, Watanabe M, Giles WR, and Imaizumi Y (1997) Molecular cloning and tissue distribution of an alternatively spliced variant of an A-type K⁺ channel α-subunit, Kv4.3 in the rat. *FEBS Lett* **420**:47–53.
6. Dilks D, Ling HP, Cockett M, Sokol P, and Numann R (1999) Cloning and expression of the human Kv4.3 potassium channel. *J Neurophysiol* **81**:1974–1977.
7. Holmqvist MH, Cao J, Hernandez-Pineda R, Jacobson MD, Carroll KI, Sung MA, Betty M, Ge P, Gilbride KJ, Brown ME, et al. (2002) Elimination of fast inactivation in Kv4 A-type potassium channels by an auxiliary subunit domain. *Proc Natl Acad Sci USA* **99**:1035–1040.
8. Yang EK, Alvira MR, Levitan ES, and Takimoto K (2001) Kvβ subunits increase expression of Kv4.3 channels by interacting with their C termini. *J Biol Chem* **276**:4839–4844.
9. Kuryshv YA, Wible BA, Gudzi TI, Ramirez AN, and Brown AM (2001) KChAP/Kvβ1.2 interactions and their effects on cardiac Kv channel expression. *Am J Physiol Cell Physiol* **281**:C290–C299.
10. Guo W, Li H, Aimond F, Johns DC, Rhodes KJ, Trimmer JS, and Nerbonne JM (2002) Role of heteromultimers in the generation of myocardial transient outward K⁺ currents. *Circ Res* **90**:586–593.
11. Franquez L, Valenzuela C, Eck J, Tamkun MM, Tamargo J, and Snyders DJ (1999) Functional expression of an inactivating potassium channel (Kv4.3) in a mammalian cell line. *Cardiovasc Res* **41**:212–219.
12. Wang H, Shi H, and Wang Z (1999) Nicotine depresses the functions of multiple cardiac potassium channels. *Life Sci* **65**:PL143–PL149.
13. Wickenden AD, Jegla TJ, Kaprielian R, and Backx PH (1999) Regional contributions of Kv1.4, Kv4.2, and Kv4.3 to transient outward K⁺ current in rat ventricle. *Am J Physiol* **276**:H1599–H1568.
14. Hoppe UC, Marban E, and Johns DC (2000) Molecular dissection of cardiac repolarization by in vivo Kv4.3 gene transfer. *J Clin Invest* **105**:1077–1084.
15. Brundel BJ, Van Gelder IC, Henning RH, Tuinenburg AE, Wietes M, Grandjean JG, Wilde AA, Van Gilst WH, and Crijns HJ (2001) Alterations in potassium channel gene expression in atria of patients with persistent and paroxysmal atrial fibrillation: differential regulation of protein and mRNA levels for K⁺ channels. *J Am Coll Cardiol* **37**:926–932.
16. An WF, Bowlby MR, Betty M, Cao J, Ling HP, Mendoza G, Hinson JW, Mattsson KI, Strassle BW, Trimmer JS, et al. (2000) Modulation of A-type potassium channels by a family of calcium sensors. *Nature (Lond)* **403**:553–556.

17. Beck EJ, Bowlby M, An WF, Rhodes KJ, and Covarrubias M (2002) Remodelling inactivation gating of Kv4 channels by KChIP1, a small-molecular-weight calcium-binding protein. *J Physiol* **538**:691–706.

18. Beck E and Covarrubias M (2001) Preferential modulation of closed-state inactivation in Kv4 K⁺ channels. *Biophys J* **81**:867–883.

19. Nakamura T, Coetzee WA, Vega-Saenz de Miera E, Artman M, and Rudy B (1997). Modulation of Kv4 channels, key components of rat ventricular transient K⁺ current, by PKC. *Am J Physiol* **273**:H1775–H1786.

20. Zagha E, Ozaita A, Chang SY, Nadal MS, Lin U, Saganich MJ, McCormack T, Akinsanya KO, Qi SY, and Rudy B (2005) DPP10 modulates Kv4-mediated A-type potassium channels. *J Biol Chem* **280**:18853–18861.

21. Rocha CA, Nadal M, Rudy B, and Covarrubias M. (2004) Inactivation gating of Kv4 K⁺ channels interacting with the dipeptidyl-aminopeptidase-like protein (DPPX), in *Proceedings of the 48th Annual Meeting of the Biophysical Society*; 2004 14–18 Feb; Baltimore, Md. Presentation 2780-Pos.

TABLE 19
K_v5.1 channels

Channel name	K _v 5.1 ^{1–4}
Description	Modifier of the K _v 2 family of channels
Other names	KH1, IK8
Molecular information	Human: 494aa, NM_002236, chr. 2p25, ⁵ <i>KCNF1</i> , GeneID: 3754, PMID: 9434767 ⁵ Mouse: 493aa, NM_201531, chr. 12 Rat: 505aa, XM_216678 (predicted), chr. 6
Associated subunits	Associates with K _v 2.1 and K _v 2.2
Functional assays	Voltage-clamp
Current	None
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Brain, heart, skeletal muscle, liver, kidney pancreas, ^{1,2,6} cardiac myocytes ⁷
Physiological functions	Modifies the gating properties of K _v 2.1 and K _v 2.2 channels
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	K _v 5.1 has no function on its own, but it has important modulatory actions on K _v 2 channels

aa, amino acids; chr., chromosome.

1. Drewe JA, Verma S, Frech G, and Joho RH (1992) Distinct spatial and temporal expression patterns of K⁺ channel mRNAs from different subfamilies. *J Neurosci* **12**:538–548.

2. Verma-Kurvari S, Border B, and Joho RH (1997) Regional and cellular expression patterns of four K⁺ channel mRNAs in the adult rat brain. *Brain Res Mol Brain Res* **46**:54–62.

3. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian *Shab* K⁺ channels. *J Biol Chem* **272**:24371–24379.

4. Kramer JW, Post MA, Brown AM, and Kirsch GE (1998) Modulation of potassium channel gating by coexpression of Kv2.1 with regulatory Kv5.1 or Kv6.1 α -subunits. *Am J Physiol* **274**:C1501–C1510.

5. Su K, Kyaw H, Fan P, Zeng Z, Shell BK, Carter KC, and Li Y (1997) Isolation, characterization, and mapping of two human potassium channels. *Biochem Biophys Res Commun* **241**:675–681.

6. UniGeneCluster Hs0.23735; OMIM no. 603787.

7. Brahmajothi MV, Morales MJ, Liu S, Rasmusson RL, Campbell DL, and Strauss HC (1996) In situ hybridization reveals extensive diversity of K⁺ channel mRNA in isolated ferret cardiac myocytes. *Circ Res* **78**:1083–1089.

TABLE 20
K_V6.1 channels

Channel name	K _V 6.1 ¹⁻⁶
Description	Modifier/silencer of K _V 2 family channels
Other names	KH2, K13
Molecular information	Human: 513aa, NM_002237, chr. 20q13, ^{6,7} <i>KCNGL1</i> , GeneID: 3755, PMID: 9434767 ⁶ Mouse: 534aa, XM_141545 (predicted), chr. 2 Rat: 514aa, XM_215951 (predicted), chr. 3
Associated subunits	Associates with K _V 2 family channels
Functional assays	Electrophysiology
Current	None
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Skeletal muscle, brain, uterus, ovary, kidney, pancreas, placenta, bone, germ cell, prostate, skin, testis, ^{6,7} cardiac myocytes (sinoatrial node) ⁸
Physiological functions	K _V 6.1 subunits when expressed alone are unable to elicit any current— however, K _V 6.1 can suppress K _V 2.1 current (less effectively than K _V 5.1), and to a lesser extent it can suppress K _V 2.2; the K _V 2.1 currents are strongly modified by K _V 6.1, which increases the time constant of activation and slows down inactivation
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	K _V 6.1 has no function on its own, but it has important modulatory actions on K _V 2 channels

aa, amino acids; chr., chromosome.

1. Drewe JA, Verma S, Frech G, and Joho RH (1992) Distinct spatial and temporal expression patterns of K⁺ channel mRNAs from different subfamilies. *J Neurosci* **12**:538–548.
2. Post MA, Kirsch GE, and Brown AM (1996) Kv2.1 and electrically silent Kv6.1 potassium channel subunits combine and express a novel current. *FEBS Lett* **399**:177–182.
3. Verma-Kurvari S, Border B, and Joho RH (1997) Regional and cellular expression patterns of four K⁺ channel mRNAs in the adult rat brain. *Brain Res Mol Brain Res* **46**:54–62.
4. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian *Shab* K⁺ channels. *J Biol Chem* **272**:24371–24379.
5. Kramer JW, Post MA, Brown AM, and Kirsch GE (1998) Modulation of potassium channel gating by coexpression of Kv2.1 with regulatory Kv5.1 or Kv6.1 α -subunits. *Am J Physiol* **274**:C1501–C1510.
6. Su K, Kyaw H, Fan P, Zeng Z, Shell BK, Carter KC, and Li Y (1997) Isolation, characterization, and mapping of two human potassium channels. *Biochem Biophys Res Commun* **241**:675–681.
7. UniGene Cluster Hs0.118695; OMIM no. *603788.
8. Brahmajothi MV, Morales MJ, Liu S, Rasmusson RL, Campbell DL, and Strauss HC (1996) In situ hybridization reveals extensive diversity of K⁺ channel mRNA in isolated ferret cardiac myocytes. *Circ Res* **78**:1083–1089.

TABLE 21
K_v6.2 channels

Channel name	K _v 6.2 ¹
Description	Modifier/silencer
Other names	None
Molecular information	Human: 466aa, NM_012283, chr. 18q22–18q23, ¹ <i>KCNQ2</i> , GeneID: 26251, PMID: 10551266 ¹ Mouse: AC145610 (genomic), chr. 18 Rat: 436aa, XM_225718 (predicted), chr. 18 Coassembles with K _v 2 family channels via the N termini ¹
Associated subunits	
Functional assays	Electrophysiology
Current	None
Conductance	Not functional on its own
Ion Selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Myocardium, fetal brain, germinal center B cells ^{1,2}
Physiological functions	Modifier/silencer, coassembles with K _v 2.1, producing K ⁺ channels with unique properties
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	K _v 6.2 has no function on its own, but it has important modulatory actions on K _v 2 channels

aa, amino acids; chr., chromosome.
1. Zhu XR, Netzer R, Bohlke K, Liu Q, and Pongs O (1999). Structural and functional characterization of Kv6.2: a new γ -subunit of voltage-gated potassium channel. *Receptors Channels* **6**:337–350.
2. UniGene Cluster Hs0.247905; OMIM no. 605696.

TABLE 22
K_v6.3 channels

Channel name	K _v 6.3 ¹
Description	Modifier/silencer
Other names	K _v 10.1
Molecular information	Human: 436aa, NM_133329, chr. 2p21, <i>KCNQ3</i> , GeneID: 170850, PMID: 11852086 ¹ Mouse: 433aa, NM_153512, chr. 17 Rat: 345aa, NM_133426, chr. 6q12 Coassembles with K _v 2.1 ¹
Associated subunits	
Functional assays	Electrophysiology
Current	None
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Whole brain (hippocampus, caudate nucleus, frontal lobe, hypothalamus, substantia nigra), spinal cord, pituitary, testis, small intestine, thymus, adrenal gland ¹
Physiological functions	Modifier/silencer, coassembles with K _v 2.1
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	K _v 6.3 has no function on its own, but it has important modulatory actions on K _v 2 channels

aa, amino acids; chr., chromosome.
1. Sano Y, Mochizuki S, Miyake A, Kitada C, Inamura K, Yokoi H, Nozawa K, Matsushime H, and Furuichi K (2002) Molecular cloning and characterization of Kv6.3, a novel modulatory subunit for voltage-gated K⁺ channel Kv2.1. *FEBS Lett* **512**:230–234.

TABLE 23
K_v6.4 channels

Channel name	K _v 6.4 ¹
Description	Modifier/silencer
Other names	None
Molecular information	Human: 519aa, NM_172347 (transcript variant 1), chr. 16q24.1, <i>KCNG4</i> , GeneID: 93107, PMID: 12060745 ¹ Mouse: 506aa, NM_025734, chr. 8, Rat: 506aa, XM_226524 (predicted), chr. 19
Associated subunits	Coassembles with K _v 2.1 ¹
Functional assays	Electrophysiology
Current	Not functional on its own
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Brain, liver, small intestine, colon ¹
Physiological functions	Regulation of membrane potential and action potential frequency by modulation of delayed rectifier potassium currents; modulates the activity of K _v 2.1 channels by causing marked changes in activation threshold and kinetics, C-type inactivation, and deactivation ¹
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	K _v 6.4 has no function on its own, but it has important modulatory actions on K _v 2 channels

aa, amino acids; chr., chromosome.

1. Ottschytch N, Raes A, Van Hoorick D, and Snyders DJ (2002) Obligatory heterotetramerization of three previously uncharacterized Kv channel-subunits identified in the human genome. *Proc Natl Acad Sci USA* **99**:7986–7991.

TABLE 24
K_v7.1 channels

Channel name	K _v 7.1
Description	Voltage-gated potassium channel, delayed rectifier
Other names	KVLQT1, ¹ slow delayed rectifier
Molecular information	Human: 676aa, NM_000218 (transcript variant 1), chr. 11p15.5, <i>KCNQ1</i> , GeneID: 3784, PMID: 8528244 ¹ Mouse: 668aa, NM_008434, chr. 7 Rat: 669aa, NM_032073, chr. 1q41
Associated subunits	KCNE1 (minK/IsK), KCNE3 [minK-related peptide 2 (MiRP2)]
Functional assays	Voltage-clamp
Current	I _{Ks} (with KCNE1), ^{2,3} I _{K,CAMP} (with KCNE3) ¹⁶
Conductance	1.8pS (KCNQ1 alone), 5pS (with KCNE1)
Ion selectivity	K ⁺
Activation	KCNQ1 alone: V _a = 12 mV, τ _a = 30, and 800 ms at +40 mV KCNQ1 + KCNE1: V _a = +8 mV, τ _a = 0.7, 1.5, and 8 s at +40 mV
Inactivation	KCNQ1 alone: V _h = +18 mV, τ _h = 130 ms at 20 mV
Activators	R-L3 (= L364373, 1 μM for KCNQ1 alone; R-L3 does not activate the KCNQ1/KCNE1 complex; the S enantiomer blocks KCNQ1) ⁴ ; mefenamic acid, niflumic acid, and 4,4'-diisothiocyanostilbene-2,2'-disulfonic acid (10–100 μM) ^{5,6}
Gating inhibitors	None
Blockers	Chromanol 293B (1 μM), ⁷ L735821 (80 nM), ⁸ mefloquine (0.88 μM), ⁹ azimilide (3 μM), ^{9,10} HMR-1556 (120 nM), XE991 (0.78 μM KCNQ1 alone; 11.1 μM KCNQ1/KCNE1), ¹¹ linopirdine (8.9 μM KCNQ1 alone)
Radioligands	None
Channel distribution	Heart, kidney, rectum, ear, germ, pancreas, lung, cochlea, placenta
Physiological functions	Repolarization of cardiac action potentials (KCNQ1 and minK/ISK/KCNE1 coassemble to form the cardiac I _{Ks} channel); potassium recycling at basolateral membrane of intestinal crypt cells (with KCNE3) and inner ear
Mutations and pathophysiology	Loss of function mutations in the <i>KCNQ1</i> gene can cause either RWS (autosomal dominant) or JLNS (autosomal recessive); RWS is characterized by congenital long QT syndrome and electrocardiographically distinguished by a prolonged QT interval and polymorphic ventricular arrhythmias (torsade de pointes), which may result in recurrent syncope, seizure, or sudden death; JLNS patients have deafness, congenital and functional heart disease, a prolonged QT interval on an electrocardiogram, and sudden death cardioauditory syndrome; <i>KCNQ1</i> is disrupted by chromosomal rearrangements in patients with Beckwith-Wiedemann syndrome, ¹³ as well as by a balanced chromosomal translocation in an embryonal rhabdoid tumor; gain-of-function mutations in <i>KCNQ1</i> cause atrial fibrillation and short QT syndrome
Pharmacological significance	Blockers developed as class III antiarrhythmic agents to target ventricular arrhythmias ^{14,15} ; activators could be useful for the treatment of some long QT syndromes ⁶

aa, amino acids; chr., chromosome; RWS, Romano-Ward syndrome; JLNS, Jervell and Lange-Nielsen syndrome; L735821, 3-(2,4-dichlorophenyl)-N-(6-methyl-5-oxo-2-phenyl-3,6-diazabicyclo[5.4.0]undeca-2,7,9,11-tetraen-4-yl)-prop-2-enamide; XE991, 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; HMR-1556, N-(6-cyano-3-hydroxy-2,2-dimethyl-chroman-4-yl)-N-methyl-ethanesulfonamide.

1. Wang Q, Curran ME, Splawski I, Burn TC, Millholland JM, Van Raay TJ, Shen J, Timothy KW, Vincent GM, de Jager T, et al. (1996) Positional cloning of a novel potassium channel gene: KVLQT1 mutations cause cardiac arrhythmias. *Nat Genet* 12:17–23.

2. Sanguinetti MC, Curran ME, Zou A, Shen J, Spector PS, Atkinson DL, and Keating MT (1996) Coassembly of K_v LQT1 and minK (IsK) proteins to form cardiac I_{Ks} potassium channel. *Nature (Lond)* 384:80–83.

3. Barhanin J, Lesage F, Guillemare E, Fink M, Lazdunski M, and Romey G (1996) K_v LQT1 and IsK (minK) proteins associate to form the I_{Ks} cardiac potassium current. *Nature (Lond)* 384:78–80.

4. Salata JJ, Jurkiewicz NK, Wang J, Evans BE, Orme HT, and Sanguinetti MC (1998) A novel benzodiazepine that activates cardiac slow delayed rectifier K⁺ currents. *Mol Pharmacol* 54:220–230.

5. Busch AE, Herzer T, Wagner CA, Schmidt F, Raber G, Waldegger S, and Lang F (1994) Positive regulation by chloride channel blockers of IsK channels expressed in *Xenopus* oocytes. *Mol Pharmacol* 46:750–753.

6. Abitbol I, Peretz A, Lerche C, Busch AE, and Attali B (1999) Stilbenes and fenamates rescue the loss of I_{Ks} channel function induced by an LQT5 mutation and other IsK mutants. *EMBO J* 18:4137–4148.

7. Yang IC, Scherz MW, Bahinski A, Bennett PB, and Murray KT (2000) Stereoselective interactions of the enantiomers of chromanol 293B with human voltage-gated potassium channels. *J Pharmacol Exp Ther* 294:955–962.

8. Tinel N, Lauritzen I, Chouabe C, Lazdunski M, and Borsotto M (1998) The KCNQ2 potassium channel: splice variants, functional and developmental expression: brain localization and comparison with KCNQ3. *FEBS Lett* 438:171–176.

9. Kang J, Chen XL, Wang L, and Rampe D (2001) Interactions of the antimalarial drug mefloquine with the human cardiac potassium channels KvLQT1/minK and HERG. *J Pharmacol Exp Ther* 299:290–296.

10. Busch AE, Busch GL, Ford E, Suessbrich H, Lang HJ, Greger R, Kunzelmann K, Attali B, and Stuhmer W (1997) The role of the IsK protein in the specific pharmacological properties of the I_{Ks} channel complex. *Br J Pharmacol* 122:187–189.

11. Wang HS, Brown BS, McKinnon D, and Cohen, IS (2000) Molecular basis for differential sensitivity of KCNQ and I_{Ks} channels to the cognitive enhancer XE991. *Mol Pharmacol* 57:1218–1223.

12. Keating MT and Sanguinetti MC (2001) Molecular and cellular mechanisms of cardiac arrhythmias. *Cell* 104:569–580.

13. Lee MP, Hu RJ, Johnson LA, and Feinberg AP (1997) Human KVLQT1 gene shows tissue-specific imprinting and encompasses Beckwith-Wiedemann syndrome chromosomal rearrangements. *Nat Genetics* 15:181–185.

14. Coghlan MJ, Carroll WA, and Gopalakrishnan M (2001) Recent developments in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. *J Med Chem* 44:1627–1653.

15. Shieh CC, Coghlan M, Sullivan JP, and Gopalakrishnan M (2000) Potassium channels: molecular defects, diseases, and therapeutic opportunities. *Pharmacol Rev* 52:557–594.

16. Schroeder BC, Waldegger S, Fehr S, Bleich M, Warth R, Greger R, and Jentsch TJ (2000) A constitutively open potassium channel formed by KCNQ1 and KCNE3. *Nature (Lond)* 403:196–199.

TABLE 25
K_v7.2 channels

Channel name	K _v 7.2
Description	Voltage-gated potassium channel, delayed rectifier
Other names	KQT2
Molecular information	Human: 872aa, NM_172107 (transcript variant 1), chr. 20q13.3, <i>KCNQ2</i> , GeneID: 3785, PMID: 9836639 ¹ Mouse: 870aa, NM_010611 (transcript variant 1), chr. 2 Rat: 852aa, NM_133322, chr. 3q43
Associated subunits	KCNQ3, KCNE2
Functional assays	Voltage-clamp
Current	M current
Conductance	5.8pS ¹³
Ion selectivity	K ⁺
Activation	V _a = 26 mV, τ _a = 157 ms at +30 mV
Inactivation	V _h = 18 mV, τ _h = 130 ms at 20 mV
Activators	Retigabine (10 μM), ² BMS204352 (1 μM) ³
Gating inhibitors	None
Blockers	Tetraethylammonium (KCNQ2 alone: 0.16 mM; KCNQ2/KCNQ3: 0.5 mM), ¹ XE991 (0.7 μM), ^{1,4} linopiridine (4.8 μM), ^{1,3} L735821 (1.5 μM) ⁵
Radioligands	None
Channel distribution	Infant brain, adult brain, fetal brain, sympathetic ganglia, lung, testis, fetal heart, adult heart, breast, eye, germ cell, placenta, small intestine, neuroblastoma ¹⁰
Physiological functions	Determines subthreshold excitability of neurons; KCNQ2 and KCNQ3 coassemble to form the M current in the brain ¹ (see "Comments"); KCNQ2 and KCNQ3 proteins are colocalized in a somatodendritic pattern on pyramidal and polymorphic neurons in the human cortex and hippocampus ¹¹ ; KCNQ2 is also expressed in the absence of KCNQ3 in some presynaptic terminals ¹¹
Mutations and pathophysiology	Benign familial neonatal convulsions (<i>EBN1</i>) with myokymia ^{6,7} ; in KCNQ2 knockout mice, homozygotes (KCNQ2 ^{−/−}) die within a few hours after birth owing to pulmonary atelectasis that is not due to the status of epileptic seizures, although their development is morphologically normal; heterozygous mice have decreased expression of KCNQ2 and show hypersensitivity to pentylenetetrazole, an inducer of seizure ¹²
Pharmacological significance	Retigabine is an anticonvulsant ² (the M current is a new target for antiepileptic therapy ^{8,9}); blockers enhance learning and memory in animal models ⁹
Comments	The M current is a slowly activating and deactivating potassium conductance that plays a critical role in determining the subthreshold excitability of neurons as well as the responsiveness to synaptic inputs; the M current was first described in peripheral sympathetic neurons, and differential expression of this conductance produces subtypes of sympathetic neurons with distinct firing patterns; the M current is also expressed in many neurons in the central nervous system

aa, amino acids; chr., chromosome; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1*H*-indol-2-one; XE991, 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; L735821, 3-(2,4-dichlorophenyl)-*N*-(6-methyl-5-oxo-2-phenyl-3,6-diazabicyclo[5.4.0]undeca-2,7,9,11-tetraen-4-yl)-prop-2-enamide.

1. Wang HS, Pan Z, Shi W, Brown BS, Wymore RS, Cohen IS, Dixon JE, and McKinnon D (1998). KCNQ2 and KCNQ3 potassium channel subunits: molecular correlates of the M-channel. *Science (Wash DC)* **282**:1890–1893.

2. Tatulian L, Delmas P, Abogadie FC, and Brown DA (2001). Activation of expressed KCNQ potassium currents and native neuronal M-type potassium currents by the anti-convulsant drug retigabine. *J Neurosci* **21**:5535–5545.

3. Schroder RL, Jespersen T, Christophersen P, Strobaek D, Jensen BS, Olesen SP (2001) KCNQ4 channel activation by BMS-204352 and retigabine. *Neuropharmacology* **40**:888–898.

4. Robbins J (2001) KCNQ potassium channels: physiology, pathophysiology, and pharmacology. *Pharmacol Ther* **90**:1–19.

5. Tinel N, Lauritzen I, Chouabe C, Lazdunski M, and Borsotto M (1998) The KCNQ2 potassium channel: splice variants, functional and developmental expression: brain localization and comparison with KCNQ3. *FEBS Lett* **438**:171–176.

6. Charlier C, Singh NA, Ryan SG, Lewis TB, Reus BE, Leach R, and Leppert M. (1998) A pore mutation in a novel KQT-like potassium channel gene in an idiopathic epilepsy family. *Nat Genet* **18**:53–55.

7. Biervert C, Schroeder BC, Kubisch C, Berkovic CF, Propping P, Jentsch TJ, and Steinlein OK (1998) A potassium channel mutation in neonatal human epilepsy. *Science (Wash DC)* **279**:403–406.

8. Cooper EC (2001) Potassium channels: how genetic studies of epileptic syndromes open paths to new therapeutic targets and drugs. *Epilepsia* **42**:49–54.

9. Coghlan MJ, Carroll WA, and Gopalakrishnan M (2001) Recent developments in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. *J Med Chem* **44**:1627–1653.

10. Smith JS, Iannotti C, Dargis P, Christian EP, and Aiyar J (2001) Differential expression of KCNQ2 splice variants: implications to M current function during neuronal development. *J Neurosci* **21**:1096–1103.

11. Cooper EC, Aldape KD, Abosch A, Barbaro NM, Berger MS, Peacock WS, Jan YN, and Jan LY (2000) Colocalization and coassembly of two human brain M-type potassium channel subunits that are mutated in epilepsy. *Proc Natl Acad Sci USA* **97**:4914–4919.

12. Watanabe H, Nagata E, Kosaki A, Nakamura M, Yokoyama M, Tanaka K, and Sasai H (2000) Disruption of the epilepsy KCNQ2 gene results in neural hyperexcitability. *J Neurochem* **75**:28–33.

13. Selyanko AA, Hadley JK, Wood IC, Abogadie FC, Delmas P, Buckley NJ, London B, and Brown DA (2001) Properties of single M-type KCNQ2/KCNQ3 potassium channels expressed in mammalian cells. *J Physiol* **534**:15–24.

TABLE 26
K_v7.3 channels

Channel name	K _v 7.3
Description	Voltage-gated potassium channel, delayed rectifier
Other names	None
Molecular information	Human: 872aa NM_004519, chr. 8q24, <i>KCNQ3</i> , GeneID: 3786, PMID: 9836639 ¹ Mouse: 873aa, NM_152923, chr. 15 Rat: 873aa, NM_031597, chr. 7q33
Associated subunits	KCNQ2, KCNQ5
Functional assays	Voltage-clamp
Current	M current ¹
Conductance	7.3pS
Ion selectivity	K ⁺
Activation	V _a = 39 mV, τ _a = 60 ms at +30 mV
Inactivation	Not established
Activators	Retigabine (<i>KCNQ3</i> alone: 0.6 μM; <i>KCNQ3/KCNQ5</i> : 1.4 μM) ² ; XE991, ³ BMS204352 (1 μM) ⁴
Gating inhibitors	None
Blockers	Tetraethylammonium (>30 mM), ⁵ linopiridine (<i>KCNQ3/KCNQ5</i> : 7.7 μM) ²
Radioligands	None
Channel distribution	Brain, testis, retina, colon, eye, head, neck
Physiological functions	Determines subthreshold excitability of neurons; <i>KCNQ2</i> and <i>KCNQ3</i> coassemble to form the M current in the brain ¹ (see “Comments”); <i>KCNQ2</i> and <i>KCNQ3</i> proteins are colocalized in a somatodendritic pattern on pyramidal and polymorphic neurons in the human cortex and hippocampus ^{7,8}
Mutations and pathophysiology	Benign familial neonatal convulsions (<i>EBN2</i>) (e.g., G263V mutation in the pore) ⁹
Pharmacological significance	Anticonvulsants (activators), cognition enhancers (blockers) ⁶
Comments	The M current is a slowly activating and deactivating potassium conductance that plays a critical role in determining the subthreshold excitability of neurons as well as the responsiveness to synaptic inputs; the M current was first described in peripheral sympathetic neurons, and differential expression of this conductance produces subtypes of sympathetic neurons with distinct firing patterns; the M current is also expressed in many neurons in the central nervous system

aa, amino acids; chr., chromosome; XE991 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1*H*-indol-2-one.

1. Wang HS, Pan Z, Shi W, Brown BS, Wymore RS, Cohen IS, Dixon JE, and McKinnon D (1998) *KCNQ2* and *KCNQ3* potassium channel subunits: molecular correlates of the M-channel. *Science (Wash DC)* **282**:1890–1893.

2. Wickenden AD, Zou A, Wagoner PK, and Jegla T (2001) Characterization of *KCNQ5/Q3* potassium channels expressed in mammalian cells. *Br. J. Pharmacol* **132**:381–384.

3. Wang HS, Brown BS, McKinnon D, and Cohen IS (2000) Molecular basis for differential sensitivity of *KCNQ*, and *I_{Ks}* channels to the cognitive enhancer XE991. *Mol Pharmacol* **57**:1218–1223.

4. Schroder RL, Jespersen T, Christophersen P, Strobaek D, Jensen BS, and Olesen SP (2001) *KCNQ4* channel activation by BMS-204352 and retigabine. *Neuropharmacology* **40**:888–898.

5. Hadley JK, Noda M, Selyanko AA, Wood IC, Abogadie FC, and Brown DA (2000) Differential tetraethylammonium sensitivity of *KCNQ1–4* potassium channels. *Br J Pharmacol* **129**:413–415.

6. Coghlan MJ, Carroll WA, and Gopalakrishnan M (2001) Recent developments in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. *J Med Chem* **44**:1627–1653.

7. Smith JS, Iannotti C, Dargis P, Christian EP, and Aiyar J (2001) Differential expression of *KCNQ2* splice variants: implications to M current function during neuronal development. *J Neurosci* **21**:1096–1103.

8. Cooper EC, Aldape KD, Abosch A, Barbaro NM, Berger MS, Peacock WS, Jan YN, and Jan LY (2000) Colocalization and coassembly of two human brain M-type potassium channel subunits that are mutated in epilepsy. *Proc Natl Acad Sci USA* **97**:4914–4919.

9. Charlier C, Singh NA, Ryan SG, Lewis TB, Reus BE, Leach RJ, and Leppert M (1998) A pore mutation in a novel KQT-like potassium channel gene in an idiopathic epilepsy family. *Nat Genet* **18**:53–55.

TABLE 27
K_v7.4 channels

Channel name	K _v 7.4
Description	Voltage-gated potassium channel, delayed rectifier
Other names	None
Molecular information	Human: 695aa, NM_004700 (transcript variant 1), chr. 1p34, <i>KCNQ4</i> , ¹ GeneID: 9132, PMID: 10025409 ¹ Mouse: 724aa, XM_143960 (predicted), chr. 4 Rat: AF249748 (partial coding sequence)
Associated subunits	KCNQ3 ²
Functional assays	Voltage-clamp
Current	IK,n
Conductance	Not established
Ion selectivity	K ⁺
Activation	V _a = 10 mV
Inactivation	Not established
Activators	Retigabine (1 μM) ³ ; BMS204352 (1 μM) ³
Gating inhibitors	None
Blockers	Tetraethylammonium (3 mM), ⁴ linopirdine (14 μM), ⁵ XE991 (5 μM), ⁵ bepridil (9.4 μM) ⁵
Radioligands	None
Channel distribution	Cochlea (outer hair cells), placenta, vestibular organs (type 1 hair cells), brainstem auditory nuclei
Physiological functions	Mediates potassium efflux from outer hair cells ^{1,6}
Mutations and pathophysiology	Mutations in <i>KCNQ4</i> cause autosomal dominant nonsyndromic deafness type 2 (DFNA2) ^{1,6}
Pharmacological significance	Anticonvulsants (activators)

aa, amino acids; chr., chromosome; XE991, 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1H-indol-2-one.

1. Kubisch C, Schroeder BC, Friedrich T, Lütjohann B, El-Amraoui A, Marlin S, Petit C, and Jentsch TJ (1999) KCNQ4, a novel potassium channel expressed in sensory outer hair cells, is mutated in dominant deafness. *Cell* **96**:437–446.

2. Schroeder BC, Waldegger S, Fehr S, Bleich M, Warth R, Greger R, and Jentsch TJ (2000) A constitutively open potassium channel formed by KCNQ1 and KCNE3. *Nature (Lond)* **403**:196–199.

3. Schroder RL, Jespersen T, Christophersen P, Strobaek D, Jensen BS, and Olesen SP (2001) KCNQ4 channel activation by BMS-204352 and retigabine. *Neuropharmacology* **40**:888–898.

4. Hadley JK, Noda M, Selyanko AA, Wood IC, Abogadie FC, and Brown DA (2000) Differential tetraethylammonium sensitivity of KCNQ1–4 potassium channels. *Br J Pharmacol* **129**:413–415.

5. Sogaard R, Ljungstrom T, Pedersen KA, Olesen SP, and Jensen BS (2001) KCNQ4 channels expressed in mammalian cells: functional characteristics and pharmacology. *Am J Physiol* **280**:C859–C866.

6. Kharkovets T, Hardelin JP, Safieddine S, Schweizer M, El-Amraoui A, Petit C, and Jentsch TJ (2000) KCNQ4, a K⁺ channel mutated in a form of dominant deafness, is expressed in the inner ear and the central auditory pathway. *Proc Natl Acad Sci USA* **97**:4333–4338.

TABLE 28
K_v7.5 channels

Channel name	K _v 7.5
Description	Voltage-gated potassium channel, delayed rectifier
Other names	None
Molecular information	Human: 932aa, NM_019842, chr. 6q14, <i>KCNQ</i> , ^{1,5} GeneID: 56479, PMID: 10787416 ¹ Mouse: 933aa, NM_023872, chr. 1 Rat: 953aa, XM_237012 (predicted), chr. 9
Associated subunits	KCNQ3
Functional assays	Voltage-clamp
Current	M current ¹
Conductance	Not established
Ion selectivity	K ⁺
Activation	V _a = 30 mV
Inactivation	Not established
Activator	Retigabine (KCNQ5/KCNQ3: 1.4 μM), ² BMS204352 (2.4 μM) ³
Gating inhibitors	None
Blockers	Tetraethylammonium (>30 mM), ¹ linopiridine (16 μM), ¹ linopiridine KCNQ5/KCNQ3 (7.7 μM), ² XE991 ³
Radioligands	None
Channel distribution	Brain, sympathetic ganglia (splice variant I), ⁴ skeletal muscle (splice variant III) ⁴
Physiological functions	Determines subthreshold excitability of neurons
Mutations and pathophysiology	A number of allelic variants have been identified
Pharmacological significance	Anticonvulsants (activators)

aa, amino acids; chr., chromosome; XE991, 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1H-indol-2-one.

1. Lerche C, Scherer CR, Seebach G, Derst C, Wei AD, Busch AE, and Steinmeyer K (2000) Molecular cloning and functional expression of KCNQ5, a potassium channel subunit that may contribute to neuronal M-current diversity. *J Biol Chem* **275**:22395–22400.

2. Wickenden AD, Zou A, Wagoner PK, and Jegla T (2001) Characterization of KCNQ5/Q3 potassium channels expressed in mammalian cells. *Br J Pharmacol* **132**:381–384.

3. Dupuis DS, Schroder RL, Jespersen T, Christensen JK, Christophersen P, Jensen BS, and Olesen SP (2002) Activation of KCNQ5 channels stably expressed in HEK293 cells by BMS-204352. *Eur J Pharmacol* **437**:129–137.

4. Schroeder BC, Hechenberger M, Weinreich F, Kubisch C, and Jentsch TJ (2000) KCNQ5, a novel potassium channel broadly expressed in brain, mediates M-type currents. *J Biol Chem* **275**:24089–24095.

TABLE 29
Kv8.1 channels

Channel name	Kv8.1 ¹⁻³
Description	Modifier/silencer
Other names	Kv2.3, HNKA
Molecular information	Human: 500aa, NM_014379, chr. 8q22.3-24.1, <i>KCNV1</i> , GeneID: 27012, PMID: 670833 ¹ Mouse: 503aa, NM_026200, chr. 15 Rat: 503aa, NM_021697, chr. 7q31
Associated subunits	Coassembles with Kv2 family channels
Functional assays	Voltage-clamp
Current	None established
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Infant brain, adult brain (layers II, IV, and VI of the cerebral cortex, hippocampus, CA1–CA4 pyramidal cell layer, granule cells of the dentate gyrus, granule cell layer, Purkinje cell layer of the cerebellum), kidney
Physiological functions	Regulation of membrane potential and action potential frequency by modulation of delayed rectifier potassium current; modulates the activity of Kv2.1 and Kv2.2 channels by changing kinetics and levels of expression and by shifting the half-inactivation potential to more polarized values
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	Kv8.1 has no function on its own, but it has important modulatory actions on Kv2 channels

aa, amino acids; chr., chromosome.
1. Hugnot JP, Salinas M, Lesage F, Guillemare E, de Weille J, Heurteaux C, Mattei MG, and Lazdunski M (1996) Kv8.1, a new neuronal potassium channel subunit with specific inhibitory properties towards Shab and Shaw channels. *EMBO J* **15**:3322–3331.
2. Salinas M, de Weille J, Guillemare E, Lazdunski M, and Hugnot JP (1997) Modes of regulation of *Shab* K⁺ channel activity by the Kv8.1 subunit. *J Biol Chem* **272**:8774–8780.
3. Chiara MD, Monje F, Castellano A, and Lopez-Barneo J (1999) A small domain in the N terminus of the regulatory α -subunit Kv2.3 modulates Kv2.1 potassium channel gating. *J Neurosci* **19**:6865–6873.

TABLE 30
Kv8.2 channels

Channel name	Kv8.2
Description	Modifier/silencer
Other names	Kv11.1 ¹
Molecular information	Human: 545aa, NM_133497, chr. 9p24.2, <i>KCNV2</i> , GeneID: 169522, PMID: 12060745 ¹ Mouse: 562aa, NM_183179, chr. 19 Rat: 561 aa, XM_220024 (predicted), chr. 1
Associated subunits	Coassembles with Kv2 family channels
Functional assays	Voltage-clamp
Current	None established
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Lung, liver, kidney, pancreas, spleen, thymus, prostate, testis, ovary, colon ¹
Physiological functions	Regulation of membrane potential and action potential frequency by modulation of delayed rectifier potassium currents; modulates the activity of Kv2.1 channels by causing small changes in activation threshold and kinetics and in C-type inactivation ¹
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	Kv8.2 has no function on its own, but it has important modulatory actions on Kv2 channels

aa, amino acids; chr., chromosome.
1. Ottschytch N, Raes A, Van Hoorick D, and Snyder DJ (2002) Obligatory heterotetramerization of three previously uncharacterized Kv channel-subunits identified in the human genome. *Proc Natl Acad Sci USA* **99**:7986–7991.

TABLE 31
K_v9.1 channels

Channel name	K _v 9.1 ¹⁻⁴
Description	Modifier/silencer
Other names	None
Molecular information	Human: 526 aa, NM_002251, chr. 20q12, <i>KCNS1</i> , GeneID: 3787, PMID: 10484328 ³ Mouse: 497 aa, NM_008435, chr. 2 Rat: 497 aa, NM_053954, chr. 3q42
Associated subunits	Coassembles with K _v 2 family channels
Functional assays	Voltage-clamp
Current	None established
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Infant brain, adult brain (frontal cortex), lens epithelium, melanocytes (in mouse brain, the distribution of K _v 9.1 is similar to K _v 9.2, with highest expression levels in the main olfactory bulb, cerebral cortex, hippocampal formation, habenula, basolateral amygdaloid nuclei, and cerebellum; K _v 9.1 and K _v 9.2 are colocalized with K _v 2.1 and/or K _v 2.2 α subunits in several regions)
Physiological functions	Regulation of membrane potential and action potential frequency by modulation of delayed rectifier potassium current; modulates the activity of K _v 2.1 and K _v 2.2 α subunits by changing kinetics and levels of expression and by shifting the half-inactivation potential to more polarised values; K _v 9.1 enhances the single-channel conductance of K _v 2.1
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	The human K _v 9.1 gene is composed of a minimum of 5 exons, with at least 2 alternatively spliced exons in the 5'-untranslated region ³

aa, amino acids; chr., chromosome.

1. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian *Shab* K⁺ channels. *J Biol Chem* **272**:24371–24379.
2. Stocker M and Kerschensteiner D (1998) Cloning and tissue distribution of two new potassium channel α -subunits from rat brain. *Biochem Biophys Res Commun* **248**:927–934.
3. Shepard AR and Rae JL (1999) Electrically silent potassium channel subunits from human lens epithelium. *Am J Physiol* **277**:C412–C424.
4. Richardson FC and Kaczmarek LK (2000) Modification of delayed rectifier potassium currents by the Kv9.1 potassium channel subunit. *Hear Res* **147**:21–30.

TABLE 32
K_v9.2 channels

Channel name	K _v 9.2 ^{1,2}
Description	Modifier/silencer
Other names	None
Molecular information	Human: 477aa, NM_020697, chr. 8q22, ³ <i>KCNS2</i> , GeneID: 3788, PMID: 9305895 ¹ Mouse: 477aa, NM_181317, chr. 15 Rat: 477 aa, NM_023966, chr. 7q22
Associated subunits	Coassembles with K _v 2 family channels
Functional assays	Voltage-clamp
Current	None established
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Infant and adult brain, retina, spinal cord (in mouse brain, the distribution of K _v 9.2 is similar to K _v 9.1, with highest expression levels in the main olfactory bulb, cerebral cortex, hippocampal formation, habenula, basolateral amygdaloid nuclei, and cerebellum; K _v 9.1 and K _v 9.2 are colocalized with K _v 2.1 and/or K _v 2.2 α subunits in several regions; also found in the retina, spinal cord, and pulmonary artery)
Physiological functions	Regulation of membrane potential and action potential frequency by modulation of delayed rectifier potassium current; modulates the activity of K _v 2.1 and K _v 2.2 α subunits by changing kinetics and levels of expression and by shifting the half-inactivation potential to more polarized values; K _v 9.1 enhances the single-channel conductance of K _v 2.1
Mutations and pathophysiology	Not established
Pharmacological significance	Not established

aa, amino acids; chr., chromosome.

1. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian *Shab* K⁺ channels. *J Biol Chem* **272**:24371–24379.
2. Davies AR and Kozlowski RZ (2001) Kv channel subunit expression in rat pulmonary arteries. *Lung* **179**:147–161.
3. Banfi S, Borsani G, Rossi E, Bernard L, Guffanti A, Rubboli F, Marchitelli A, Giglio S, Coluccia E, Zollo M, et al. (1996) Identification and mapping of human cDNAs homologous to *Drosophila* mutant genes through EST database searching. *Nat Genet* **13**:167–174.

TABLE 33
K_v9.3 channels

Channel name	K _v 9.3 ^{1–3}
Description	Modifier/silencer
Other names	None
Molecular information	Human: 491aa, NM_023966, NM_002252, chr. 2p24, <i>KCNS3</i> (see 'Comments'), GeneID: 3790, PMID: 9362476 ¹ Mouse: 491aa, NM_173417, chr. 12 Rat: 491aa, NM_031778, chr. 6q14
Associated subunits	Coassembles with K _v 2 family channels
Functional assays	Voltage-clamp
Current	K _v 9.3/K _v 2.1 and ATP-dependent delayed rectifier channel in oxygen-sensitive pulmonary myocytes
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	K _v 9.3/K _v 2.1 heteromers inactivate in a fast and complete fashion from intermediate closed states but in a slow and incomplete manner from open states ⁴
Activators	None
Gating inhibitors	None
Blockers	Hypoxia blocks K _v 9.3/K _v 2.1 channels ⁵
Radioligands	None
Channel distribution	Brain, breast, colon, eye, lens, heart, kidney, muscle, lung, testis, skin, stomach, uterus ⁶ ; also found in lens epithelium ³
Physiological functions	Regulation of membrane potential in pulmonary artery myocytes
Mutations and pathophysiology	Not established
Pharmacological significance	Pulmonary artery hypertension
Comments	The human K _v 9.3 gene is intronless across the coding region 3'-UTR and all of the analysed 5'-UTR

aa, amino acids; chr., chromosome; UTR, untranslated region.
1. Patel AJ, Lazdunski M, and Honore E (1997) Kv2.1/Kv9.3, a novel ATP-dependent delayed-rectifier K⁺ channel in oxygen-sensitive pulmonary artery myocytes. *EMBO J* **16**:6615–6625.
2. Stocker M and Kerschensteiner D (1998) Cloning and tissue distribution of two new potassium channel α -subunits from rat brain. *Biochem Biophys Res Commun* **248**:927–934.
3. Shepard AR and Raem JL (1999) Electrically silent potassium channel subunits from human lens epithelium. *Am J Physiol* **277**, C412–C424.
4. Kerschensteiner D and Stocker M (1999) Heteromeric assembly of Kv2.1 with Kv9.3: effect on the state dependence of inactivation. *Biophys J* **77**:248–257.
5. Hulme JT, Coppock EA, Felipe A, Martens JR, and Tamkun MM (1999) Oxygen sensitivity of cloned voltage-gated K⁺ channels expressed in the pulmonary vasculature. *Circ Res* **85**:489–497.
6. UniGeneCluster Hs0.47584; OMIM no. 603888.

TABLE 34
K_v10.1 channels

Channel name	K _v 10.1
Description	Voltage-gated potassium channel, delayed rectifier
Other names	eag1a, eag1b, KCNH1a, KCNH1b, <i>ether-à-go-go</i> ¹⁻⁴
Molecular information	Human: 989aa, NM_172362, chr. 1q32-41, <i>KCNH1</i> (see "Comments"), GeneID: 3756, PMID: 8159766 ² Mouse: 989aa, NM_010600, chr. 1 Rat: 962aa, NM_031742, chr. 13q27
Associated subunits	Hyperkinetic (Hk), ⁵ CaM, ⁶ Slob, ⁷ epsin, ⁸ KCR1 (K channel regulator) ⁹
Functional assays	Voltage-clamp
Current	Delayed rectifier
Conductance	Not established
Ion selectivity	K ⁺ and Ca, ^{2+ 10} variable Cs ⁺
Activation	Extracellular Mg ²⁺ and other divalent cations slow activation in a dose- and voltage-dependent manner, based on their enthalpy of hydration ¹¹ ; low external pH also slows activation
Inactivation	Not established
Activators	Hyperpolarization slows down the kinetics of activation; depolarization accelerates the kinetics of activation ³
Gating inhibitors	None
Blockers	Quinidine (1.4 μM), ¹² calcium/calmodulin (480 nM) ^{6,13}
Radioligands	None
Channel distribution	Brain (amygdala, caudate nucleus, cerebral cortex, cerebellum, putamen, hippocampus, frontal lobe, occipital lobe, temporal lobe, subthalamic nucleus; not in substantia nigra, thalamus, or medulla oblongata), myoblasts, skeletal muscle (ESTs, but not detected by Northern), melanoma cells, ectopic expression in cancer cell lines and many tumor cells from different tissues, spiral ligament in rat ¹⁴⁻¹⁶
Physiological functions	Role in controlling the cell cycle and/or cell proliferation ^{17,18} ; eag-1 is thought to encode the noninactivating delayed rectifier potassium channel K _{NI} that is activated at the onset of human myoblast differentiation ⁴
Mutations and pathophysiology	K _v 10.1 has been associated with human cervical carcinoma ²¹
Pharmacological significance	K _v 10.1 blockers might have use in cancer therapy
Comments	This channel has a GFG (rather than the common GYG) potassium channel signature sequence, a PAS domain in the distal part of the cytosolic N terminus, a cNBD domain in the proximal portion of the C terminus, a C-terminal assembly domain (CAD), a CaM-binding domain, a bNLS domain in the C terminus, and a C-terminal domain required for assembly ¹⁹ ; the TCC domain at the C-terminal end of K _v 10 and K _v 11 confers specificity for multimer formation, allowing K _v 10.1/K _v 10.2 heteromerization and K _v 11.1 homomerization but not K _v 10.x/K _v 11.1 heteromerization ²² ; this C-terminal TCC domain has been identified in many other channels, and mutations of the TCC have been found to be linked to genetic channelopathies; conductance properties have been shown to change with the cell cycle ²⁰

aa, amino acids; chr., chromosome; CaM, calmodulin; TCC, tetramerizing coiled-coiled; EST, expressed sequence tag.

- Warmke J, Drysdale R, and Ganetzky B (1991) A distinct potassium channel polypeptide encoded by the *Drosophila* eag locus. *Science (Wash DC)* **252**:1560–1562.
- Warmke JW and Ganetzky B (1994) A family of potassium channel genes related to eag in *Drosophila* and mammals. *Proc Natl Acad Sci USA* **91**:3438–3442.
- Ludwig J, Terlau H, Wunder F, Bruggemann A, Pardo LA, Marquardt A, Stuhmer W, and Pongs O (1994) Functional expression of a rat homologue of the voltage gated ether à go-go potassium channel reveals differences in selectivity and activation kinetics between the *Drosophila* channel and its mammalian counterpart. *EMBO J* **13**:4451–4458.
- Ochiodoro T, Bernheim L, Liu JH, Bijlenga P, Sinnreich M, Bader CR, and Fischer-Lougheed J (1998) Cloning of a human ether-à-go-go potassium channel expressed in myoblasts at the onset of fusion. *FEBS Lett* **434**:177–182.
- Wilson GF, Wang Z, Chouinard SW, Griffith LC, and Ganetzky B (1998) Interaction of the K channel β subunit, Hyperkinetic, with eag family members. *J Biol Chem* **273**:6389–6394.
- Schönherr R, Lober K, and Heinemann SH (2000) Inhibition of human ether à go-go potassium channels by Ca²⁺/calmodulin. *EMBO J* **19**:3263–3271.
- Schopperle WM, Holmqvist MH, Zhou Y, Wang J, Wang Z, Griffith LC, Keselman I, Kusinitz F, Dagan D, and Levitan IB (1998) Slob, a novel protein that interacts with the Slowpoke calcium-dependent potassium channel. *Neuron* **20**:565–573.
- Piros ET, Shen L, and Huang XY (1999) Purification of an EH domain-binding protein from rat brain that modulates the gating of the rat ether-à-go-go channel. *J Biol Chem* **274**:33677–33683.
- Hoshi N, Takahashi H, Shahidullah M, Yokoyama S, and Higashida H (1998) KCR1, a membrane protein that facilitates functional expression of non-inactivating K⁺ currents associated with rat EAG voltage-dependent K⁺ channels. *J Biol Chem* **273**:23080–23085.
- Bruggemann A, Pardo LA, Stuhmer W, and Pongs O (1993) *Ether-à-go-go* encodes a voltage-gated channel permeable to K⁺ and Ca²⁺ and modulated by cAMP. *Nature (Lond)* **365**:445–448.
- Terlau H, Ludwig J, Steffan R, Pongs O, Stuhmer W, and Heinemann SH (1996) Extracellular Mg²⁺ regulates activation of rat eag potassium channel. *Pflueg Arch Eur J Physiol* **432**:301–312.
- Schönherr R, Gessner G, Lober K, and Heinemann SH (2002) Functional distinction of human EAG1 and EAG2 potassium channels. *FEBS Lett* **514**:204–208.
- Stansfeld CE, Roper J, Ludwig J, Weseloh RM, Marsh SJ, Brown DA, and Pongs O (1996) Elevation of intracellular calcium by muscarinic receptor activation induces a block of voltage-activated rat ether-à-go-go channels in a stably transfected cell line. *Proc Natl Acad Sci USA* **93**:9910–9914.
- Lecain E, Sauvaget E, Crisanti P, Van Den Abbeele T, and Huy PT (1999) Potassium channel ether à go-go mRNA expression in the spiral ligament of the rat. *Hear Res* **133**:133–138.
- Meyer R, Schönherr R, Gavrilova-Ruch O, Wohlrab W, and Heinemann SH (1999) Identification of ether à go-go and calcium-activated potassium channels in human melanoma cells. *J Membr Biol* **171**:107–115.
- Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding subthreshold-operating voltage-gated K⁺ channels in brain. *J Neurosci* **21**:4609–4624.
- Pardo LA, del Camino D, Sanchez A, Alves F, Bruggemann A, Beckh S, and Stuhmer W (1999) Oncogenic potential of EAG K⁺ channels. *EMBO J* **18**:5540–5547.
- Camacho J, Sanchez A, Stuhmer W, and Pardo LA (2000) Cytoskeletal interactions determine the electrophysiological properties of human EAG potassium channels. *Pflueg Arch Eur J Physiol* **441**:167–174.
- Ludwig J, Owen D, and Pongs O (1997) Carboxy-terminal domain mediates assembly of the voltage-gated rat ether-à-go-go potassium channel. *EMBO J* **16**:6337–6345.
- Pardo LA, Bruggemann A, Camacho J, and Stühmer W (1998) Cell-cycle related changes in the conducting properties of r-eag K⁺ channels. *J Cell Biol* **143**:767–775.
- Fariás LM, Ocana DB, Diaz L, Larrea F, Avila-Chavez E, Cadena A, Hinojosa LM, Lara G, Villanueva LA, Vargas C, Hernandez-Gallegos E, et al. (2004) *Ether à go-go* potassium channels as human cervical cancer markers. *Cancer Res* **64**:6996–7001.
- Jenke M, Sanchez A, Monje F, Stuhmer W, Weseloh RM, and Pardo LA (2003) C-terminal domains implicated in the functional surface expression of potassium channels. *EMBO J* **22**:395–403.

TABLE 35
K_v10.2 channels

Channel name	K _v 10.2
Description	Outward-rectifying, noninactivating voltage-dependent K ⁺ currents ^{3–5}
Other names	eag2 ^{1–5}
Molecular information	Human: 987aa, NM_139318 (transcript variant 1), chr. 14q23.1, <i>KCNH5</i> (see “Comments”), GeneID: 27133, PMID: 9738473 ² Mouse: 988aa, NM_172805, chr. 12 Rat: 988aa, NM_133610, chr. 6q24
Associated subunits	Hyperkinetic (Hk), ⁶ CaM, Slob, KCR1 (potassium channel regulator)
Functional assays	Voltage-clamp
Current	Outward-rectifying
Conductance	Not established
Ion selectivity	K ⁺
Activation	Activates at –100 mV (rat) ³
Inactivation	Noninactivating
Activators	None
Gating inhibitors	None
Blockers	Quinidine (152 μM), ⁵ intracellular calcium (nanomolar) ⁴
Radioligands	None
Channel distribution	Brain (layer IV of the cerebral cortex; thalamus, inferior colliculus, olfactory bulb, and certain brainstem nuclei) ^{3,4}
Physiological functions	Not established
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	This channel has a GFG (rather than the common GYG) potassium channel signature sequence, a PAS domain in the distal part of the cytosolic N terminus, a cNBD domain in the proximal portion of the C terminus, a C-terminal assembly domain (CAD), a CaM-binding domain, a bNLS domain in the C terminus, and a C-terminal domain is required for assembly ⁷ ; the TCC domain at the C-terminal end of K _v 10 and K _v 11 confers specificity for multimer formation, allowing K _v 10.1/K _v 10.2 heteromerization and K _v 11 homomerization but not K _v 10.x/K _v 11.x heteromerization ⁸ ; this C-terminal TCC domain has been identified in many other channels, and mutations of the TCC have been found to be linked to genetic channelopathies

aa, amino acids; chr., chromosome; CaM, calmodulin; TCC, tetramerizing coiled-coiled.

1. Shi W, Wang HS, Pan Z, Wymore RS, Cohen IS, McKinnon D, and Dixon JE (1998) Cloning of a mammalian elk potassium channel gene and EAG mRNA distribution in rat sympathetic ganglia. *J Physiol* **511**:675–682.

2. Occhiodoro T, Bernheim L, Liu JH, Bijlenga P, Sinnreich M, Bader CR, and Fischer-Lougheed J (1998) Cloning of a human *ether-à-go-go* potassium channel expressed in myoblasts at the onset of fusion. *FEBS Lett* **434**:177–182.

3. Saganich MJ, Vega-Saenz de Miera E, Nadal MS, Baker H, Coetzee WA, and Rudy B (1999) Cloning of components of a novel subthreshold-activating K⁺ channel with a unique pattern of expression in the cerebral cortex. *J Neurosci* **19**:10789–10802.

4. Ludwig J, Weseloh R, Karschin C, Liu Q, Netzer R, Engeland B, Stansfeld C, and Pongs O (2000) Cloning and functional expression of rat eag2, a new member of the *ether-à-go-go* family of potassium channels and comparison of its distribution with that of eag1. *Mol Cell Neurosci* **16**:59–70.

5. Schonherr R, Gessner G, Lober K, and Heinemann SH (2002) Functional distinction of human EAG1 and EAG2 potassium channels. *FEBS Lett* **514**:204–208.

6. Wilson GF, Wang Z, Chouinard SW, Griffith LC, and Ganetzky B (1998) Interaction of the K channel β subunit, Hyperkinetic, with eag family members. *J Biol Chem* **273**:6389–6394.

7. Ludwig J, Owen D, and Pongs O (1997) Carboxy-terminal domain mediates assembly of the voltage-gated rat *ether-à-go-go* potassium channel. *EMBO J* **16**:6337–6345.

8. Jenke M, Sanchez A, Monje F, Stuhmer W, Weseloh RM, and Pardo LA (2003) C-terminal domains implicated in the functional surface expression of potassium channels. *EMBO J* **22**:395–403.

TABLE 36
K_v11.1 channels

Channel name	K _v 11.1
Description	Voltage-gated potassium channel with inwardly rectifying properties
Other names	Human <i>ether-à-go-go</i> -related gene, HERG, erg1, Hergb ¹⁻⁸
Molecular information	Human: 1159aa, NM_000238 (transcript variant 1), chr. 7q35-36, ¹ <i>KCNH2</i> , GeneID: 3757, PMID: 8159766 ¹ Mouse: 1162aa, NM_013569, chr. 5 Rat: 1163aa, NM_053949, chr. 4q11 minK, ^{9,25} possibly MiRP1 (KCNE2) ¹⁰
Associated subunits	
Functional assays	Voltage-clamp
Current	Cardiac I _{Kr} current ^{3,26}
Conductance	2pS (in physiological [K] _o), 10pS (100 mM [K] _o) ¹¹
Ion selectivity	K ⁺
Activation	Activation at currents more positive than -50 mV ^{3,26}
Inactivation	Exhibits C-type inactivation ⁴ ; inward rectification arises from a rapid and voltage-dependent inactivation process that reduces conductance at positive voltages ^{3,26,27}
Activators	None
Gating inhibitors	None
Blockers	Astemizole (1 nM), ¹³ BeKM-1 (3 nM), ¹⁴ ergtoxin (12 nM), ¹⁵ sertindole (3 nM), dofetilide (15–35 nM), ¹⁶ cisapride (6–40 nM), pimozone (18 nM), terfenadine (56 nM), halofantrine (200 nM), BRL32872 (240 nM), E-4031 (7.7 nM), CT haloperidol (1 μM), imipramine (3 μM), cocaine (5 μM), ketoconazole
Radioligands	None
Channel distribution	Heart, leiomyosarcoma, hippocampus, neuroblastoma, blood cells, brain, kidney, liver, lung, ovary, pancreas, testis, prostate, small intestine, tonsil, uterus, microglia
Physiological functions	HERG proteins form cardiac I _{Kr} channels ^{3,26} ; in the heart, HERG channels produce a resurgent current during repolarization ²⁰ due to the recovery from C-type inactivation ⁴ and a slow deactivation due to an interaction with an N-terminal domain (AA2–16) and the internal mouth of the pore ^{1,22} ; HERG contains a tetramerization domain called NAB and a structurally defined PAS domain in distinct regions of the N terminus ¹⁷ ; HERG forms a complex with MiRP1, ¹⁰ but it is as yet unclear whether MiRP1 forms a stable part of the channel itself or is otherwise involved in regulation of HERG expression or stability ²³
Mutations and pathophysiology	Mutations of this gene cause the autosomal dominant long QT syndrome 2 due to gating defects ²⁸ and trafficking abnormalities ^{29–33} and a prolonged QT interval on the electrocardiogram; syncope, sudden cardiac death, ventricular fibrillation, and torsades de pointes are also implicated in acquired long QT syndrome; mutations in MiRP1 are the cause of long QT syndrome 6 and are also found in many tumors ^{18,19}
Pharmacological significance	Proarrhythmic potential (QT prolongation) of histamine H ₁ receptor antagonists, antipsychotics, and tricyclic antidepressants that leads to torsades de pointes in some individuals (acquired long QT syndrome)
Comments	A shorter isoform encoded by an alternative transcript (1b) of K _v 11.1 ^{5,7} or a truncated isoform ⁶ can coassemble with and modulate the behavior of full-length HERG and Merg1, the mouse ortholog; the TCC domain at the C-terminal end of K _v 10 and K _v 11 confers specificity for multimer formation, allowing K _v 10.1/K _v 10.2 heteromerization and K _v 11 homomerization, but not K _v 10.x/K _v 11.x heteromerization ²⁴ ; this C-terminal TCC domain has been identified in many other channels, and mutations of the TCC are found to be linked to genetic channelopathies; C terminus interacts with Golgi matrix protein GM130 ³⁴

aa, amino acids; chr., chromosome; MiRP1, MinK-related peptide 1; TCC, tetramerizing coiled-coiled; E-4031, *N*-[4-[[1-[2-(6-methyl-2-pyridinyl)ethyl]-4-piperidinyl]carbonyl]phenyl]methanesulfonamide dihydrochloride.

1. Warmke JW and Ganetzky B (1994) A family of potassium channel genes related to eag in *Drosophila* and mammals. *Proc Natl Acad Sci USA* **91**:3438–3442.
2. Curran ME, Splawski I, Timothy KW, Vincent GM, Green ED, and Keating MT (1995) A molecular basis for cardiac arrhythmia: HERG mutations cause long QT syndrome. *Cell* **80**:795–803.
3. Sanguinetti MC, Jiang C, Curran ME, and Keating MT (1995) A mechanistic link between an inherited and an acquired cardiac arrhythmia: HERG encodes the I_{Kr} potassium channel. *Cell* **81**:299–307.
4. Smith PL, Baukrowitz T, and Yellen G (1996) The inward rectification mechanism of the HERG cardiac potassium channel. *Nature (Lond)* **379**:833–836.
5. Lees-Miller JP, Kondo C, Wang L, and Duff HJ (1997) Electrophysiological characterization of an alternatively processed ERG K⁺ channel in mouse and human hearts. *Circ Res* **81**:719–726.
6. Kupersmidt S, Snyders DJ, Raes A, and Roden DM (1998) A K⁺ channel splice variant common in human heart lacks a C-terminal domain required for expression of rapidly activating delayed rectifier current. *J Biol Chem* **273**:27231–27235.
7. London B, Trudeau MC, Newton KP, Beyer AK, Copeland NG, Gilbert DJ, Jenkins NA, Satler CA, and Robertson GA (1997) Two isoforms of the mouse *ether-à-go-go*-related gene co-assemble to form channels with properties similar to the rapidly activating component of the cardiac delayed rectifier K⁺ current. *Circ Res* **81**:870–878.
8. Itoh T, Tanaka T, Nagai R, Kamiya T, Sawayama T, Nakayama T, Tomoike H, Sakurada H, Yazaki Y, and Nakamura Y (1998) Genomic organization and mutational analysis of HERG, a gene responsible for familial long QT syndrome. *Hum Genet* **102**:435–439.
9. McDonald TV, Yu Z, Ming Z, Palma E, Meyers MB, Wang KW, Goldstein SA, and Fishman GI (1997) A minK-HERG complex regulates the cardiac potassium current I_{Kr}. *Nature (Lond)* **388**:289–292.
10. Abbott GW, Sesti F, Splawski I, Buck ME, Lehmann MH, Timothy KW, Keating MT, and Goldstein SA (1999) MiRP1 forms I_{Kr} potassium channels with HERG and is associated with cardiac arrhythmia. *Cell* **97**:175–187.
11. Kiehn J, Lacerda AE, Wible B, and Brown AM (1996) Molecular physiology and pharmacology of HERG: single-channel currents and block by dofetilide. *Circulation* **94**:2572–2579.
12. Zhou Z, Gong Q, Ye B, Fan Z, Makielski JC, Robertson GA, and January CT (1998) Properties of HERG channels stably expressed in HEK 293 cells studied at physiological temperature. *Biophys J* **74**:230–241.
13. Suessbrich H, Waldegger S, Lang F, and Busch AE (1996) Blockade of HERG channels expressed in *Xenopus* oocytes by the histamine receptor antagonists terfenadine and astemizole. *FEBS Lett* **385**:77–80.

14. Korolkova YV, Kozlov SA, Lipkin AV, Pluzhnikov KA, Hadley JK, Filippov AK, Brown DA, Angelo K, Strobaek D, Jespersen T, et al. (2001) An ERG channel inhibitor from the scorpion *Buthus eupeus*. *J Biol Chem* **276**:9868–9876.

15. Pardo-Lopez L, Zhang M, Liu J, Jiang M, Possani LD, and Tseng GN (2002) Mapping the binding site of a human *ether-à-go-go*-related gene-specific peptide toxin (ErgTx) to the channel's outer vestibule. *J Biol Chem* **277**:16403–16411.

16. Spector PS, Curran ME, Keating MT, and Sanguinetti MC (1996) Class III antiarrhythmic drugs block HERG, a human cardiac delayed rectifier K⁺ channel: open-channel block by methane sulfonanilides. *Circ Res* **78**:499–503.

17. Morais Cabral JH, Lee A, Cohen SL, Chait BT, Li M, and Mackinnon R (1998) Crystal structure and functional analysis of the HERG potassium channel N terminus: eukaryotic PAS domain. *Cell* **95**:649–655.

18. Bianchi L, Wible B, Arcangeli A, Taglialatela M, Morra F, Castaldo P, Crociani O, Rosati B, Faravelli L, Olivetto M, and Wanke E (1998) herg encodes a K⁺ current highly conserved in tumors of different histogenesis: a selective advantage for cancer cells? *Cancer Res* **58**:815–822.

19. Smith GA, Tsui HW, Newell E, Jiang X, Zhu XP, Tsui FW, and Schlichter LC (2002) Functional up-regulation of HERG K⁺ channels in neoplastic hematopoietic cells. *J Biol Chem* **277**:18528–18534.

20. Zhou Z, Gong Q, Ye B, Fan Z, Makielski JC, Robertson GA, and January CT (1998) Properties of HERG channels stably expressed in HEK 293 cells. *Biophys J* **74**:230–241.

21. Wang J, Trudeau MC, Zappia AM, and Robertson GA (1998) Regulation of deactivation by an amino terminal domain in human *ether-à-go-go*-related gene potassium channels. *J Gen Physiol* **112**:637–647.

22. Wang J, Myers CD, and Robertson GA (2000) Dynamic control of deactivation gating by a soluble amino-terminal domain in HERG K⁺ channels. *J Gen Physiol* **115**:749–758.

23. Weerapura M, Nattel S, Chartier D, Caballero R, and Hebert TE (2002) A comparison of currents carried by HERG, with and without coexpression of MiRP1, and the native rapid delayed rectifier current. Is MiRP1 the missing link? *J Physiol* **540**:15–27.

24. Jenke M, Sanchez A, Monje F, Stuhmer W, Weseloh RM, and Pardo LA (2003) C-terminal domains implicated in the functional surface expression of potassium channels. *EMBO J* **22**:395–403.

25. Finley MR, Li Y, Hua F, Lillich J, Mitchell KE, Ganta S, Gilmour RF Jr, and Freeman LC (2002) Expression and coassociation of ERG1, KCNQ1, and KCNE1 potassium channel proteins in horse heart. *Am J Physiol Heart Circ Physiol* **283**:H126–H138.

26. Trudeau M, Warmke JW, Ganetzky B, and Robertson GA (1995) HERG, a human inward rectifier in the voltage-gated potassium channel family. *Science (Wash DC)* **269**:92–95.

27. Spector PS, Curran EM, Zou A, Keating MT, and Sanguinetti MC (1996) Fast inactivation causes rectification of the IKr channel. *J Gen Physiol* **107**:611–619.

28. Sanguinetti MC, Curran ME, Spector PS, and Keating MT (1996) Spectrum of HERG K⁺-channel dysfunction in an inherited cardiac arrhythmia[published erratum appears in *Proc Natl Acad Sci USA* (1996) **93**:8796]. *Proc Natl Acad Sci USA* **93**:2208–2212.

29. Furutani M, Trudeau MC, Hagiwara N, Seki A, Gong Q, Zhou Z, Imamura S, Nagashima H, Kasanuki H, Takao A, et al. (1999) Novel mechanism associated with an inherited cardiac arrhythmia: defective protein trafficking by the mutant HERG (G601S) potassium channel. *Circulation* **99**:2290–2294.

30. Zhou Z, Gong Q, and January CT (1999) Correction of defective protein trafficking of a mutant HERG potassium channel in human long QT syndrome. Pharmacological and temperature effects. *J Biol Chem* **274**:31123–31126.

31. Ficker E, Dennis AT, Obejero-Paz CA, Castaldo P, Taglialatela M, and Brown AM (2000) Retention in the endoplasmic reticulum as a mechanism of dominant-negative current suppression in human long QT syndrome. *J Mol Cell Cardiol* **32**:2327–2337.

32. Ficker E, Thomas D, Viswanathan PC, Dennis AT, Priori SG, Napolitano C, Memmi M, Wible BA, Kaufman ES, Iyengar S, et al. (2000) Novel characteristics of a misprocessed mutant HERG channel linked to hereditary long QT syndrome. *Am J Physiol Heart Circ Physiol* **279**:H1748–H1756.

33. Rajamani S, Anderson CL, Anson BD, and January CT (2002) Pharmacological rescue of human K⁺ channel long-QT2 mutations: human *ether-à-go-go*-related gene rescue without block. *Circulation* **105**:2830–2835.

34. Roti Roti EC, Myers CD, Ayers RA, Boatman DE, Delfosse SA, Chan EK, Ackerman MJ, January CT, and Robertson GA (2002) Interaction with GM130 during HERG ion channel trafficking: disruption by type 2 congenital long QT syndrome mutations. *J Biol Chem* **277**:47779–47785.

TABLE 37
K_V11.2 channels

Channelname	K _V 11.2
Description	Voltage-gated potassium channel
Other names	erg2 ^{1,2}
Molecular information	Human: 994aa, NM_030779 (transcript variant 1) chr. 17q23.3, <i>KCNH6</i> , GeneID: 81033, PMID: 10414305 ⁶ Rat: 950aa, NM_053937, chr. 10q32.1
Associated subunits	See “Comments”
Functional assays	Voltage-clamp
Current	Not established
Conductance	Not established
Ion selectivity	Not established
Activation	Not established
Inactivation	Not established
Activators	None
Gating inhibitors	None
Blockers	Sipatrigine
Radioligands	None
Channel distribution	Brain, ² uterus, leiomyosarcoma, hippocampus, neuroblastoma, lactotrophs, ³ GH3/B6 cells, rat pituitary ⁴
Physiological functions	Not established
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	K _V 11.1, K _V 11.2, and K _V 11.3 can form heteromultimers ⁵

aa, amino acids; chr., chromosome.

1. Schafer R, Wulfsen I, Behrens S, Weinsberg F, Bauer CK, and Schwarz JR (1999) The erg-like potassium current in rat lactotrophs. *J Physiol* **518**:401–416.

2. Ganetzky BS and Titus SA (2000) inventors, Wisconsin Alumni Research Foundation, assignee. Potassium ion channel genes and proteins. U.S. patent A6,087,488. 2000 11 Jul.

3. Shi W, Wymore RS, Wang HS, Pan Z, Cohen IS, McKinnon D, and Dixon JE (1997) Identification of two nervous system-specific members of the erg potassium channel gene family. *J Neurosci* **17**:9423–9432.

4. Wulfsen I, Hauber HP, Schiemann D, Bauer CK, and Schwarz JR (2000). Expression of mRNA for voltage-dependent and inward-rectifying K channels in GH3/B6 cells and rat pituitary. *J Neuroendocrinol* **12**:263–272.

5. Wimmers S, Wulfsen I, Bauer CK, and Schwarz JR (2001). Erg1, erg2 and erg3 K channel subunits are able to form heteromultimers. *Pflug Arch Eur J Physiol* **441**:450–455.

6. Ganetzky B, Robertson GA, Wilson GF, Trudeau MC, and Titus SA (1999) The eag family of K⁺ channels in *Drosophila* and mammals. *Ann NY Acad Sci* **868**:356–369.

TABLE 38
K_V11.3 channels

Channel name	K _V 11.3
Description	Voltage-gated potassium channel
Other names	erg3 ^{1–3}
Molecular information	Human: 1196aa, NM_033272 (transcript variant 1), chr. 2q24.2, <i>KCNH7</i> , GeneID: 90134, PMID: 10414305 ⁹ Mouse: 1195aa, NM_133207, chr. 2 Rat: 1195aa, NM_131912, chr. 3q21
Associated subunits	See “Comments”
Functional assays	Voltage-clamp
Current	Not established
Conductance	Not established
Ion selectivity	K ²⁺
Activation	Activated at –50 mV ² (see “Comments”)
Inactivation	Not established
Activators	None
Gating inhibitors	None
Blockers	Sertindole (43 nM) ² and pimoizide (103 nM) ²
Radioligands	None
Channel distribution	Brain, sympathetic ganglia, CA pyramidal neurons, ⁴ lactotrophs, ⁵ GH3/B6 cells, rat pituitary ⁶
Physiological functions	Not established
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	Thyrotropin-releasing hormone reduces K _V 11.3 currents and shifts the voltage dependence of activation by 6 mV ⁷ ; K _V 11.1, K _V 11.2, and K _V 11.3 can form heteromultimers ⁸

aa, amino acids; chr., chromosome.

1. Shi W, Wymore RS, Wang HS, Pan Z, Cohen IS, McKinnon D, and Dixon JE (1997) Identification of two nervous system-specific members of the erg potassium channel gene family. *J Neurosci* **17**:9423–9432.

2. Kang J, Chen XL, and Rampe D (2001) The antipsychotic drugs sertindole and pimoizide block erg3, a human brain K⁺ channel. *Biochem Biophys Res Commun* **286**:499–504.

3. Ganetzky BS and Titus SA (2000) inventors, Wisconsin Alumni Research Foundation, assignee. Potassium ion channel genes and proteins. U.S. patent A6,087,488. 2000 11 Jul.

4. Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding subthreshold-operating voltage-gated K⁺ channels in brain. *J Neurosci* **21**:4609–4624.

5. Schafer R, Wulfsen I, Behrens S, Weinsberg F, Bauer CK, and Schwarz JR (1999) The erg-like potassium current in rat lactotrophs. *J Physiol* **518**:401–416.

6. Wulfsen I, Hauber HP, Schiemann D, Bauer CK, and Schwarz JR (2000) Expression of mRNA for voltage-dependent and inward-rectifying K channels in GH3/B6 cells and rat pituitary. *J. Neuroendocrinol* **12**:263–272.

7. Schledermann W, Wulfsen I, Schwarz JR, and Bauer CK (2001) Modulation of rat erg1, erg2, erg3 and HERG K⁺ currents by thyrotropin-releasing hormone in anterior pituitary cells via the native signal cascade. *J Physiol* **532**:143–163.

8. Wimmers S, Wulfsen I, Bauer CK, and Schwarz JR (2001) Erg1, erg2 and erg3 K channel subunits are able to form heteromultimers. *Pflueg Arch Eur J Physiol* **441**:450–455.

9. Ganetzky B, Robertson GA, Wilson GF, Trudeau MC, and Titus SA (1999) The eag family of K⁺ channels in *Drosophila* and mammals. *Ann NY Acad Sci* **868**:356–369.

TABLE 39
K_V12.1 channels

Channel name	K _V 12.1
Description	Slowly activating and deactivating voltage-gated potassium channel ¹
Other names	elk1, ¹ elk3 ²
Molecular information	Human: 1107aa, NM_144633, chr. 3p24.3, <i>KCNH8</i> , GeneID: 131096, PMID: 12890647 ³ Mouse: 1102aa, NM_001031811, chr. 17 Rat: 1102aa, NM_145095, chr. 9q11 (see “Comments”)
Associated subunits	Not established
Functional assays	Voltage-clamp
Current	None identified
Conductance	Not established
Ion selectivity	K ⁺
Activation	Not established
Inactivation	Not established
Activators	None
Gating inhibitors	None
Blockers	Ba ²⁺ ¹
Radioligands	None
Channel distribution	Sympathetic ganglia, testis, brain, colon, lung, uterus, pre-B cell leukemia (ESTs) ^{1,2}
Physiological functions	Not established
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	There is a light oxygen voltage (LOV) and cyclic nucleotide binding (CNB) domain in the N and C terminus, respectively.

aa, amino acids; chr., chromosome.

1. Shi W, Wang HS, Pan Z, Wymore RS, Cohen IS, McKinnon D, and Dixon JE (1998) Cloning of a mammalian elk potassium channel gene and EAG mRNA distribution in rat sympathetic ganglia *J Physiol* **511**:675–682.

2. Engeland B, Neu A, Ludwig J, Roeper J, and Pongs O (1998) Cloning and functional expression of rat *ether-à-go-go*-like K⁺ channel genes. *J Physiol* **513**:647–654.

3. Zou A, Lin Z, Humble M, Creech CD, Wagoner PK, Krafte D, Jegla TJ, and Wickenden AD (2003) Distribution and functional properties of human KCNH8 (Elk1) potassium channels. *Am J Physiol Cell Physiol* **285**:C1356–C1366.

TABLE 40
K_V12.2 channels

Channel name	K _V 12.2
Description	Voltage-gated potassium channel
Other names	BEC1, ¹ Elk2 ²
Molecular information	Human: 1083aa, NM_012284, chr. 12q13, ¹ <i>KCNH3</i> , GeneID: 23416, PMID: 10455180 ¹ Mouse: 1095aa, NM_010601, chr. 15 Rat: 1087aa, NM_017108, 7q36
Associated subunits	None determined
Functional assays	Voltage-clamp
Current	Not established
Conductance	Not established
Ion selectivity	K ⁺
Activation	Not established
Inactivation	Fast ^{1,2}
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Infant brain, lung (small cell carcinoma), eye (retinoblastoma), sciatic nerve, cortex, amygdala, hippocampus (mainly in CA1 and CA3 pyramidal cell body layers and in the granule cell layers of the dentate gyrus); in the striatal regions, including the putamen and caudate nucleus, lymphocytes, leukemias, and NG108-15 cell line ¹⁻⁵
Physiological functions	Not established
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	There is a light oxygen voltage (LOV) and cyclic nucleotide binding (CNB) domain in the N and C terminus, respectively.

aa, amino acids; chr., chromosome.
1. Miyake A, Mochizuki S, Yokoi H, Kohda M, and Furuichi K (1999) New *ether-à-go-go* K⁺ channel family members localized in human telencephalon. *J Biol Chem* **274**:25018–25025.
2. Engeland B, Neu A, Ludwig J, Roeper J, and Pongs O (1998) Cloning and functional expression of rat *ether-à-go-go*-like K⁺ channel genes. *J Physiol* **513**:647–654.
3. Meves H, Schwarz JR, and Wulfsen I (1999) Separation of M-like current and ERG current in NG108–15 cells. *Br J Pharmacol* **127**:1213–1223.
4. Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding subthreshold-operating voltage-gated K⁺ channels in brain. *J Neurosci* **21**:4609–4624.
5. Smith GA, Tsui HW, Newell EW, Jiang X, Zhu XP, Tsui FW, and Schlichter LC (2002) Functional up-regulation of HERG K⁺ channels in neoplastic hematopoietic cells. *J Biol Chem* **277**:18528–18534.

TABLE 41
K_V12.3 channels

Channel name	K _V 12.3
Description	Slowly activating voltage-gated potassium channel
Other names	BEC2, ¹ elk1 ²
Molecular information	Human: 1017aa, NM_012285, <i>KCNH4</i> , chr. 17q21.2, GeneID: 23415, PMID: 10455180 ¹ Rat: 1017aa, NM_053630, chr. 10q32.1 (see “Comments”)
Associated subunits	None
Functional assays	Voltage-clamp
Current	Not established
Conductance	Not established
Ion Selectivity	K ⁺
Activation	Threshold for activation is 90 mV ²
Inactivation	Not established
Activators	None
Gating inhibitors	None
Blockers	Ba ²⁺ ²
Radioligands	None
Channel distribution	Brain (telencephalon), ^{1,3} neuroblastoma, esophagus, oligodendroglioma, lung, primary B-cell neoplasia, cerebellum, pituitary gland ⁴
Physiological functions	Not established
Mutations and pathophysiology	Not established
Pharmacological significance	Not established
Comments	There are light oxygen voltage (LOV) and cyclic nucleotide binding (CNB) domains in the N and C terminus, respectively.

aa, amino acids; chr., chromosome.
1. Miyake A, Mochizuki S, Yokoi H, Kohda M, and Furuichi K (1999) New *ether-à-go-go* K⁺ channel family members localized inhuman telencephalon. *J Biol Chem* **274**:25018–25025.
2. Engeland B, Neu A, Ludwig J, Roeper J, and Pongs O (1998) Cloning and functional expression of rat *ether-à-go-go*-like K⁺ channel genes. *J Physiol* **513**:647–654.
3. Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding sub threshold-operating voltage-gated K⁺ channels in brain. *J Neurosci* **21**:4609–4624.
4. Wulfsen I, Hauber HP, Schiemann D, Bauer CK, and Schwarz JR (2000) Expression of mRNA for voltage-dependent and inward-rectifying K channels in GH3/B6 cells and rat pituitary. *J Neuroendocrinol* **12**:263–272.