

XIV. International Union of Pharmacology Nomenclature in Nitric Oxide Research^a

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I. Introduction	137
II. Nitric oxide synthases	138
III. Inhibitors of nitric oxide synthase	138
IV. Nitric oxide donors	140
V. Recommended nomenclature and abbreviations	140
A. Abbreviations for isoenzymes of nitric oxide synthase	140
B. Abbreviations for nitric oxide synthase substrates	140
C. Abbreviations for inhibitors of nitric oxide synthase	140
D. Abbreviations for certain nitric oxide donors	140
E. Nitregic nerves	141
VI. Conclusion	141
VII. Acknowledgments	141
VIII. References	141

I. Introduction

The discovery that the biological actions of endothelium-derived relaxing factor (Furchgott and Zawadzki,

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^c Abbreviations: NO, nitric oxide; NOS, NO synthase.

1980) are due to the endogenous release of nitric oxide (NO)^c (Palmer et al., 1987; Ignarro et al., 1987; Khan and Furchgott, 1987) revealed the existence of a ubiquitous biochemical pathway (Moncada et al., 1989). NO is formed from the amino acid L-arginine by a family of enzymes, the NO synthases (NOSs), and plays a role in many physiological functions. Its formation in vascular endothelial cells, in response to chemical stimuli and to physical stimuli such as shear stress, maintains a vasodilator tone that is essential for the regulation of blood flow and pressure (Moncada et al., 1989; Vanhoutte, 1989; Furchgott, 1990; Ignarro, 1990; Vane et al., 1990; Luscher, 1991). NO produced by the endothelium and/or platelets also inhibits platelet aggregation and adhesion, inhibits leukocyte adhesion and modulates smooth muscle cell proliferation (Moncada and Higgs, 1993). NO is synthesized in neurons of the central nervous system, where it acts as a neuromediator with many physiological functions, including the formation of memory, coordination between neuronal activity and blood flow, and modulation of pain (Garthwaite, 1991; Snyder and Brecht, 1992). In the peripheral nervous system, NO is now known to be the mediator released by a widespread network of nerves, previously recognized as nonadrenergic and noncholinergic. These nerves mediate some forms of neurogenic vasodilation and regulate certain gastrointestinal, respiratory and genitourinary functions (Gillespie et al., 1990; Rand, 1992; Toda, 1995). These physiological actions of NO are mediated by activation of the soluble guanylate cyclase and consequent

increase in the concentration of cyclic guanosine monophosphate in target cells (Murad et al., 1990; Ignarro, 1991).

In addition, NO is generated in large quantities during host defense and immunological reactions (Nathan and Hibbs, 1991; Nussler and Billiar, 1993). Such generation of NO was first observed in activated macrophages (Hibbs et al., 1988; Marletta et al., 1988; Stuehr et al., 1989), where it contributes to their cytotoxicity against tumor cells, bacteria, viruses and other invading microorganisms. The cytostatic/cytotoxic actions of NO result from its inhibitory actions on key enzymes in the respiratory chain and in the synthesis of deoxyribonucleic acid in the target cells (Hibbs et al., 1990; Nguyen et al., 1992). NO may also interact with oxygen-derived radicals to produce other toxic substances (Hibbs, 1992) such as peroxynitrite (Beckman et al., 1990). Peroxynitrite is a powerful oxidant; however, there seem to be very effective mechanisms for its removal and inactivation (Moro et al., 1994). Thus, NO plays a role in immunological host defense and is also involved in the pathogenesis of conditions such as septic shock and inflammation.

NO is a gas at temperatures down to -152°C . It is slightly soluble in many solvents and can diffuse relatively easily across biological membranes. Its solubility in water is low so that it can only occur in dilute solution. Nitrite, rather than nitrite plus nitrate, is believed to be the product of NO in oxygenated water (see Butler et al., 1995; Williams, 1996). Table 1 shows several chemical species related to NO.

II. Nitric Oxide Synthases

NOS is a heme-containing enzyme with a sequence similarity to cytochrome P-450 reductase. Several isoforms of NOS are now known to exist, two of which are constitutive and one of which is inducible by immuno-

TABLE 1
NO and related species

Common Name	Abbreviation/Symbol
Nitric oxide ^a	NO
Nitroxyl anion	NO ⁻
Nitrosonium ^b	NO ⁺
Nitrogen dioxide	NO ₂
Peroxynitrite	ONOO ⁻
Superoxide ^c	O ₂ ⁻
Nitrite	NO ₂ ⁻
Nitrate	NO ₃ ⁻
Nitrite + nitrate ^d	NO _x
Nitrosothiols	RSNO

^a The unpaired electron does *not* need to be written into the abbreviation for NO as NO[•].

^b Although some compounds act as donors of a species with an NO⁺-like character (i.e., are nitrosating agents), NO⁺ does not exist as such in aqueous solution.

^c NO reacts with the superoxide anion to form peroxynitrite.

^d In some conditions, S-nitrosothiols may also contribute to the NO_x measured by chemiluminescence.

logical stimuli (for reviews see Knowles and Moncada, 1994; Morris and Billiar, 1994; Nathan and Xie, 1994; Sessa, 1994; Stuehr and Griffith, 1992). The constitutive NOS (cNOS) that was first discovered in the vascular endothelium has been designated as eNOS, whereas that present in the brain, spinal cord and peripheral nervous system is termed nNOS. The form of NOS induced by immunological or inflammatory stimuli is known as iNOS. A comparison of the properties of these three major isoforms of the enzyme is shown in table 2.

The complementary deoxyribonucleic acids for all three of these isoforms have been cloned from several species, including humans (Charles et al., 1993; Geller et al., 1993; Marsden et al., 1992; Nakane et al., 1993). This has revealed further differences among them, which are shown in table 3. Knockout mice have been generated for each of the three NOS isoforms and have provided useful information concerning the role of each isoform and the effects of its deletion in whole animals (Huang and Fishman, 1996).

The apparent association of the three NOS isoenzymes with the endothelium, neurons and inducibility (table 2) is an oversimplification. For example, eNOS is located not only in the vascular endothelial cells but also in platelets (Radomski et al., 1990) and in certain neuronal populations in the brain (Dinerman et al., 1994), whereas nNOS has been found in the epithelium of the bronchi and trachea (Kobzik et al., 1993), as well as in skeletal muscle (Kobzik et al., 1994). Furthermore, some differences have been identified between iNOSs obtained from different tissues within the same species (Mohaupt et al., 1994). In addition, the constitutive eNOS can be induced in certain situations such as during chronic exercise (Sessa et al., 1994) or during pregnancy, when both eNOS and iNOS are induced (Weiner et al., 1994), whereas iNOS appears to be present constitutively in some tissues, including human bronchial epithelium (Kobzik et al., 1993), rat kidney (Mohaupt et al., 1994) and some fetal tissues (Baylis et al., 1994).

III. Inhibitors of Nitric Oxide Synthase

The generation of NO from L-arginine proceeds via the formation of N^ω-hydroxy-L-arginine (Pufahl et al., 1992). This L-arginine/NO pathway can be inhibited by several analogues of L-arginine, of which the first to be identified was N^ω-monomethyl-L-arginine (see Moncada and Higgs, 1993). Before the discovery of NO as a biological mediator, N^ω-monomethyl-L-arginine had been found to prevent L-arginine-dependent cytotoxicity in murine macrophages (see Hibbs et al., 1990). It has subsequently been shown to be a competitive inhibitor of the formation of NO in the vasculature and elsewhere (see Moncada et al., 1989), and, as such, it has become a valuable tool for unraveling the biological actions of NO.

Many other analogues of L-arginine are now known to act as competitive (and in some cases, irreversible) inhibitors of both the constitutive and the inducible NOS.

TABLE 2
Isoforms of NOS

	<i>Endothelial NOS (eNOS)</i> ^a (Type III NOS, NOS-3)	<i>Neuronal NOS (nNOS)</i> ^a (Type I NOS, NOS-1, bNOS)	<i>Inducible NOS (iNOS)</i> ^a (Type II NOS, NOS-2, macNOS, hepNOS)
Primary regulation	Ca ²⁺ /calmodulin	Ca ²⁺ /calmodulin	Gene expression
Subcellular location	Membrane >> cytosol	Cytosol?	Cytosol >> membrane
NO output ^b	Low (pmolar)	Low (pmolar)	High (μmolar)
Function	Cell signaling	Cell signaling	Cytotoxic Cytostatic Cytoprotective

^a Previously used alternative abbreviation.

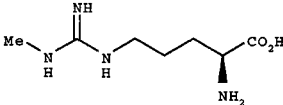
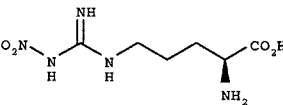
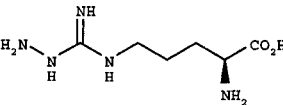
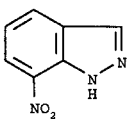
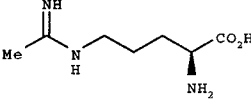
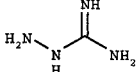
^b All three isolated purified isoforms of NOS have comparable specific activities. Output in this case refers to differences in levels of enzyme expression and in duration of activation in the in vivo situation.

TABLE 3
Molecular biology of human NOS isoforms^a

Gene	eNOS	nNOS	iNOS
Location	7q35–7q36	12q24.2	17cen–17q11.2
Structure	26 exons	29 exons	27 exons
Size	21 kb	160 kb	37 kb
Peptide	1203 aa	1434 aa	1153 aa
(deduced)	133 kDa	160 kDa	131 kDa

^a Examples of accession numbers: eNOS - M95296; nNOS - U17327; iNOS - X73029.

Other non-amino acid compounds that mimic the guanidinium moiety of L-arginine also inhibit the enzyme. Some inhibitors of NOS are shown in figure 1. Interestingly, asymmetric dimethyl-L-arginine (N^G-N^G-dimethyl-L-arginine, L-ADMA), an inhibitor of NOS, and N^ω-monomethyl-L-arginine are present in human plasma and urine (Vallance et al., 1992). Accumulation of these compounds in the plasma may contribute to the pathophysiology of some conditions.

Compound	Abbreviation	Structure	Inhibitory potency
N ^ω -monomethyl-L-arginine	L-NMMA		nNOS = eNOS > iNOS
N ^ω -nitro-L-arginine	L-NA		nNOS = eNOS >> iNOS
N ^ω -amino-L-arginine	L-NAA		nNOS = iNOS > eNOS
7-nitroindazole	7-NI		nNOS = eNOS = iNOS
N-δ-iminoethyl-L-ornithine	L-NIO		iNOS > eNOS = nNOS*
Aminoguanidine			iNOS > eNOS = nNOS

* Despite its lack of selectivity on the isolated enzymes, 7NI shows apparent pharmacological selectivity for nNOS.

FIG. 1. Some inhibitors of NOS.

There are several clinical situations in which it may be desirable to inhibit the production of NO, either by a constitutive NOS (for example, in cerebral ischemia or epilepsy, in which overproduction of NO by nNOS may lead to neurotoxicity) or by iNOS in conditions such as septic shock or certain chronic inflammatory diseases. Although much effort is being put into the search for a selective inhibitor of iNOS and some of the known inhibitors have shown some degree of selectivity in vitro toward one or another NOS isoform (fig. 1), thus far, the drugs available affect both the inducible and the constitutive isoforms (for review, see Griffith and Gross, 1996).

IV. Nitric Oxide Donors

The clinical actions of the nitrovasodilators are now known to result from their ability to liberate NO (Feelisch, 1991); thus, the term "NO donors" has been adopted for this class of drug. Such compounds include the organic nitrates (e.g., glyceryl trinitrate) and nitrites (e.g., amyl nitrite), inorganic nitroso compounds (e.g., sodium nitroprusside), sydnonimines (e.g., molsidomine) and S-nitrosothiols (e.g., S-nitrosoglutathione) (Feelisch and Stamler, 1996 and table 4).

Although NO donors have been used traditionally as vasodilators, other clinical applications are emerging as our understanding of the biological actions of NO itself increases. Thus, compounds such as S-nitrosoglutathione, which potently inhibits platelet aggregation at concentrations that do not affect blood pressure (de Belder et al., 1995), may be useful in the treatment of certain thrombotic disorders. Other NO donors have been shown to reduce intimal thickening in injured arteries in an animal model (Lefer and Lefer, 1994). Impaired generation of NO by nitrenergic nerves, i.e., those that liberate NO as a neuromodulator, may underlie certain gastrointestinal, genitourinary and respiratory disorders. In such cases, NO donors may mimic nitrenergic nerve-mediated responses and have been shown to be effective in the treatment of achalasia and other malfunctions of sphincters in the gastrointestinal tract, as well as in the treatment of impotence in diabetes (see Moncada and Higgs, 1995).

TABLE 4
Recommended abbreviations for certain NO donors

Compound	Class	Abbreviation
Glyceryl trinitrate	Organic nitrate	GTN
Sodium nitroprusside	Inorganic iron complex	SNP
S-nitroso-L-cysteine	S-nitrosothiol	CysNO
S-nitrosoglutathione	S-nitrosothiol	GSNO
S-nitroso-N-acetyl-L-cysteine	S-nitrosothiol	N-ac-CysNO
S-nitroso-N-acetyl-DL-penicillamine	S-nitrosothiol	SNAP
3-morpholinosydnonimine	Sydnonimine	SIN-1

V. Recommended Nomenclature and Abbreviations

In the research literature of the past few years regarding the biological actions of NO, several different names or abbreviations have been used by investigators to identify the same enzyme or the same substance. To avoid confusion arising from nonuniform nomenclature and abbreviations, the following recommendations are made (see sections V.A to V.E.). These reflect current usage and, as such, contain anomalies. In the future, new compounds will be named and abbreviated with greater consistency.

A. Abbreviations for the Isoenzymes of Nitric Oxide Synthase

The abbreviations for the isoenzymes of the NOS are:

- eNOS for the isoform originally found in endothelial cells;
- nNOS for that originally found in neuronal tissue; and
- iNOS for the isoform induced in macrophages and other cell types in response to endotoxin and/or various cytokines.

B. Abbreviations for Nitric Oxide Synthase Substrates

The recommended abbreviations for NOS substrates are as follows:

- L-arginine: full name or L-Arg;
- L-homoarginine: full name or L-homoArg; and
- N^ω-hydroxy-L-arginine: L-OHArg.

C. Abbreviations for Inhibitors of Nitric Oxide Synthase

The recommended abbreviations for inhibitors of NOS are as follows:

- N^ω-monomethyl-L-arginine: L-NMMA;
- N^ω-nitro-L-arginine: L-NA;
- N^ω-nitro-L-arginine methyl ester: L-NAME;
- N-iminoethyl-L-ornithine: L-NIO;
- N^ω-amino-L-arginine: L-NAA;
- N^ω-N^{ω'}-dimethyl-L-arginine: L-ADMA;
- N^ω-N^{ω'}-dimethyl-L-arginine: L-SDMA;
- L-canavanine: full name;
- aminoguanidine: full name; and
- 7-nitroindazole: 7-NI.

The use of the L-isomer should be specified.

Other compounds have recently been described as inhibitors of NOS but are not in common use. These include the isothioureas (Garvey et al., 1994; Southan et al., 1995) and L-thiocitrulline (Frey et al., 1994).

D. Abbreviations for Certain Nitric Oxide Donors

The recommended abbreviation for certain NO donors are shown in table 4.

E. Nitrergic Nerves

The term 'nitrergic' should be applied to nerves whose transmitter function depends on the release of NO or to transmission mechanisms that are brought about by NO.

VI. Conclusion

Since the identification in 1987 of NO as a biological mediator, more than 14,000 papers have been published in this field. Some basic guidelines for standardization of terminology have been given in this short document. As the amount of work in this area continues to grow, it will be necessary to update and expand these recommendations.

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REFERENCES

- BAYLIS, S. A., WEINER, C. P., MONCADA, S., AND CHARLES, I. G.: In vivo expression of inducible nitric oxide synthase in the human fetus (abstract 3). Presented at the First International Conference on Biochemistry and Molecular Biology of Nitric Oxide, Los Angeles, California, 1994, p. 66.
- BECKMAN, J. S., BECKMAN, T. W., CHEN, J., MARSHALL, P. A., AND FREEMAN, B. A.: Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proc. Natl. Acad. Sci. USA* **87**: 1620–1624, 1990.
- BUTLER, A. R., FLITNEY, F. W., AND WILLIAMS, D. L. H.: NO, nitrosonium ions, nitroxide ions, nitrosothiols and iron-nitrosyls in biology: a chemist's perspective. *Trends Pharmacol. Sci.* **16**: 18–22, 1995.
- CHARLES, I. G., PALMER, R. M. J., HICKERY, M. S., BAYLISS, M. T., CHUBB, A. P., HALL, V. S., MOSS, D. W., AND MONCADA, S.: Cloning, characterization, and expression of a cDNA encoding an inducible nitric oxide synthase from the human chondrocyte. *Proc. Natl. Acad. Sci. USA* **90**: 11419–11423, 1993.
- DE BELDER, A., LEES, C., MARTIN, J., MONCADA, S., AND CAMPBELL, S.: Treatment of HELLP syndrome with nitric oxide donor. *Lancet* **345**: 124–125, 1995.
- DINERMAN, J. L., DAWSON, T. M., SCHELL, M. J., SNOWMAN, A., AND SNYDER, S. H.: Endothelial nitric oxide synthase localized to hippocampal pyramidal cells: implications for synaptic plasticity. *Proc. Natl. Acad. Sci. USA* **91**: 4214–4218, 1994.
- FEELISCH, M.: The biochemical pathways of nitric oxide formation from nitrovasodilators: appropriate choice of exogenous NO donors and aspects of preparation and handling of aqueous NO solutions. *J. Cardiovasc. Pharmacol.* **17**(suppl. 3): S25–S33, 1991.
- FEELISCH, M., AND STAMLER, J. S.: Donors of nitrogen oxides. *In Methods in Nitric Oxide Research*, ed. by M. Feelisch and J. S. Stamler, pp. 71–115, Wiley and Sons Ltd., New York, 1996.
- FREY, C., NARAYANAN, K., MCMILLAN, K., SPACK, L., GROSS, S. S., MASTERS, B. S., AND GRIFFITH, O. W.: L-thiocitrulline: a stereospecific, heme-binding inhibitor of nitric oxide synthases. *J. Biol. Chem.* **269**: 26083–26091, 1994.
- FURCHGOTT, R. F.: Studies on endothelium-dependent vasodilation and the endothelium-derived relaxing factor. *Acta. Physiol. Scand.* **139**: 257–270, 1990.
- FURCHGOTT, R. F., AND ZAWADZKI, J. V.: The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature (Lond.)* **288**: 373–376, 1980.
- GARTHWAITE, J.: Glutamate, nitric oxide and cell-cell signalling in the nervous system. *Trends Neurosci.* **14**: 60–67, 1991.
- GARVEY, E. P., OPLINGER, J. A., TANOURY, G. J., SHERMAN, P. A., FOWLER, M., MARSHALL, S., HARMON, M. F., PAITH, J. E., AND FURFINE, E. S.: Potent and selective inhibition of human nitric oxide synthases: inhibition by non-amino acid isothioureas. *J. Biol. Chem.* **269**: 26669–26676, 1994.
- GELLER, D. A., NUSSLER, A. K., DI-SILVIO, M., LOWENSTEIN, C. J., SHAPIRO, R. A., WANG, S. C., SIMMONS, R. L., AND BILLIAR, T. R.: Cytokines, endotoxin, and glucocorticoids regulate the expression of inducible nitric oxide synthase in hepatocytes. *Proc. Natl. Acad. Sci. USA* **90**: 522–526, 1993.
- GILLESPIE, J. S., LIU, X., AND MARTIN, W.: The neurotransmitter of the non-adrenergic non-cholinergic inhibitory nerves to smooth muscle of the genital system. *In Nitric Oxide from L-Arginine: A Bioregulatory System*, ed. by S. Moncada and E. A. Higgs, pp. 147–164, Elsevier Science Publishers B. V., Amsterdam, 1990.
- GRIFFITH, O. W., AND GROSS, S. S.: Inhibitors of nitric oxide synthases. *In Methods in Nitric Oxide Research*, ed. by M. Feelisch and J. S. Stamler, pp. 187–220, Wiley and Sons Ltd., New York, 1996.
- HIBBS, J. B. JR.: Overview of cytotoxic mechanisms and defence of the intracellular environment against microbes. *In The Biology of Nitric Oxide: Enzymology, Biochemistry and Immunology*, vol. 2, ed. by S. Moncada, M. A. Marletta, J. B. Hibbs Jr., and E. A. Higgs, pp. 201–206, Portland Press, London, 1992.
- HIBBS, J. B. JR., TAINTOR, R. R., VAVRIN, Z., AND RACHLIN, E. M.: Nitric oxide: a cytotoxic activated macrophage effector molecule. *Biochem. Biophys. Res. Commun.* **157**: 87–94, 1988.
- HIBBS, J. B. JR., TAINTOR, R. R., VAVRIN, Z., GRANGER, D. L., DRAPIER, J. -C., AMBER, I. J., AND LANCASTER, J. R. JR.: Synthesis of nitric oxide from a terminal guanidino nitrogen atom of L-arginine: a molecular mechanism regulating cellular proliferation that targets intracellular iron. *In Nitric Oxide from L-Arginine: A Bioregulatory System*, ed. by S. Moncada and E. A. Higgs, pp. 189–223, Elsevier, Amsterdam, 1990.
- HUANG, P. L., AND FISHMAN, M. C.: Genetic analysis of nitric oxide synthase isoforms: targeted mutation in mice. *J. Mol. Med.* **74**: 415–421, 1996.
- IGNARRO, L. J.: Biosynthesis and metabolism of endothelium-derived nitric oxide. *Ann. Rev. Pharmacol. Toxicol.* **30**: 535–560, 1990.
- IGNARRO, L. J.: Heme-dependent activation of guanylate cyclase by nitric oxide: a novel signal transduction mechanism. *Blood Vessels* **28**: 67–73, 1991.
- IGNARRO, L. J., BUGA, G. M., WOOD, K. S., BYRNS, R. E., AND CHAUDHURI, G.: Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proc. Natl. Acad. Sci. USA* **84**: 9265–9269, 1987.
- KHAN, M. T., AND FURCHGOTT, R. F.: Additional evidence that endothelium-derived relaxing factor is nitric oxide. *In Pharmacology*, ed. by M. J. Rand and C. Raper, pp. 341–344, Elsevier, Amsterdam, 1987.
- KNOWLES, R. G., AND MONCADA, S.: Nitric oxide synthases in mammals. *Biochem. J.* **298**: 249–258, 1994.
- KOZBIK, L., BREDT, D. S., LOWENSTEIN, C. J., DRAZEN, J., GASTON, B., SUGARBAKER, D., AND STAMLER, J. S.: Nitric oxide synthase in human and rat lung: immunocytochemical and histochemical localization. *Am. J. Respir. Cell Mol. Biol.* **9**: 371–377, 1993.
- KOZBIK, L., REID, M. B., BREDT, D. S., AND STAMLER, J. S.: Nitric oxide in skeletal muscle. *Nature (Lond.)* **372**: 546–548, 1994.
- LEFER, A. M., AND LEFER, D. J.: Therapeutic role of nitric oxide donors in the treatment of cardiovascular disease. *Drugs Future* **19**: 665–672, 1994.
- LUSCHER, T. F.: Endothelium-derived nitric oxide: the endogenous nitrovasodilator in the human cardiovascular system. *Eur. Heart J.* **12**(suppl. E): E2–E11, 1991.
- MARLETTA, M. A., YOON, P. S., IYENGAR, R., LEAF, C. D., AND WISHNOK, J. S.: Macrophage oxidation of L-arginine to nitrite and nitrate: nitric oxide is an intermediate. *Biochemistry* **27**: 8706–8711, 1988.
- MARSDEN, P. A., SCHAPPERT, K. T., CHEN, H. S., FLOWERS, M., SUNDELL, C. L., WILCOX, J. N., LAMAS, S., AND MICHEL, T.: Molecular cloning and characterization of human endothelial nitric oxide synthase. *FEBS Lett.* **307**: 287–293, 1992.
- MOHAUPT, M. G., ELZIE, J. L., AHN, K. Y., CLAPP, W. L., WILCOX, C. S., AND KONE, B. C.: Differential expression and induction of mRNAs encoding two inducible nitric oxide synthases in rat kidney. *Kidney Int.* **46**: 653–665, 1994.
- MONCADA, S., AND HIGGS, E. A.: The L-arginine-nitric oxide pathway. *N. Engl. J. Med.* **329**: 2002–2012, 1993.
- MONCADA, S., AND HIGGS, E. A.: Molecular mechanisms and therapeutic strategies related to nitric oxide. *FASEB J.* **9**: 1319–1330, 1995.
- MONCADA, S., PALMER, R. M. J., AND HIGGS, E. A.: Biosynthesis of nitric oxide from L-arginine: a pathway for the regulation of cell function and communication. *Biochem. Pharmacol.* **38**: 1709–1715, 1989.
- MORO, M. A., DARLEY-USMAR, V. M., GOODWIN, D. A., READ, N. G., ZAMORAPINO, R., FEELISCH, M., RADOMSKI, M. W., AND MONCADA, S.: Paradoxical fate and biological action of peroxynitrite on human platelets. *Proc. Natl. Acad. Sci. USA* **91**: 6702–6706, 1994.
- MORRIS, S. M. JR., AND BILLIAR, T. R.: New insights into the regulation of inducible nitric oxide synthesis. *Am. J. Physiol.* **266**: E829–E839, 1994.
- MURAD, F., ISHII, K., FÖRSTERMANN, U., GORSKY, L., KERWIN, J. F. JR., POLLOCK, J., AND HELLER, M.: EDRF is an intracellular second messenger and autacoid to regulate cyclic GMP synthesis in many cells. *Adv. Second Messenger Phosphoprotein Res.* **24**: 441–448, 1990.
- NAKANE, M., SCHMIDT, H. H. W., POLLOCK, J. S., FÖRSTERMANN, U., AND MURAD, F.: Cloned human brain nitric oxide synthase is highly expressed in skeletal muscle. *FEBS Lett.* **316**: 175–180, 1993.
- NATHAN, C. F., AND HIBBS, J. B. JR.: Role of nitric oxide synthesis in macrophage antimicrobial activity. *Curr. Opin. Immunol.* **3**: 65–70, 1991.
- NATHAN, C., AND XIE, Q. -W.: Regulation of biosynthesis of nitric oxide. *J. Biol. Chem.* **269**: 13725–13728, 1994.
- NGUYEN, T., BRUNSON, D., CRESPI, C. L., PENMAN, B. W., WISHNOK, J. S., AND TANNENBAUM, S. R.: DNA damage and mutation in human cells exposed to nitric oxide in vitro. *Proc. Natl. Acad. Sci. USA* **89**: 3030–3034, 1992.
- NUSSLER, A. K., AND BILLIAR, T. R.: Inflammation, immunoregulation and inducible nitric oxide synthase. *J. Leukoc. Biol.* **54**: 171–178, 1993.
- PALMER, R. M. J., FERRIGE, A. G., AND MONCADA, S.: Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature (Lond.)* **327**: 524–526, 1987.
- PUFAHL, R. A., NANJAPPAN, P. G., WOODARD, R. W., AND MARLETTA, M. A.: Mechanistic probes of N-hydroxylation of L-arginine by the inducible nitric

- oxide synthase from murine macrophages. *Biochemistry* **31**: 6822–6828, 1992.
- RADOMSKI, M. W., PALMER, R. M. J., AND MONCADA, S.: An L-arginine/nitric oxide pathway present in human platelets regulates aggregation. *Proc. Natl. Acad. Sci. USA* **87**: 5193–5197, 1990.
- RAND, M. J.: Nitrgic transmission: nitric oxide as a mediator of non-adrenergic, non-cholinergic neuro-effector transmission. *Clin. Exp. Pharmacol. Physiol.* **19**: 147–169, 1992.
- SESSA, W. C.: The nitric oxide synthase family of proteins. *J. Vasc. Res.* **31**: 131–143, 1994.
- SESSA, W. C., PRITCHARD, K., SEYEDI, N., WANG, J., AND HINTZE, T. H.: Chronic exercise in dogs increases coronary vascular nitric oxide production and endothelial cell nitric oxide synthase gene expression. *Circ. Res.* **74**: 349–353, 1994.
- SNYDER, S. H., AND BREDT, D. S.: Biological roles of nitric oxide. *Sci. Am.* **266**: 68–71, 1992.
- SOUTHAN, G. J., SZABO, C., AND THIEMERMANN, C.: Isothioureas: potent inhibitors of nitric oxide synthases with variable isoform selectivity. *Br. J. Pharmacol.* **114**: 510–516, 1995.
- STUEHR, D. J., AND GRIFFITH, O. W.: Mammalian nitric oxide synthases. *Adv. Enzymol. Relat. Areas Mol. Biol.* **65**: 287–346, 1992.
- STUEHR, D., GROSS, S., SAKUMA, I., LEVI, R., AND NATHAN, C.: Activated murine macrophages secrete a metabolite of arginine with the bioactivity of endothelium-derived relaxing factor and the chemical reactivity of nitric oxide. *J. Exp. Med.* **169**: 1011–1020, 1989.
- TODA, N.: Nitric oxide and the regulation of cerebral arterial tone. *In Nitric Oxide in the Nervous System*, ed. by S. Vincent, pp. 207–225, Academic Press Ltd., Orlando, 1995.
- VALLANCE, P., LEONE, A., CALVER, A., COLLIER, J., AND MONCADA, S.: Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *Lancet* **339**: 572–575, 1992.
- VANE, J. R., ANGGARD, E. E., AND BOTTING, R. M.: Regulatory functions of the vascular endothelium. *N. Engl. J. Med.* **323**: 27–36, 1990.
- VANHOUTTE, P. M.: Endothelium and control of vascular function. *Hypertension* **13**: 658–667, 1989.
- WEINER, C. P., LIZASOAIN, I., BAYLIS, S. A., KNOWLES, R. G., CHARLES, I. G., AND MONCADA, S.: Induction of calcium-dependent nitric oxide synthases by sex hormones. *Proc. Natl. Acad. Sci. USA* **91**: 5212–5216, 1994.
- WILLIAMS, R. J. P.: Nitric oxide in biology: its role as a ligand. *Chem. Soc. Rev.* **25**: 77–83, 1996.