

# International Union of Basic and Clinical Pharmacology. LXXXIII: Classification of Prostanoid Receptors, Updating 15 Years of Progress<sup>S</sup>

D. F. Woodward, R. L. Jones, and S. Narumiya

*Department of Biological Sciences, Allergan Inc., Irvine, California (D.F.W.); Strathclyde Institute of Pharmacy & Biomedical Sciences, University of Strathclyde, Glasgow, Scotland, United Kingdom (R.L.J.); and Department of Pharmacology, Kyoto University Faculty of Medicine, Yoshida, Kyoto, Japan (S.N.)*

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Address correspondence to: Dr. David F. Woodward, Dept. of Biological Sciences RD3-2B, Allergan, Inc., 2525 Dupont Dr., Irvine, CA 92612. E-mail: woodward\_david@allergan.com

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**Abstract**—It is now more than 15 years since the molecular structures of the major prostanoid receptors were elucidated. Since then, substantial progress has been achieved with respect to distribution and function, signal transduction mechanisms, and the design of agonists and antagonists (<http://www.iuphar-db.org/DATABASE/FamilyIntroductionForward?familyId=58>). This review systematically details these advances. More recent developments in prostanoid receptor research are included. The DP<sub>2</sub> receptor, also termed CRTH2, has little structural resemblance to DP<sub>1</sub> and other receptors described in the original prostanoid receptor classifica-

tion. DP<sub>2</sub> receptors are more closely related to chemoattractant receptors. Prostanoid receptors have also been found to heterodimerize with other prostanoid receptor subtypes and nonprostanoids. This may extend signal transduction pathways and create new ligand recognition sites: prostacyclin/thromboxane A<sub>2</sub> heterodimeric receptors for 8-*epi*-prostaglandin E<sub>2</sub>, wild-type/alternative (alt4) heterodimers for the prostaglandin FP receptor for bimatoprost and the prostamides. It is anticipated that the 15 years of research progress described herein will lead to novel therapeutic entities.

## I. Introduction

### A. Receptor Classification (circa 1994)

**1. Receptor Subtypes.** The major prostaglandins (PGs<sup>1</sup>), PGD<sub>2</sub>, PGE<sub>2</sub>, PGF<sub>2α</sub>, prostacyclin (PGI<sub>2</sub>), and thromboxane A<sub>2</sub> (TxA<sub>2</sub>) preferentially interact with dedicated receptors designated DP, EP, FP, IP, and TP, respectively

(Kennedy et al., 1982; Coleman et al., 1984). Although largely based on functional studies using a limited range of agonists, and an even more limited range of antagonists, the original classification of prostanoid receptors has entirely withstood the tests of time and scrutiny. Prostanoid receptor subtypes were also proposed. Four subtypes of EP receptor (EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub>, and EP<sub>4</sub>) have

been described (Coleman et al., 1994b). Two PGD<sub>2</sub>-sensitive receptors were suggested, but only DP<sub>1</sub> was described in 1994, although diverse pharmacological evidence described a second PGD<sub>2</sub>-sensitive receptor (Jones, 1976a,b, 1978; Narumiya and Toda, 1985; Woodward et al., 1990, 1993b; Rangachari and Betti, 1993; Fernandes and Crankshaw, 1995).

<sup>1</sup>Abbreviations: AH 13205, *trans*-2-(4-(1-hydroxyhexyl)phenyl)-5-oxocyclopentaneheptanoic acid; AH 23848, 7-(5-(((1,1-biphenyl)-4-yl)methoxy)-2-(4-morpholinyl)-3-oxocyclopentyl)-4-heptanoic acid; Akt, protein kinase B; AL-6556, 9-chloro-15-cyclohexyl-11,15-dihydroxy-3-oxa-16,17,18,19,20-pentanoic acid; ApoE, apolipoprotein E; BAL, bronchoalveolar lavage; Bay U 3405, ramatroban; BM-13177, sulotroban; BM-13505, daltroban; BMS-180291, ifetroban; BNP, brain natriuretic protein; BW 245C, 5-(6-carboxyhexyl)-1-(3-cyclohexyl-3-hydroxypropyl)hydantoin; BW A868C, 3-(3-[1,1'-biphenyl]-4-yl-3-hydroxypropyl)-2,5-dioxo-4-imidazolidineheptanoic acid, ethyl ester; CAY-10399, "2-series" analog of butaprost-FA; CGRP, calcitonin gene-related peptide; CJ-023,423, *N*-((2-(4-(2-ethyl-4,6-dimethyl-1*H*-imidazo(4,5-*c*)pyridin-1-yl)phenyl)ethyl)amino)carbonyl)-4-methylbenzenesulfonamide; CJ-42794, (S)-4-(1-(5-chloro-2-(4-fluorophenoxy)benzamido)ethyl)benzoic acid; CNS, central nervous system; COX, cyclooxygenase; CRTH2, chemoattractant receptor-homologous molecule expressed on Th2 cells; Cyr 61, cysteine-rich angiogenic protein 61; DC, dendritic cell; DNFB, dinitrofluorobenzene; EGF, epidermal growth factor; Epacs, exchange proteins activated by cAMP; ERK, extracellular signal regulated protein kinase; GR 63799X, (4-benzamidophenyl)-(Z)-[1*R*,2*R*,3*R*]-3-hydroxy-2-[(2*R*)-2-hydroxy-3-phenoxypropoxy]-5-oxocyclopentyl]hept-5-enoate; GR-32191, vapiprost; GRK, G protein-coupled receptor kinase; HIF, hypoxia-inducible factor; HPGDS, hematopoietic prostaglandin D synthase; I-BOP, [1*S*-[1*α*,2*α*(Z),3*β*(1*E*,3*S*<sup>\*</sup>),4*α*]-7-[3-[3-hydroxy-4-(4-iodophenoxy)-1-butenyl]-7-oxabi-cyclo[2.2.1]hept-2-yl]5-heptenoic acid; ICI-192605, 6-(2-(2-chlorophenyl-4-hydroxyphenyl)-1,3-dioxan-5-yl)hexenoic acid; ICI-80205, 16-*p*-chlorophenoxy-*ω*-tetranor PGE<sub>2</sub>; ICI-81008, fluprostenol; IFN, interferon; IL, interleukin; IOP, intraocular pressure; iP, isoprostane; KO, knockout; KP-496, (2-(*N*-(4-(4-chlorobenzenesulfonylamino)butyl)-*N*-(3-(4-isopropylthiazol-2-yl)methoxy)benzyl)sulfamoyl)benzoic acid; L-888,607, (9-((4-chlorophenyl)thio)-6-fluoro-2,3-dihydro-1*H*-pyrrolo(1,2-*a*)indol-1-yl)acetic acid; LC, Langerhans cell; LPS, lipopolysaccharide; MAP, mitogen-activated protein; MB-28767, 11-deoxy-16-phenoxy PGE<sub>2</sub>; MEK, mitogen-activated protein kinase kinase; MF-266-1, 1-(5-(3-(2-(benzyloxy)-5-chlorophenyl)-2-thienyl)pyridin-3-yl)-2,2,2-trifluoroethane-1,1-diol; MK-0524, laropiprant; MLR, mixed lymphocyte reaction; NMDA, *N*-methyl-D-aspartate; ONO-11120, 11*α*-carba-12-(2'*S*-hydroxy-3'-phenyl-propylamino)-9*α*,11*α*-isopropylideno-*ω*-octanor-prost-5*Z*-enoic acid; ONO-8711, 6-((2*S*,3*S*)-3-(4-chloro-2-methylphenylsulfonamino)ethyl)-bicyclo(2.2.2)octan-2-yl)-5*Z*-hexenoic acid; OVA, ovalbumin; PF-04475270, 5-(3-(2-(3-hydroxy-4-(3-(trifluoromethyl)phenyl)butyl)-5-oxopyrrolidin-1-yl)propyl)thiophene-2-carboxylate; PG, prostaglandin; PGDS, prostaglandin D synthase; PGI<sub>2</sub>, prostacyclin; PI, phosphatidylinositol; PI3K, phosphoinositide-3-kinase; PKA, protein kinase A; PKC, protein kinase C; PLC, phospholipase C; PPAR, peroxisome proliferator-activated receptor; PTH, parathyroid hormone; RANKL, receptor activator of nuclear factor-*κ*B ligand; rc, recombinant; Rho, rhodopsin; S-145, 5,7-(3-phenylsulfonylamino(2.2.1)bicyclohept-2-yl)heptenoic acid; S-18886, terutroban; SC-19220, 8-chloro-dibenzo(Z)[b,f][1,4]oxazepine-10(11*H*)-carboxylic acid, 2-acetylhydrazide; SQ-29548, 7-(3-((2-((phenylamino)carbonyl)hydrazino)methyl)-7-oxabicyclo(2.2.1)hept-2-yl)-5-heptenoic acid; STAT, signal transducer and activator of transcription; TGF, transforming growth factor; Th, T helper; THG-113.31, Ile-Leu-Gly-His-(D-Cit)-Asp-Tyr-Lys; TM, transmembrane domain; TNF, tumor necrosis factor; TR4979, butaprost; TS-022, (4-((1*R*,2*S*,3*R*,5*R*)-5-chloro-2-((S)-3-cyclohexyl-3-hydroxyprop-1-ynyl)-3-hydroxycyclopentyl)butylthio) acetic acid monohydrate; TxA<sub>2</sub>, thromboxane A<sub>2</sub>; U-46619, 9-11-dideoxy-11*α*,9*α*-epoxymethanoprostaglandin F<sub>2α</sub>; VEGF, vascular endothelial growth factor; WT, wild type.

**2. Molecular Structure.** Prostanoid receptors are G protein-coupled receptors. Although the overall homology between those receptors cloned in the 1990s was not high, there were several conserved regions. The first prostanoid receptor to be structurally identified was TP. This was achieved by purifying the TP receptor protein using the high-affinity, radiolabeled ligand S-145 (Ushikubi et al., 1989). Based on a partial amino acid sequence, the cDNA for the TP receptor was obtained (Hirata et al., 1991). The other prostanoid receptors were cloned by homology based screening and by 1994, DP<sub>1</sub>, FP, IP, and all PGE<sub>2</sub>-sensitive receptors had been structurally identified (Coleman et al., 1994b; Hirata et al., 1994; Nakagawa et al., 1994; Regan et al., 1994b).

The deduced amino acid sequences for human DP<sub>1</sub>, EP<sub>1-4</sub>, FP, IP, and TP receptors, together with DP<sub>2</sub>, are compared in Fig. 1. Hydrophobicity analysis of the sequences indicated seven membrane-spanning segments, an extracellular N terminus, and an intracellular -COOH terminus typical of rhodopsin-type, G-protein-coupled receptors. Regions of significant homology occur in the seventh transmembrane domain and the second extracellular loop. The highly conserved arginine in the seventh transmembrane (TM) domain has been proposed as the interaction site for the carboxylate group, which is common to all natural prostanoids. Additional determinants of prostanoid receptor binding have been suggested as follows. Two conserved Cys residues in the first and second extracellular loops are believed to form a disulfide bridge critical for stabilization of the receptor conformation (Narumiya et al., 1999). The second extracellular loop connecting TM4 and TM5 contains an invariant Trp-Cys-Phe triplet (Pierce et al., 1995). There are one or more consensus sequences for N-glycosylation of arginine residues in the amino terminal region (Narumiya et al., 1999). N-glycosylation may also be important for ligand binding, at least for TP receptors (Chiang and Tai, 1998).

Studies on the molecular evolution of prostanoid receptors suggest a PGE<sub>2</sub>-sensitive entity as the ancestral receptor (Regan et al., 1994b; Boie et al., 1995; Toh et al., 1995; Foord et al., 1996). This may have occurred not only by gene duplication but also by chromosomal duplication (Duncan et al., 1995). These phylogenetic analyses suggested two major branches of prostanoid receptor evolution: one group G<sub>s</sub>-coupled (DP<sub>1</sub>, EP<sub>2</sub>, EP<sub>4</sub>, and IP) and the other group G<sub>i</sub>- (EP<sub>3</sub>) or G<sub>q</sub>-coupled (EP<sub>1</sub>, FP, and TP), providing three clusters (Table 1). For most receptors, mRNA splicing variants have been identified.

The DP<sub>2</sub> receptor, although recognizing PGD<sub>2</sub> as a primary natural ligand, has no significant homology with DP<sub>1</sub> or other prostanoid receptors described in the original classification (Abe et al., 1999; Hirai et al., 2001; Hata et al., 2005). DP<sub>2</sub> (CRTH2) is more closely related to chemoattractant receptors, such as C<sub>5α</sub> and *N*-formyl peptide receptors (Abe et al., 1999; Hirai et al., 2001). The arginine residue in the seventh membrane-

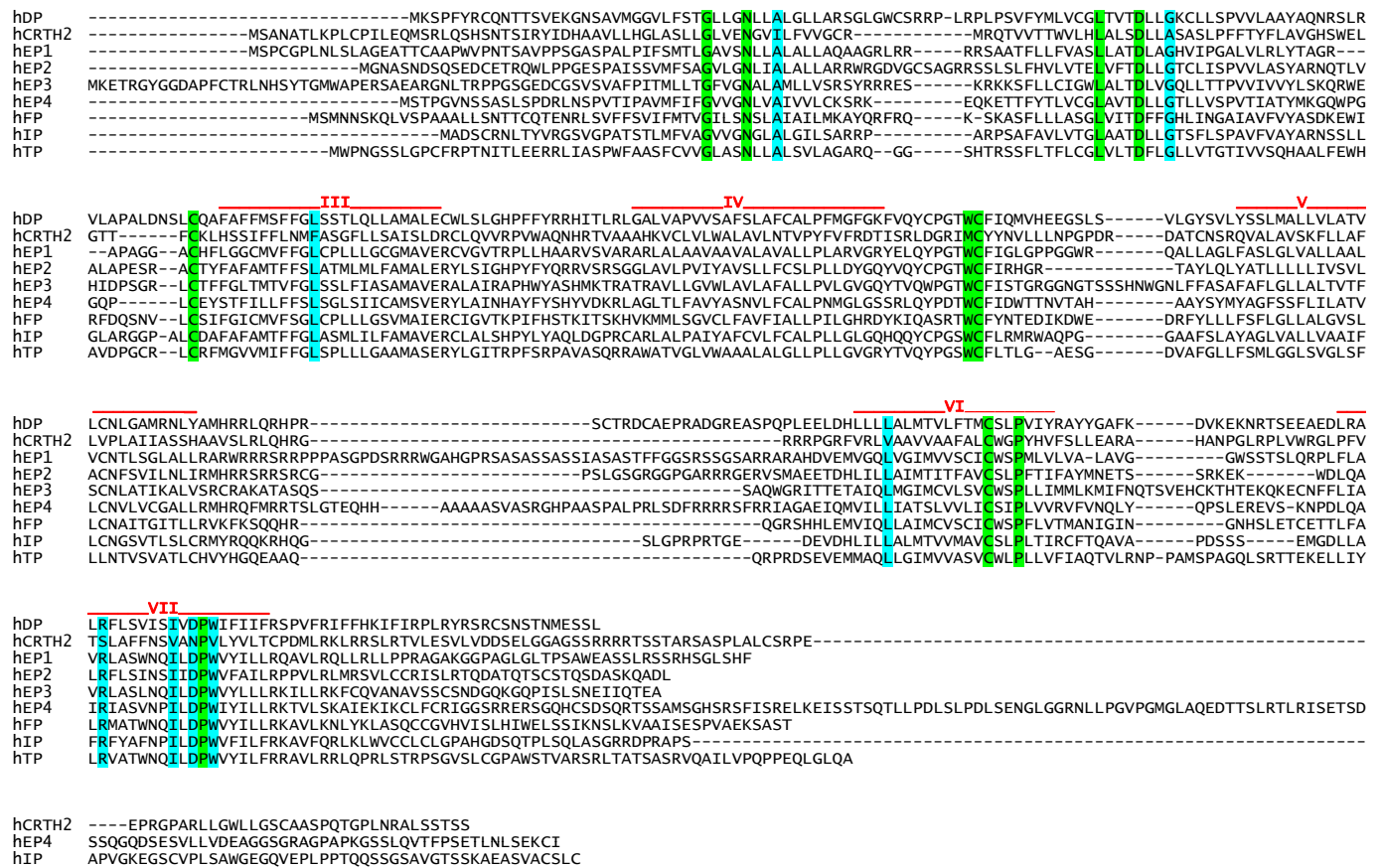


FIG. 1. Deduced amino sequence of human prostanoid receptors, in alignment. Amino acid residues with 100% homology between all receptors are highlighted in green. Amino acid residues with complete homology, except for DP<sub>2</sub>, are highlighted in blue.

spanning domain, which is conserved in all receptors of the original classification, is not present in DP<sub>2</sub> (Fig. 1). Replacement of the corresponding serine by mutation to alanine had little effect on PGD<sub>2</sub> binding (Hata et al., 2005). In contrast, replacement of lysine 209 in the fifth membrane-spanning domain by alanine greatly reduced PGD<sub>2</sub> binding to DP<sub>2</sub> receptors. Interaction of the carboxylate of PGD<sub>2</sub> with Lys-209 would place PGD<sub>2</sub> in an opposite orientation in the binding pocket to that proposed for other prostanoid receptors (Hata et al., 2005).

**3. Second Messenger Signaling.** Early signal transduction studies and a paucity of receptor selective ligands provided little insight into the pharmacology of prostanoid-sensitive receptors. The cloning of each prostanoid receptor and their transfection into and expres-

sion in cultured cells led to rapid elucidation of their G protein coupling characteristics, at least with respect to G<sub>q</sub>, G<sub>s</sub>, and G<sub>i</sub>. DP<sub>(1)</sub>, EP<sub>2</sub>, EP<sub>4</sub>, and IP receptors were classified as G<sub>s</sub>-coupled; EP<sub>1</sub>, FP, and TP receptors were designated as G<sub>q</sub>-coupled; EP<sub>3</sub> receptors seemed capable of coupling to both G<sub>q</sub> and G<sub>i</sub> (Coleman et al., 1994b). During the past decade and a half, the DP<sub>2</sub> (CRTH2) receptor was discovered as a second G<sub>i</sub>-coupled receptor. These findings are summarized in Table 1. The signaling cascades associated with prostanoid receptor stimulation have been further investigated to reveal diverse pathways.

**4. Agonist and Antagonist Drugs.** The pharmacological characterization of prostanoid receptors was built on isolated tissue and cultured cell studies (Coleman et al., 1984). Despite an inherent reliance on pharmacological intuition and deduction and on low-throughput assays, highly selective ligands were obtained for certain receptors. 5-(6-Carboxyhexyl)-1-(3-cyclohexyl-3-hydroxypropyl)hydantoin (BW 245C) and 3-(3-[1,1'-biphenyl]-4-yl-3-hydroxypropyl)-2,5-dioxo-4-imidazolidineheptanoic acid, ethyl ester (BW A868C) (Giles et al., 1989) were discovered as a selective agonist and antagonist, respectively, for DP<sub>1</sub> receptors, previously designated DP receptors. *trans*-2-(4-(1-Hydroxyhexyl)phenyl)-5-oxocyclopentaneheptanoic acid (AH 13205) (Nials et al., 1993)

TABLE 1  
Primary G protein coupling for prostanoid receptors

Prostanoid Receptor Subtype	Prostanoid Molecular Evolution Cluster	G Protein	Second Messenger
DP <sub>1</sub>	1	G <sub>s</sub>	↑ cAMP
EP <sub>1</sub>	2	G <sub>s</sub>	↑ [Ca <sup>2+</sup> ] <sub>i</sub>
EP <sub>2</sub>	1	G <sub>s</sub>	↑ cAMP
EP <sub>3</sub>	3	G <sub>i</sub>	↓ cAMP
EP <sub>4</sub>	1	G <sub>s</sub>	↑ cAMP
FP	2	G <sub>q</sub>	↑ [Ca <sup>2+</sup> ] <sub>i</sub> , PI turnover
IP	1	G <sub>s</sub>	↑ cAMP
TP	2	G <sub>q</sub>	↑ [Ca <sup>2+</sup> ] <sub>i</sub> , PI turnover



and butaprost (Gardiner, 1986) were selective EP<sub>2</sub> agonists and (4-benzamidophenyl)-(Z)-7-[(1R,2R,3R)-3-hydroxy-2-[(2R)-2-hydroxy-3-phenoxypropoxy]-5-oxocyclopentyl]hept-5-enoate (GR 63799X) was a potent and selective EP<sub>3</sub> agonist (Bunce et al., 1991). Fluprostenol (Dukes et al., 1974; Coleman et al., 1984) and 17-phenyl PGF<sub>2α</sub> (Woodward et al., 1995a) were known as selective FP agonists and, ultimately, formed the structural platform for launching antiglaucoma drugs. Above all, several potent and selective TP receptor antagonists were invented based on their potential utility for treating cardiovascular disease. Indeed, S-145 was sufficiently potent and selective to enable isolation of the TP receptor protein (Ushikubi et al., 1989; Hirata et al., 1991).

## B. Summary of New Developments

**1. Prostaglandin D<sub>2</sub> (CRTh2) Receptors.** The eight prostanoid receptors described by the pharmacology-based classification were rapidly discovered by homology-based screening after structural identification of TP receptors. This made it unlikely that further similar receptors would emerge, and this has proved to be the case. More recently discovered prostanoid receptors are quite different. A second PGD<sub>2</sub>-sensitive receptor was long suggested by functional studies but, when discovered, was found to be structurally quite distinct (Hirai et al., 2001). The DP<sub>2</sub> receptor, also and originally designated (CRTh2), is more closely related to chemo-attractant receptors. The DP<sub>2</sub> receptor mediates effects that are opposed to those produced by DP<sub>1</sub> receptor stimulation in some instances. Given the lack of structural identity between DP<sub>1</sub> and DP<sub>2</sub> (CRTh2) receptors, it is not surprising that ligand recognition can markedly diverge. For example, the cyclooxygenase (COX) inhibitor indomethacin actually stimulates DP<sub>2</sub> (CRTh2) receptors (Hirai et al., 2002; Stubbs et al., 2002), albeit weakly.

**2. Prostanoid Receptor Heterodimerization.** The discovery that G protein-coupled receptors may heterodimerize has explained certain pharmacological anomalies. Receptor heterodimerization offers the option of creating novel binding sites without evolution of a dedicated encoding gene. It also provides a means of closely regulating the activity of local hormones simultaneously released by intimately combining their receptors. IP/TP receptor heterodimerization provides an example of both phenomena: 1) a recognition site for 8-*epi*-PGE<sub>2</sub> is created and 2) the pathological effects of TP are limited because cAMP levels are increased by TxA<sub>2</sub> activating the associated heterodimeric protein (Wilson et al., 2004).

The pharmacology of PGF<sub>2α</sub>-ethanolamide (prosta-mide F<sub>2α</sub>), and its analog bimatoprost, was elucidated by a longer and more traditional route. Numerous agonist studies suggested a pharmacological identity distinct from the classic prostanoid FP receptor; this was confirmed by the eventual discovery of selective antagonists (Woodward et al., 2008). All this led to the same place in that the bimatoprost recognition site was modeled by

cotransfecting the wild-type FP receptor and an alternative mRNA splicing variant thereof (Liang et al., 2008).

**3. Gene Deletion Studies.** Gene deletion studies with mice lacking each of the individual prostanoid receptors have enabled further elucidation of prostanoids in health and disease. Moreover, they have revealed important functions that had not been previously appreciated. The roles of prostanoids in inflammation and immune regulation provide a good example. It is now clearer that prostanoids exert both pro-inflammatory and anti-inflammatory effects and regulate gene expression in mesenchymal and epithelial cells at inflammatory sites (Narumiya, 2009). This self-regulatory role, with prostanoids exerting a dual role in a singular process, is often seen in immunology-based reactions. It had been held that prostanoids exerted very little role in immunity, but gene deletion studies revealed that prostanoids work at many levels in immune responses (Narumiya, 2003). A striking example of the contribution of gene deletion is the discovery of EP<sub>1</sub> receptor involvement in suppressing impulsive behavior in response to stress (Furuyashiki and Narumiya, 2009). These findings will provide novel direction from discovering additional prostanoid-based therapeutics.

Gene deletion studies are an important step forward in elucidating the role of prostanoids in physiology and pathophysiology, especially when viewed in the context of small-molecule-based research. There are potential pitfalls in using small molecules to determine the pharmacological basis of experimental disease and thereby to discover new therapeutic approaches. False-negative results may occur by virtue of inadequate bioavailability at the target tissue: metabolic disposition and pharmacokinetics are traditionally a terminal step in the drug discovery process that occurs before clinical evaluation. Given the very high attrition rate in drug discovery, false-positive animal model data are clearly commonplace. Seemingly beneficial effects in animal models of human disease may be misleading for several reasons. Off-target pharmacology can never be fully assessed, even by screens aimed at 400 to 500 targets. Drug-induced toxicity is rarely overt in acute animal models but is likely to produce misleading positive effects, certainly in pain and inflammation models. Indications of toxicity could be obtained from such living animal studies but this rarely seems to be the case. Blood pressure and heart rate could readily be monitored by tail cuff instrumentation. Circulating leukocyte viability could be determined by *in vitro* studies on inflammatory cells that routinely determine cell viability. Studies employing genetically modified mice are more reliable in that there can be confidence that there is 1) no hidden off-target pharmacology, 2) no toxicity of unknown origin, and 3) no bioavailability issue. Thus, the use of genetically modified mice provides a solution to certain drug discovery complications; species differences and potential compensatory mechanisms remain a concern. Fi-

nally, the choice of disease models to complement gene deletion studies is also an important consideration. Many animal models, for example LPS-induced uveitis (Caspi, 2006), seem to be models in search of a human disease to mimic.

**4. New Agonists and Antagonists.** The introduction of recombinant receptor technologies permitted high-throughput assays. This has resulted in selective and potent antagonists for all known prostanoid receptors, with the exception of EP<sub>2</sub>. Potent and selective agonists have now been discovered for all prostanoid receptors, with the possible exception of EP<sub>1</sub>. These pharmacological tools, with information provided by gene deletion studies, will result in therapies based on informed modulation of prostanoid-mediated events. Prostanoid-based therapeutics will realize its full potential.

Parallel to fitting ligands to known receptors, unexplained pharmacological characteristics associated with certain prostanoids have been successfully studied. The activities of certain "orphan" prostanoids, although not actually described as such, are now understood. The surprisingly high potency of the PGD<sub>2</sub> metabolites 13,14-dihydro 15-keto PGD<sub>2</sub> (Jones, 1976a,b; Jones and Wilson, 1978) and PGJ<sub>2</sub> (Woodward et al., 1990) was explained by the discovery of the DP<sub>2</sub> (CRTh2) receptor. Heterodimerization between IP and TP receptors provides a site for interaction with 8-*epi*-PGE<sub>2</sub> (Wilson et al., 2004). Likewise, PG-ethanolamides, and their analog bimatoprost, have a predilection for cells cotransfected with wild type and an alternative mRNA splicing variant of the FP receptor (Liang et al., 2008).

**5. Cell Signaling.** Elucidation of the cell signaling pathways associated with prostanoid receptors has provided an additional dimension to the body of information required to conceptualize therapeutic applications. In many instances, the repertoire of signaling pathways is now known to be quite expansive for some receptors. These will be described for each individual prostanoid receptor.

## II. Receptor Types, Subtypes, and mRNA Splicing Variants

### A. DP<sub>1</sub> Receptors

**1. Second Messenger Signaling.** DP<sub>1</sub> receptors are G<sub>s</sub>-coupled and stimulate cAMP formation (Gorman et al., 1977; Whittle et al., 1978; Schafer et al., 1979; Halushka et al., 1989; Goh and Nakajima, 1990; Hirata et al., 1994; Boie et al., 1995). Cells expressing DP<sub>1</sub> receptors also elicit an increase in [Ca<sup>2+</sup>]<sub>i</sub> when stimulated by PGD<sub>2</sub> or the selective DP<sub>1</sub> agonist BW 245C, but this is cAMP-dependent (Boie et al., 1995). This increase in [Ca<sup>2+</sup>]<sub>i</sub> may result from activation of protein kinase A (PKA) and subsequent involvement of L-type Ca<sup>2+</sup> channels and the ryanodine receptor (Zaccolo, 2009). This seems to be a minor pathway for transducing DP<sub>1</sub> receptor signaling, cAMP/PKA activation being almost invariably involved. No evidence for the involvement of ex-

change proteins activated by cAMP (Epacs) seems to have emerged.

**2. Distribution and Biological Functions.** Among the classic prostanoid receptors, DP<sub>1</sub> is the least abundant in tissues and exhibits a relatively narrow distribution. Northern blot analyses detected expression only in the retina and small intestine (Boie et al., 1995). In mice, DP<sub>1</sub> receptor expression is moderate in the ileum; weak in the stomach, lung, and uterus; and absent elsewhere (Hirata et al., 1994). Functional studies, however, have described DP<sub>1</sub> receptors present in certain cells. In some cases, transcripts have provided supportive data. Human platelets express functional DP<sub>1</sub> receptors, which inhibit aggregation (Whittle et al., 1983; Giles et al., 1989; Trist et al., 1989; Seiler et al., 1990). Effects of PGD<sub>2</sub> on vascular smooth muscle vary according to species (Giles and Leff, 1988). In humans, effects were restricted to facial flushing and nasal congestion (Heavey et al., 1984), with no meaningful alteration in blood pressure. Similar effects were apparent for the DP<sub>1</sub> agonist BW 245C (Giles and Leff, 1988). Thus, despite evidence for presynaptic DP receptors that would enhance norepinephrine release (Molderings et al., 1994), this does not seem to translate into gross cardiovascular events. In contrast, the skin flushing associated with nicotinic acid used for treating dyslipidemia is DP<sub>1</sub> receptor-mediated (Cheng et al., 2006).

Compelling pharmacological evidence for DP<sub>1</sub> receptors in the human myometrium has been advanced (Senior et al., 1992; Fernandes and Crankshaw, 1995). DP<sub>1</sub> receptor stimulation would mediate tocolysis. It is noteworthy that Northern blotting did not identify a DP<sub>1</sub> receptor transcript in the human uterus (Boie et al., 1995). There are additional examples of DP<sub>1</sub> receptors being identified in specific cell types or localized regions in which Northern blot analyses of tissue would suggest otherwise.

The initial studies on DP<sub>1</sub> receptor distribution showed undetectable to very low levels in the brains of both mice and humans (Hirata et al., 1994; Boie et al., 1995). Because PGD<sub>2</sub> has well documented activity in the CNS, these data were interpreted to mean that expression was limited to certain areas and/or specific cells (Narumiya et al., 1999). This has proven correct, when subject to careful examination, for not only the brain but also other tissues (Gerashchenko et al., 1998; Wright et al., 1998). A pharmacological rationale for the various central effects of PGD<sub>2</sub> and its analogs is therefore tenable. These effects include sleep regulation (Urade and Hayaishi, 1999), neuroprotection (Liang et al., 2005b; Saleem et al., 2007b; Thura et al., 2009) allodynia (Minami et al., 1997), and hyperalgesia (Telleria-Diaz et al., 2008). DP<sub>1</sub> receptor involvement in neurotransmission is not limited to the CNS, and a local effect on pruritus has been reported (Arai et al., 2004, 2007; Sugimoto et al., 2007). The pruritic activity associated with scratching, in an atopic dermatitis-like

model in NC/Nga mice, was inhibited by PGD<sub>2</sub> but not metabolites known to stimulate DP<sub>2</sub> receptors (Arai et al., 2004). These effects on experimental pruritus were confirmed using a DP<sub>1</sub> agonist, (4-((1*R*,2*S*,3*R*,5*R*)-5-chloro-2-((*S*)-3-cyclohexyl-3-hydroxyprop-1-ynyl)-3-hydroxycyclopentyl)butylthio) acetic acid monohydrate (TS-022), and the antagonist BW A868C to establish the pharmacological basis of the attenuated itch response (Arai et al., 2007). In addition, accelerated repair of the disrupted cutaneous barrier was reported for TS-022 (Arai et al., 2007), an effect ascribed not only to DP<sub>1</sub> but also to EP<sub>3</sub> and EP<sub>4</sub> receptors in mice (Honma et al., 2005). A physiological role for PGD<sub>2</sub> has also been contemplated, in that PGD<sub>2</sub> released by mechanical scratching may be autoinhibitory, limiting the extent of the scratching response and preventing skin damage (Sugimoto et al., 2007; Takaoka et al., 2007). The NC/Nga model may, at least in part, be operational with respect to the antipruritic effects of DP<sub>1</sub> agonists by virtue of a reduced level of endogenous PGD<sub>2</sub> production (Takaoka et al., 2007). The decrease in PGD<sub>2</sub> production in the late phase of dermatitis and scratching in NC/Nga mice, together with increased DP<sub>1</sub> receptor expression assists in explaining the potent antipruritic activity of TS-022 (Sugimoto et al., 2007). Related to atopic dermatitis, DP<sub>1</sub> receptor stimulation impedes TNF- $\alpha$ -induced migration of human Langerhans cells (LCs) and additional chemotactic events, which strongly decreases the recruitment of inflammatory cells in a model of murine atopic dermatitis (Angeli et al., 2004). The beneficial effects of TS-022 and BW 245C cannot be attributed solely to amelioration of pruritus by scratching behavior.

A study in mice also revealed that DP<sub>1</sub> receptor stimulation inhibited airway inflammation and suppressed asthma by modulating dendritic cells and inducing regulatory T cells (Hammad et al., 2007). PGD<sub>2</sub> also affects human dendritic cell differentiation and modulates the pattern of immunoregulatory cytokine production, favoring naive T cells toward a Th2 phenotype (Gosset et al., 2005). PGD<sub>2</sub> may contribute to the control of allergic reactions and tumor formation (Gosset et al., 2005; Torres et al., 2008).

DP<sub>1</sub> antagonists have also been the subject of anti-inflammatory studies. PGD<sub>2</sub> is the major prostanoid produced by mast cells; this presents an attractive target for DP-receptor drug design. Despite the evidence for DP<sub>1</sub> receptor-mediated pathological increases in blood flow and engorgement of blood vessels in the nasal mucosa, clinical trials on the DP<sub>1</sub> antagonist laropiprant demonstrated no efficacy in patients with allergic rhinitis or asthma (Philip et al., 2009). For therapeutic modalities based on attenuating the activity of PGD<sub>2</sub>, consideration of DP<sub>2</sub> (CRTh2)-mediated events is probably of greater importance. The significance of DP<sub>1</sub> receptor activation in inflammation and immune responses is best appreciated when considered in the context of DP<sub>2</sub> (CRTh2) receptors.

DP<sub>1</sub> receptor expression is high in the retina (Boie et al., 1995), but it could be argued that this finding has not been followed up to its full extent. PGD<sub>2</sub> induces heme oxygenase-1 expression in the retinal pigmented epithelium, an enzyme important for photoreceptor survival (Satarug et al., 2008). DP<sub>1</sub> receptors exerted only a marginal influence on heme oxygenase-1 expression, the DP<sub>2</sub> (CRTh2) receptor being important (Satarug et al., 2008). In addition to the retina, DP receptor expression occurs in the iris and ciliary body (Gerashchenko et al., 1998). Expression in the ciliary body entirely correlates with the ocular hypotensive activity of DP<sub>1</sub> agonists and their mechanism of action (i.e., increased uveoscleral outflow) (Toris et al., 2006). DP<sub>1</sub> receptor mRNA is also located in mucus-secreting goblet cells and the columnar epithelium of the rat gastrointestinal tract (Wright et al., 1998), but this does not transition into the eye, where goblet cell secretion is attributable to DP<sub>2</sub> receptor pharmacology (Woodward et al., 1990, 1993b). Likewise, the DP<sub>1</sub> receptor has been implicated in eosinophil trafficking (Schratl et al., 2007), but this was not observed in ocular studies (Woodward et al., 1990, 1993b). The most prominent effect of DP<sub>1</sub> receptor stimulation in the eye is on intraocular pressure (IOP) (Goh et al. 1988; Nakajima et al., 1991; Woodward et al., 1993b). DP<sub>1</sub> effects are summarized in Table 2 in the therapeutics section.

**3. Gene Deletion Studies.** One line of DP (DP<sub>1</sub>)-deficient mice has been generated (Matsuoka et al., 2000). Using this line of mice, physiological roles of DP<sub>1</sub> in allergy and immunity, sleep induction and other brain functions, and tumor progression and angiogenesis have been examined. Given that PGD<sub>2</sub> is a major PG produced by mast cells and released in large amount after antigen challenge, Matsuoka et al. (2000) examined its role in allergic inflammation using ovalbumin (OVA)-induced asthma model. Sensitization and aerosol challenge of DP(-/-) mice with OVA-induced increases in the serum concentration of IgE similar to those observed in wild-type mice. However, the DP(-/-) animals developed substantially reduced asthmatic responses in this model; the concentrations of Th2 cytokines and the extents of lymphocyte accumulation and eosinophil infiltration in the lungs of the mutant animals after OVA challenge were greatly reduced compared with those apparent in the wild type. These observations indicate that PGD<sub>2</sub> serves as a mediator of asthmatic responses. The authors found that DP<sub>1</sub> is induced in airway epithelial cells after the challenge and suggested that PGD<sub>2</sub> acts on epithelial cells to induce various allergy-associated genes to facilitate inflammation. On the other hand, Angeli et al. (2001), studying *Schistosoma mansoni* infection in the skin, found that parasite-derived PGD<sub>2</sub> acts at DP<sub>1</sub> receptors in LCs to induce their retention in the epidermis. They further showed that this retention can be mimicked by administration of a DP<sub>1</sub> agonist, BW 245C, during local TNF- $\alpha$  treatment and



TABLE 2  
*Potential therapeutic application of DP<sub>1</sub> agonists*

DP <sub>1</sub> Agonist	Route	Dose	Species	Experimental Model	Indication	Reference
572C85 192C86	Topical	2–250 $\mu$ g	Rabbit, cat	Intraocular pressure	Glaucoma	Matsugi et al., 1995
AL-6598	Topical	0.01%	Monkey	Intraocular pressure	Glaucoma	Toris et al., 2006
BW 245C	Intrathecal	83 ng/kg (IC <sub>50</sub> )	Mouse	Nociceptin-induced allodynia	Chronic pain	Minami et al., 1997
BW 245C	Topical	2.5 $\mu$ g	Rabbit	Intraocular pressure	Glaucoma	Goh et al., 1988
BW 245C	Topical	50 $\mu$ M	Mouse	Epicutaneous sensitization to OVA	Atopic dermatitis	Angeli et al., 2004
BW 245C	Intratracheal	100 $\mu$ M	Mouse	OVA-alum asthma	Asthma	Hammad et al., 2007
BW 245C		1–1000 nM	Rat	Cell/tissue culture	Neurodegenerative diseases	Liang et al., 2005b
BW 245C	In vitro <sup>+</sup> (–/– DP <sub>1</sub> mice)	0.05–5 $\mu$ M	Mouse	Ischemia-reperfusion injury	Cerebral ischemic disorders	Saleem et al., 2007b
BW 245C	Topical	0.01–1%	Rabbit	Intraocular pressure	Glaucoma	Woodward et al., 1990
BW 245C AS 702224	Intracavernosal	BW 245C, 59 nM; AS 702224, 29 nM (EC <sub>50</sub> in isolated human tissue)	Rabbit, rat, human cavernosal tissue	Penile tumescence	Erectile dysfunction	Brugger et al., 2008
SQ-27986	Topical	2.5–25 $\mu$ g	Rabbit	Intraocular pressure	Glaucoma	Woodward et al., 1993
SQ-27986	Topical	1–30 $\mu$ g	Monkey	Intraocular pressure	Glaucoma	Crawford et al., 1992
TS-022	Topical	2.5 → 250 nM	Mouse	NC/Nga mice	Atopic dermatitis	Arai et al., 2007

AL-6598, 2-[[[(2Z)-4-[(1R,2R,3R,5R)-5-chloro-2-[(3R)-3-cyclohexyl-3-hydroxypropyl]-3-hydroxycyclopentyl]-2-buten-1-yl]oxy]-1-methylethyl ester, acetic acid.

that the retention of LCs in the skin by *S. mansoni* infection or by BW 245C was impaired in DP<sub>1</sub>-deficient mice. In DP<sub>1</sub>-deficient mice, LCs migrated to draining lymph nodes and triggered a Th2 response (Hervé et al., 2003). Such inhibitory modulation of dendritic cell (DC) function by DP stimulation is also found in DCs in the lung. Hammad et al. (2007) found that inhalation of BW 245C in mice during OVA challenge attenuated the asthmatic response and that this inhibitory effect of BW 245C was dependent on DP<sub>1</sub> expression by lung DCs. They suggested that this DP<sub>1</sub>-dependent modulation of lung DCs operates in the actual process of allergic inflammation in the OVA-induced asthma response, because chimeric mice with DP(–/–) hematopoietic cells exhibited an enhanced airway response. Therefore, there is both enhancement and attenuation of PGD<sub>2</sub>-DP<sub>1</sub> signaling-dependent pathways by different types of cells in allergic inflammation, and the final outcome shown in the study by Matsuoka et al. (2000) seems to represent the net effects of these pathways. Given the sleep-inducing effect of PGD<sub>2</sub>, Mizoguchi et al. (2001) examined localization of DP<sub>1</sub> in the brain and examined whether DP<sub>1</sub> is involved in PGD<sub>2</sub>-mediated sleep. They found that DP<sub>1</sub> in the brain is mainly localized in arachnoid trabecular cells in the leptomeninges of the basal forebrain, and that infusion of PGD<sub>2</sub> into the subarachnoid space of this region increased the extracellular adenosine level and induces an increase in the amount of nonrapid eye movement sleep in wild-type mice and not in DP<sub>1</sub>-deficient mice. Although this study unequivocally demonstrated the involvement of DP<sub>1</sub> in PGD<sub>2</sub>-induced sleep, the baseline sleep-wake patterns were essentially identical between WT and DP(–/–) mice. These results suggest either that the DP<sub>1</sub>-mediated system may not be crucial for physiological sleep regulation or that some other system may effectively compensate

for the loss of the DP system involved in the regulation of sleep. The same group of researchers (Qu et al., 2006) reported that administration of SeCl<sub>4</sub>, as in patients with insomnia, inhibits PGD<sub>2</sub> production and induces insomnia in WT mice but not in DP(–/–) mice. Qu et al. (2006) further showed that administration of a specific DP<sub>1</sub> antagonist, ONO-4127Na (pA<sub>2</sub>, 9.73 for human DP<sub>1</sub>) (Torisu et al. 2003a,b), markedly reduced the sleep period in rats (Qu et al., 2006). On the basis of these findings, they concluded that the PGD<sub>2</sub>-DP<sub>1</sub> signaling is involved in regulation of physiological sleep. The importance of DP in the brain has also been studied from the viewpoint of neuroinflammation. Mohri et al. (2006) detected significantly higher PGD<sub>2</sub> concentration in the brain of the genetic demyelination *twitcher* mouse and found that this was due to induction of hematopoietic type PGD synthase in the microglia of these mice. They further found that DP<sub>1</sub> was induced on astroglia, and loss of DP<sub>1</sub> impaired astrogliosis and demyelination and lessened clinical severity in this line of mice. The same group (Taniguchi et al., 2007) also examined the role of the PGD<sub>2</sub>-DP<sub>1</sub> signaling in hypoxic ischemic encephalopathy in neonatal mice. They subjected 7-day old pups of WT mice or mice deficient in L-PGDS, HPGDS, both L-PGDS and HPGDS, DP<sub>1</sub>, or CRTh2 to left carotid artery ligation and hypoxia exposure and examined the infarct size. They found that the infarct size was significantly enhanced in mice deficient in both L-PGDS and HPGDS or mice deficient in DP<sub>1</sub>. Given the induction of DP<sub>1</sub> in endothelial cells and more severe damage of endothelial cells in DP(–/–) mice, these authors suggested that the PGD<sub>2</sub>-DP<sub>1</sub> signaling exerts a protective action against hypoxic ischemic injury by an action on endothelial cells. Finally, Murata et al. (2008) used DP(–/–) mice and reported that the PGD<sub>2</sub>-DP<sub>1</sub> signaling is involved in suppression of tumor-associated angio-



genesis and hyperpermeability. In this experiment, they implanted Lewis lung cancer cells onto the back of WT and DP(−/−) mice and found enhanced tumor growth in DP(−/−) mice. Furthermore, they found decreased apoptosis and increased angiogenesis and vascular leakage associated with tumors in DP<sub>1</sub>(−/−) mice. Given the expression of DP<sub>1</sub> in endothelial cells and attenuation of angiogenesis and permeability in model systems, they suggested that PGD<sub>2</sub>-DP<sub>1</sub> signaling exerts inhibitory actions in tumor-associated angiogenesis and plasma leakage.

**4. Agonists and Antagonists.** As a general premise, position, type and configuration of oxygen substituents on the cyclopentane ring determines the receptor specificity of natural prostanoids, although no single prostanoid is truly specific. In addition, high agonist potency is generally dependent on a C1-carboxylate, *trans* geometry of the side-chains (8 $\alpha$ ,12 $\beta$  as conventionally drawn), and a 15S-15-hydroxy moiety. The recent work of Ungrin et al. (2001) on the human EP<sub>1</sub> receptor illustrates these relationships well. There are, however, important deviations from this general picture, including the retention of high potency in C1-primary alcohol and 16-hydroxy prostanoids. In addition, quite small alterations to the  $\omega$  terminus often results in large changes in potency. For example, the EP<sub>1</sub> binding affinity of 17S,20-

dimethyl-3,7-dithia PGE<sub>1</sub> is 63-fold higher than its 17R epimer (Maruyama et al., 2002a). Modification of the  $\omega$  chain has, therefore, been an attractive and successful strategy in searching for selective agonists, especially because the synthetic approach is usually ring construction  $\rightarrow$   $\alpha$ -chain attachment  $\rightarrow$   $\omega$ -chain attachment. Non-prostanoid agonists have also been described for DP<sub>2</sub>, EP<sub>2</sub>, EP<sub>3</sub>, and IP receptors.

Replacement of the  $\omega$ -pentyl terminus with a cyclohexyl moiety has been a popular strategy in DP<sub>1</sub> agonist development. For example, the hydantoin ring analog BW 245C (Town et al., 1983; Whittle et al., 1983) is commonly used as a standard agonist; it exhibits minimal DP<sub>2</sub> agonism (Monneret et al., 2001; Yoshimura-Uchiyama et al., 2004). Restricting the conformational mobility of the  $\alpha$ -chain in hydantoin-type analogs conserves DP<sub>1</sub> agonist potency [e.g., BW-587C (Fig. 2); Baraclough et al., 1996], whereas the introduction of methyl, ethyl, and *n*-propyl groups at N<sup>10</sup> progressively reduces efficacy (Leff and Giles, 1992); N<sup>10</sup>-benzyl substitution affords pure DP<sub>1</sub> antagonism (BW A868C). Other potent DP<sub>1</sub> agonists containing a 15-cyclohexyl group include RS-93520 (Fig. 2), a prostacyclin analog with completely inverted stereochemistry in the bicyclic ring (Alvarez et al., 1991; Crider et al., 1999) and SQ-27986 (Fig. 2), a PGH<sub>2</sub> analog with correspondingly in-

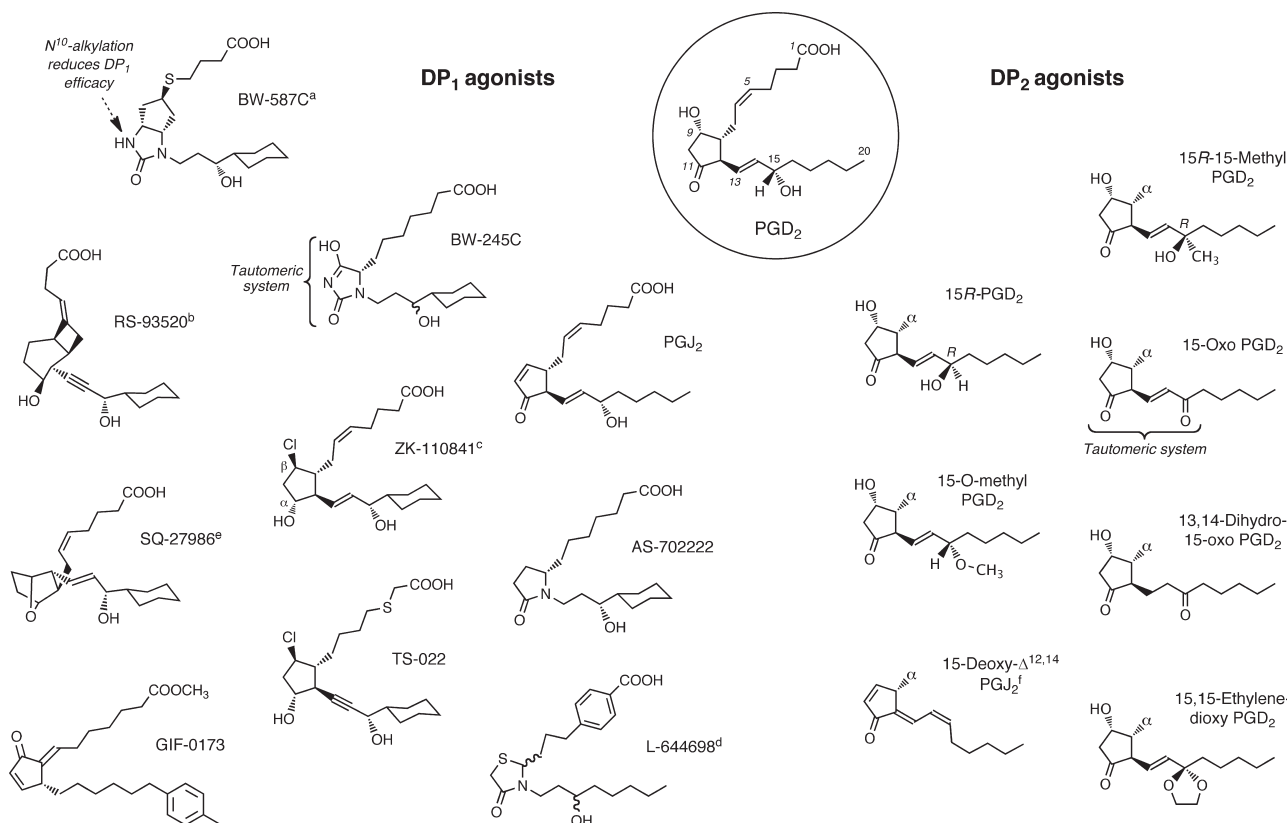


FIG. 2. Structures of agonists for prostanoide DP<sub>1</sub> and DP<sub>2</sub> receptors. PGD<sub>2</sub>, the most active natural agonist, is shown in the circle. Promotion of DP<sub>2</sub>-selectivity mainly involves alterations to C15;  $\alpha$  indicates natural 2-series side-chain. <sup>a</sup>, ring system related to 6 $\beta$ -PGI<sub>1</sub>. <sup>b</sup>, PGI<sub>2</sub> analog with unnatural ring chirality. <sup>c</sup>, EP<sub>1</sub> and EP<sub>2</sub> agonism retained. <sup>d</sup>, some EP<sub>2</sub> agonism remains. <sup>e</sup>, PGH<sub>2</sub> analog with unnatural ring chirality. <sup>f</sup>, 14-*cis* isomer is major component.

verted stereochemistry (see Fig. 2; Seiler et al., 1990). Although the 9 $\beta$ -chloro-11 $\alpha$ -hydroxy ring system in ZK-110841 (Fig. 1) (Thierauch et al., 1988; Ney and Schrör, 1991) and its 3-oxa analog ZK-118182 (Darius et al., 1994) affords high DP<sub>1</sub> agonism, high binding affinity for EP<sub>1</sub> receptors is also found (Wright et al., 1998; Ungrin et al., 2001; Sharif and Davis, 2002). ZK-110841, ZK-118182, and the 13,14-dihydro analog of the latter [9-chloro-15-cyclohexyl-11,15-dihydroxy-3-oxa-16,17,18,19,20-pentano-5-prostenoic acid (AL-6556)] also seem to behave as partial agonists in human functional EP<sub>2</sub> systems (EC<sub>50</sub>, ~200–700 nM) (Sharif et al., 2000, 2004); compare with structure of ONO-AE1-259 in Fig. 4.

Dehydration of the ring system of PGD<sub>2</sub> to give PGJ<sub>2</sub> results in retention of DP<sub>1</sub> agonism (Fukushima et al., 1982; Mahmud et al., 1984; Wright et al., 1998) and approximately a 5-fold loss of DP<sub>2</sub> agonism (Monneret et al., 2002). The related 4-oxo-thiazolidine L-644698 (Fig. 2) shows high DP<sub>1</sub>/DP<sub>2</sub> selectivity but still retains some affinity for EP<sub>2</sub> receptors (K<sub>i</sub>, 270 nM; human recombinant) (Wright et al., 1998); indeed, complete elimination of EP<sub>2</sub> agonism in the search for a selective DP<sub>1</sub> agonist is not an easy task.

The first useful DP<sub>1</sub> antagonist was the xanthone-carboxylic acid AH-6809 (Fig. 5) (Keery and Lumley, 1988). However, it has low affinity (pA<sub>2</sub>, 5.9–6.6), shows considerable EP<sub>1</sub> antagonism and also nonspecificity in the low micromolar range. Subsequently, BW A868C (Fig. 3), which is related to the DP<sub>1</sub> agonist BW 245C, emerged as a selective, competitive blocker of high affinity (human DP<sub>1</sub>: pA<sub>2</sub>, >9) (Giles et al., 1989; Lydford et al., 1996); it is still a first choice antagonist.

The last 10 years has seen the development of other chemical classes of DP<sub>1</sub> antagonist (Fig. 3) (Medina and Liu, 2006), driven by renewed interest in the pathogenic role of PGD<sub>2</sub> in allergic disorders (see Pettipher, 2008).

Emerging from bicyclic prostanoid analogs showing TP antagonism (Narisada et al., 1988), a pinane ring system with a sulfonamido linkage to an aromatic group led to high affinity, as in S-5751 (Fig. 3) (Arimura et al., 2001). A quite different approach has involved the use of indole-3-acetic acid, present in nonsteroidal anti-inflammatory drugs such as indomethacin, as a pharmacophore. In ONO-AE3-237 (Fig. 3), the acetate unit is located on C4; its binding pK<sub>i</sub> for the human rc-DP<sub>1</sub> receptor is 7.7 (Torisu et al., 2004). In the Merck series of DP<sub>1</sub> antagonists, the positions of the carboxylic group and the substituents on the benzene ring of the indole template were optimized to yield MK-0524 (laropiprant) (Sturino et al., 2007). Laropiprant has very high affinity for the human rc-DP<sub>1</sub> receptor (pK<sub>i</sub>, 10.5); it also has ~300-fold lower affinity for the corresponding TP receptor, which requires consideration when using it to characterize prostanoid receptors.

**5. Therapeutics.** DP<sub>1</sub> receptor agonists are not currently used therapeutically. DP<sub>1</sub> agonists are not used to treat glaucoma because of the initial ocular hypertensive spike and an unacceptable incidence of ocular surface hyperemia (Nakajima et al., 1991). Perhaps the most promising of the newly advanced medical hypotheses is their potential utility in treating pruritus and atopic dermatitis (Angeli et al., 2004; Arai et al., 2004, 2007; Sugimoto et al., 2007). DP<sub>1</sub> receptor antagonists, notably BW A868C, have been available for a long time (Giles et al., 1989). The report that laropiprant has no efficacy in patients with allergic rhinitis and asthma (Philip et al., 2009) is discouraging. It is likely that combined DP<sub>1</sub>/DP<sub>2</sub> therapies offer greater promise for treating allergic diseases (Mitsumori, 2004; Pettipher, 2008; Jones et al., 2009). One therapeutic concept that emerged with a positive clinical outcome was the utility of a DP<sub>1</sub> antagonist to limit the cardiovascular side

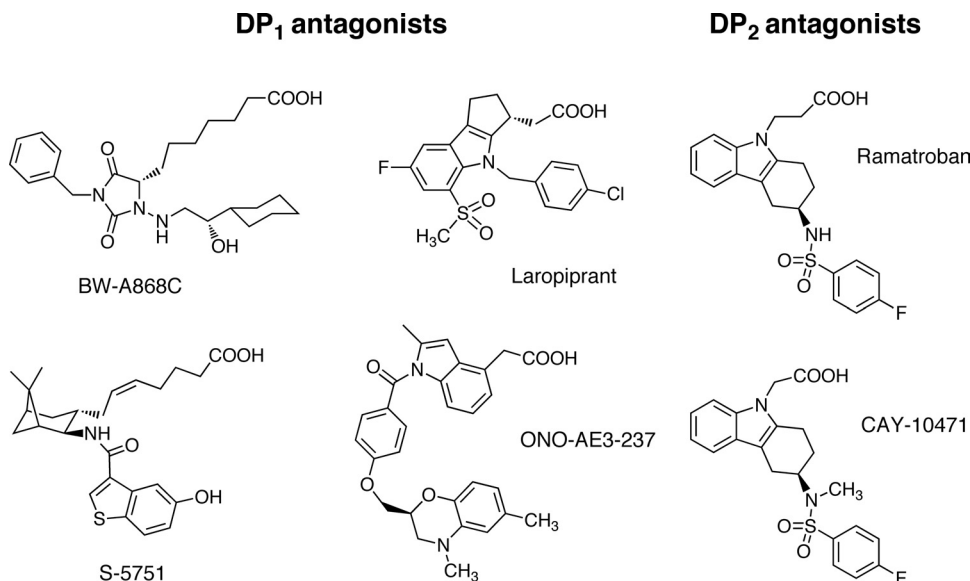


FIG. 3. Structures of representative DP<sub>1</sub> and DP<sub>2</sub> (CRTH2) antagonists.

effects of niacin. The investigational product Cordaptive was not approved as a drug in the United States but seems to have fared better as Tredaptive in Europe (Jones et al., 2009). Thus, the potent vasodilator effects of DP<sub>1</sub> receptor stimulation discovered so many years ago (Coleman et al., 1994b) have successfully transitioned into treatment for nicotinic acid-induced flushing in patients with dyslipidemia (Cheng et al., 2006; Paolini et al., 2009). Finally, DP<sub>1</sub> receptors have been implicated in the development of thyroid eye disease, which affects approximately 40% of patients with Graves hyperthyroidism. DP<sub>1</sub> receptors were found to be an important factor in promoting hyaluronan production, which would be a major contribution to exophthalmos (Guo et al., 2010). Thus, laropiprant would be of potential value in this debilitating ophthalmic condition. The potential therapeutic utilities of DP<sub>1</sub> receptor agonists and antagonists are summarized in Tables 2 and 3, respectively.

## B. Prostaglandin E<sub>2</sub> Receptors

### 1. EP<sub>1</sub> Receptors.

*a. Second messenger signaling.* The EP<sub>1</sub> receptor has long been linked to Ca<sup>2+</sup> mobilization, with a negligible PI response (Funk et al., 1993; Watabe et al., 1993; Katoh et al., 1995). Ca<sup>2+</sup> mobilization patterns appear variable according to cell type studied. It is still not entirely clear which G protein(s) may be involved. The involvement of the phospholipase C (PLC)/PKC pathway in EP<sub>1</sub>-mediated trophoblast and osteoblast stimulation implies that EP<sub>1</sub> receptors may couple to G<sub>q</sub> (Nicola et al., 2005; Tang et al., 2005). Compared with other prostanoid receptor studies, the signaling properties of EP<sub>1</sub> receptors have received little attention. EP<sub>1</sub> receptor mediated dephosphorylation of phosphatase and tensin homolog deleted on chromosome 10 and protein kinase B (Akt) (Zhou et al., 2008), and NO/cGMP pathway effects have been reported (Bachteeva et al., 2007).

The coexpression of more than one prostanoid receptor or isoform adds to the diversity of effects already inherent to prostanoid receptor stimulation. These include, but are not limited to, transactivation and cross-desensitization. EP<sub>1</sub> receptor stimulation results in desensitization of TP receptors by PKC-mediated phosphorylation of C-terminal residues (Kelley-Hickie and Kinsella, 2004). An EP<sub>1</sub> receptor-mediated transactivation of epidermal growth factor receptors, with resultant Akt activation, has also been reported (Han and Wu,

2005). The potential for transactivation obviates the need to distinguish such an event from signaling cascades directly emanating from activation of EP<sub>1</sub> receptors per se.

*b. Distribution and biological functions.* Contractile EP<sub>1</sub> receptors do not have a widespread distribution in higher species and are more common in guinea pigs and murine species (Coleman et al., 1994b). In human tissues and cells, functional EP<sub>1</sub> receptors have been demonstrated in the myometrium (Senior et al., 1991), pulmonary veins (Norel et al., 2004), mast cells (Wang and Lau, 2006) colonic longitudinal muscle (Smid and Svensson, 2009), and keratinocytes (Konger et al., 2009).

Northern blotting revealed EP<sub>1</sub> transcription in the lung, stomach, and kidney of mice (Watabe et al., 1993). Functional studies in knockout mice and the employment of pharmacological tools have provided evidence for physiological participation for EP<sub>1</sub> receptors in each of these organs. EP<sub>1</sub> receptors produce airway constriction in mice, but this seems to be neuronally mediated rather than a direct smooth muscle effect (Tilley et al., 2003). EP<sub>1</sub> receptors are also claimed to mediate PGE<sub>2</sub>-induced surfactant secretion from rat alveolar type II cells (Morsy et al., 2001). In the stomach, EP<sub>1</sub> receptors seem to have detrimental and beneficial effects. Histamine-induced gastric injury, in the form of increased vascular permeability, is worsened by EP<sub>1</sub> receptor activation (Hase et al., 2003). On the other hand, the EP<sub>1</sub> receptor affords cytoprotection to the gastric mucosa against hemorrhagic lesions produced by indomethacin and HCl/C<sub>2</sub>H<sub>5</sub>OH injury (Kunikata et al., 2001; Takeuchi et al., 2001a,c). EP<sub>1</sub> receptors are essential for HCO<sub>3</sub><sup>-</sup> secretion in response to mucosal acidification in the stomach (Takeuchi et al., 2006). A dual role for EP<sub>1</sub> receptors in esophagitis is clearer, whereby PGE<sub>2</sub> has a protective effect at low doses and a deleterious effect at high doses (Yamato et al., 2005). In the kidney, PGE<sub>2</sub> activates EP<sub>1</sub> receptors to inhibit Na<sup>+</sup> absorption by the renal collecting duct (Guan et al., 1998). EP<sub>1</sub> receptors up-regulate transcription of the Na,K-ATPase β subunit in Madin-Darby canine kidney cells (Matlhagela and Taub, 2006) and attenuate up-regulation of epithelial sodium channel mRNA in inner medullary collecting duct cells by aldosterone (González et al., 2009). EP<sub>1</sub> antagonists have been claimed as useful in the prevention of diabetic nephropathy (Makino et al., 2002) and hypertension-induced renal injury (Suganami et al., 2003). Severe renal impairment has been reported in

TABLE 3  
Potential therapeutic application DP<sub>1</sub> antagonists

DP <sub>1</sub> Antagonist	Route	Dose	Species	Experimental Model	Indication	Reference
Laropiprant	Oral	37.5 → 300 mg (human dose)	Mice, humans	NA-induced cutaneous vasodilation in mice	Adjunctive therapy for dyslipidemia	Cheng et al., 2006; Paolini et al., 2009
S-5751	Oral	30 mg/kg	Sheep	<i>Ascaris suum</i> -induced asthma	asthma	Shichijo et al., 2009

NA, nicotinic acid.



glomerulonephritic EP<sub>1</sub> knockout mice, underscoring an important role for EP<sub>1</sub> in pathological renal conditions (Rahal et al., 2006). Although a few arguably beneficial effects may be ascribed to EP<sub>1</sub> receptor activation, most effects are pathophysiological.

Prevention of EP<sub>1</sub> receptor activation has been purported as an attractive proposition for many diseases but colon cancer, systemic hypertension and, above all, inflammation and associated pain have received the most attention. Cyclooxygenases and their products have long been considered to play a role in colon carcinogenesis. Animal models have provided a direct link between EP<sub>1</sub> receptors and colon cancer. Aberrant crypt foci, induced by azoxymethane, were reduced by the EP<sub>1</sub> antagonists 6-((2S,3S)-3-(4-chloro-2-methylphenylsulfonylaminomethyl)-bicyclo(2.2.2)octan-2-yl)-5Z-hexenoic acid (ONO-8711) (Watanabe et al., 2000; Kawamori et al., 2001a) and ONO-8713 (Fig. 5) (Watanabe et al., 2000). A reduction in intestinal polyps was observed in the adenomatous polyposis coli gene knockout mouse model of tumorigenesis (Watanabe et al., 1999; Kitamura et al., 2003b).

A role for EP<sub>1</sub> receptor in cardiovascular homeostasis is indicated by knockout mouse studies (Audoly et al., 1999; Stock et al., 2001). Physiological regulation by presynaptic EP<sub>1</sub> receptors has been recently implicated in nitrergic neurovascular transmission (Jadhav et al., 2009). In models of hypertension, blockade of EP<sub>1</sub> receptors or gene deletion seems to confer antihypertensive effects in diabetic mice (Rutkai et al., 2009) and spontaneously hypertensive rats (Guan et al., 2007). EP<sub>1</sub> antagonist treatment also dramatically improved arteriolar lesions (Suganami et al., 2003).

The involvement of EP<sub>1</sub> receptors in inflammation, inflammatory pain and hyperalgesia, and neuropathic pain has been a major research focus. The anti-inflammatory activity of EP<sub>1</sub> antagonists has been reviewed extensively (Jones et al., 2009). EP<sub>1</sub> receptors contribute to neuronal sensitization at peripheral sites (Omote et al., 2001) and at several levels in the CNS. EP<sub>1</sub> receptors are localized in dorsal root ganglion neurons (Nakayama et al., 2004). Intrathecal PGE<sub>2</sub> causes hyperalgesia in response to innocuous mechanical stimuli, an effect found to be EP<sub>1</sub> receptor-mediated (Minami et al., 1994; Nakayama et al., 2004). Intrathecal administration of the EP<sub>1</sub> antagonist ONO-8711 was shown to inhibit only the late phase of the mechanical hyperalgesic response associated with carrageenan-induced rat paw edema (Nakayama et al., 2002) and postoperative pain (Omote et al., 2002). These results correlate with a study on intra-articular Kaolin injection, in which spinal application of an EP<sub>1</sub> agonist caused hyperexcitability 7 to 11 h after administration of the inflammatory stimulus (Bär et al., 2004). This time-dependent late phase response was not observed for EP<sub>2</sub> and EP<sub>4</sub> agonists (Bär et al., 2004). Set against these findings with respect to EP<sub>1</sub> receptors in the spinal dorsal horn mediating

hyperalgesia, microinjection of PGE<sub>2</sub> into the ventromedial hypothalamus produced an EP<sub>1</sub> receptor-mediated antinociceptive effect (Hosoi et al., 1999). Electrophysiological evidence has been provided for EP<sub>1</sub>-mediated hypoalgesia in response to noxious pinching of facial skin after lateral cerebroventricular administration of a receptor selective agonist and antagonists (Oka et al., 1997). These findings suggest that spinal processing of peripheral input may be subsequently relayed by EP<sub>1</sub> receptors to higher centers, where the same (EP<sub>1</sub>) receptors attenuate transmission. A strong, centrally mediated override by EP<sub>1</sub> receptors does not, however, seem to be the case because systemically administered EP<sub>1</sub> antagonists are widely reported to be analgesic and antiallodynic (Hall et al., 2007a; Jones et al., 2009).

The EP<sub>1</sub> receptor plays additional significant roles in the CNS. Of considerable interest is the role of EP<sub>1</sub> receptors in controlling stress-induced impulse behavior. Thus, in mice lacking EP<sub>1</sub> receptors, stress induces impulsive aggression, an exaggerated acoustic startle response, impaired cliff avoidance, and social dysfunction (Matsuoka et al., 2005). This behavioral phenotype was reproduced in wild-type mice by an EP<sub>1</sub> antagonist and corrected by a dopaminergic antagonist (Matsuoka et al., 2005), establishing a link between EP<sub>1</sub>, DP<sub>1</sub>, and D<sub>2</sub> receptor function (Kitaoka et al., 2007). EP<sub>1</sub> receptor stimulation has also been shown to cause hyperthermia (Oka and Hori, 1994; Oka et al., 2003b).

Beyond studies on tissues and living animals, EP<sub>1</sub> expression and functional analyses in individual cell types has produced interesting results. In human primary keratinocytes, EP<sub>1</sub> receptors evoked intracellular Ca<sup>2+</sup> mobilization and were shown to be expressed in the epidermis (Konger et al., 2005a). The growth of malignant keratinocytes (Thompson et al., 2001) and regulation of keratinocyte differentiation (Konger et al., 2009) seem to be EP<sub>1</sub> receptor-dependent. EP<sub>1</sub> receptor stimulation caused differentiation of uncommitted T cells to a Th1 phenotype, which are involved in cell-mediated immune reactions, such as dinitrofluorobenzene (DNFB) contact sensitivity (Nagamachi et al., 2007). Blockade of EP<sub>1</sub> receptors has been shown to inhibit receptor activator of nuclear factor- $\kappa$ B ligand (RANKL)-induced osteoclastogenesis (Tsujisawa et al., 2005). Hypoxia induces increased EP<sub>1</sub> receptor expression in osteoclasts (Lee et al., 2007a) by a signal transduction pathway involving HIF-1 $\alpha$  (Genetos et al., 2009). A systematic study on EP<sub>1</sub> receptor-mediated up-regulation of HIF-1 $\alpha$  implicated G<sub>i</sub> coupling with activation of a PI3K/Akt/mammalian target of rapamycin signaling pathway, and HIF-1 $\alpha$  induction was associated with phosphorylation of the ribosomal protein 56 (Ji et al., 2010). Most noteworthy is perhaps the involvement of EP<sub>1</sub> receptors in the proliferation of growth plate chondrocytes, the growth plate functioning to ossify long bones (Brochhausen et al., 2006).

*c. Gene deletion studies.* EP<sub>1</sub>(-/-) mice appeared quite normal in all respects, including morphological characteristics (Ushikubi et al., 1998). Prostanoid EP<sub>1</sub> receptors have long been heavily implicated in all aspects of nociception, from hyperalgesia to neuropathic pain. The advent of knockout mice has enabled follow-up of these observations, which were made with agonists and antagonists and the inherent complications of off-target pharmacology, drug-induced toxicity, and substandard experimental design. A role for EP<sub>1</sub> receptors in pain was supported by studies on the stretching/writhing response to noxious chemical stimuli (Stock et al., 2001). Nevertheless, other EP<sub>1</sub> gene deletion studies failed to confirm the substantive role of EP<sub>1</sub> receptors in pain and inflammation suggested by agonist/antagonists studies in living animal models (Jones et al., 2009). According to gene deletion studies, IP receptors would be regarded as more important mediators of pain and inflammation (Murata et al., 1997; Ueno et al., 2000; Honda et al., 2006). Studies on EP<sub>1</sub>(-/-) mice actually indicated an increase in thermal nociception, EP<sub>1</sub> receptors being suggested to exert centrally mediated control of thermal pain sensitivity (Popp et al., 2009).

Perhaps the most intriguing aspect of CNS function that emerged from gene deletion studies is the role of EP<sub>1</sub> receptors in regulating stress responses (Furuyashiki and Narumiya, 2009). Stress, defined as a condition where body homeostasis is perturbed (Furuyashiki and Narumiya, 2009), elicits an adaptive response. This may take the form of febrile, neuroendocrine, and behavioral responses, and EP<sub>1</sub> receptors participate in all of these. Although EP<sub>3</sub> seems to be the dominant receptor in mediating fever (Ushikubi et al., 1998; Oka et al., 2003a; Furuyashiki and Narumiya, 2009), EP<sub>1</sub> and EP<sub>3</sub> receptors are equally important in PGE<sub>2</sub>-evoked adrenocorticotrophic hormone and glucocorticoid release via the activation of corticotrophin-releasing hormone-containing neurons in the paraventricular nucleus of the hypothalamus (Matsuoka et al., 2003). The behavioral effects observed in EP<sub>1</sub>(-/-) mice are notable. Social withdrawal, impulse aggression, impaired cliff avoidance, and an enhanced acoustic startle response were apparent (Matsuoka et al., 2005). These behaviors were attributed to a lack of inhibition of impulsive activity, implying that EP<sub>1</sub> receptors suppress impulsive activity under stress (Matsuoka et al., 2005). The EP<sub>1</sub> receptor is also neurotoxic and has been postulated as the essential downstream effector of COX-2-induced neurocytotoxicity (Kawano et al., 2006). In EP<sub>1</sub> receptor-deficient mice, it was found that COX-2-derived PGE<sub>2</sub> does not mediate NMDA cytotoxicity (Kawano et al., 2006). EP<sub>1</sub> receptor gene deletion also ameliorated brain injury produced by oxygen/glucose deprivation (Kawano et al., 2006) and ischemic damage produced by middle cerebral artery occlusion (Ahmad et al., 2006a; Kawano et al., 2006). In contemplating EP<sub>1</sub> receptor participation in the middle cerebral artery occlusion

model, it is important to take into consideration that cerebral blood flow is significantly increased in EP<sub>1</sub>(-/-) mice (Saleem et al., 2007a). EP<sub>1</sub> receptors have also been implicated in innate immune responses in the CNS. PGE<sub>2</sub>, signaling via either EP<sub>1</sub> or EP<sub>2</sub> receptors, is essential for Toll-like 4 receptor-mediated depletion of intermediate progenitor cells from the hippocampal subgranular zone (Keene et al., 2009).

A role for EP<sub>1</sub> receptors in carcinogenesis has been confirmed by gene deletion studies. In the azoxymethane colon cancer model, formation of aberrant crypt foci was reduced by approximately 40% in EP<sub>1</sub>(-/-) mice (Watanabe et al., 1999), and the number of tumors formed was essentially halved (Kawamori et al., 2005). In the methylcholanthrene-induced sarcoma model (MGC 101 mice), long-term growth was attenuated in EP<sub>1</sub>-deficient mice (Axelsson et al., 2005; Wang et al., 2005). The effects of cyclooxygenase inhibition on tumor progression were primarily on cell proliferation and apoptosis; angiogenesis was not an obvious primary determinant of onset and progression of tumor development (Axelsson et al., 2005).

Resting systolic blood pressure was reduced by approximately 10 mm Hg in EP<sub>1</sub> receptor-deficient mice (Stock et al., 2001). These findings transitioned into cardiovascular hypertension, where EP<sub>1</sub> receptor disruption spared the protein kinase N locus and reduced the acute vasopressor response and chronic hypertension produced by angiotensin II (Guan et al., 2007). The hypotension in EP<sub>1</sub>-deficient mice, notably male mice, was reported to elicit physiological compensation that manifested as increased pulse rate, increased renin mRNA levels in the kidney, and increased plasma renin activity (Stock et al., 2001). In experimental glomerulonephritis, there was reduced urine osmolality, and overall renal damage was more acute in EP<sub>1</sub>(-/-) mice (Rahal et al., 2006). PGE<sub>2</sub>, via EP<sub>1</sub>, modulates urine concentration not modulated in the renal collecting duct but within the hypothalamus to promote arginine vasopressin biosynthesis in response to water deprivation (Kennedy et al., 2007). In a model of bladder outlet obstruction, detrusor hyperactivity was negligible in EP<sub>1</sub> receptor knockout mice (Schröder et al., 2004).

Two further findings from EP<sub>1</sub>(-/-) mice are noteworthy: 1) EP<sub>1</sub> receptors seem to be critically involved in shifting the Th1/Th2 balance to Th1 dominance (Nagamachi et al., 2007); this was demonstrated in a therapeutic sense by reduced DNFB-induced contact sensitivity in EP<sub>1</sub> receptor-deficient mice (Nagamachi et al., 2007). 2) Adaptive gastric cytoprotection is apparently mediated by EP<sub>1</sub> receptors (Takeuchi et al., 2001a,c).

*d. Agonists and antagonists.* 17-Phenyl PGE<sub>2</sub> (Fig. 4) has modest EP<sub>1</sub>/EP<sub>3</sub> selectivity (Lawrence et al., 1992) and is a useful agonist in Schild antagonism protocols because of its high potency. ONO-DI-004 (Fig. 4) is a more selective EP<sub>1</sub> agonist (Okada et al., 2000; Suzawa et al., 2000), resulting from development of 6-oxo PGE<sub>1</sub> via its

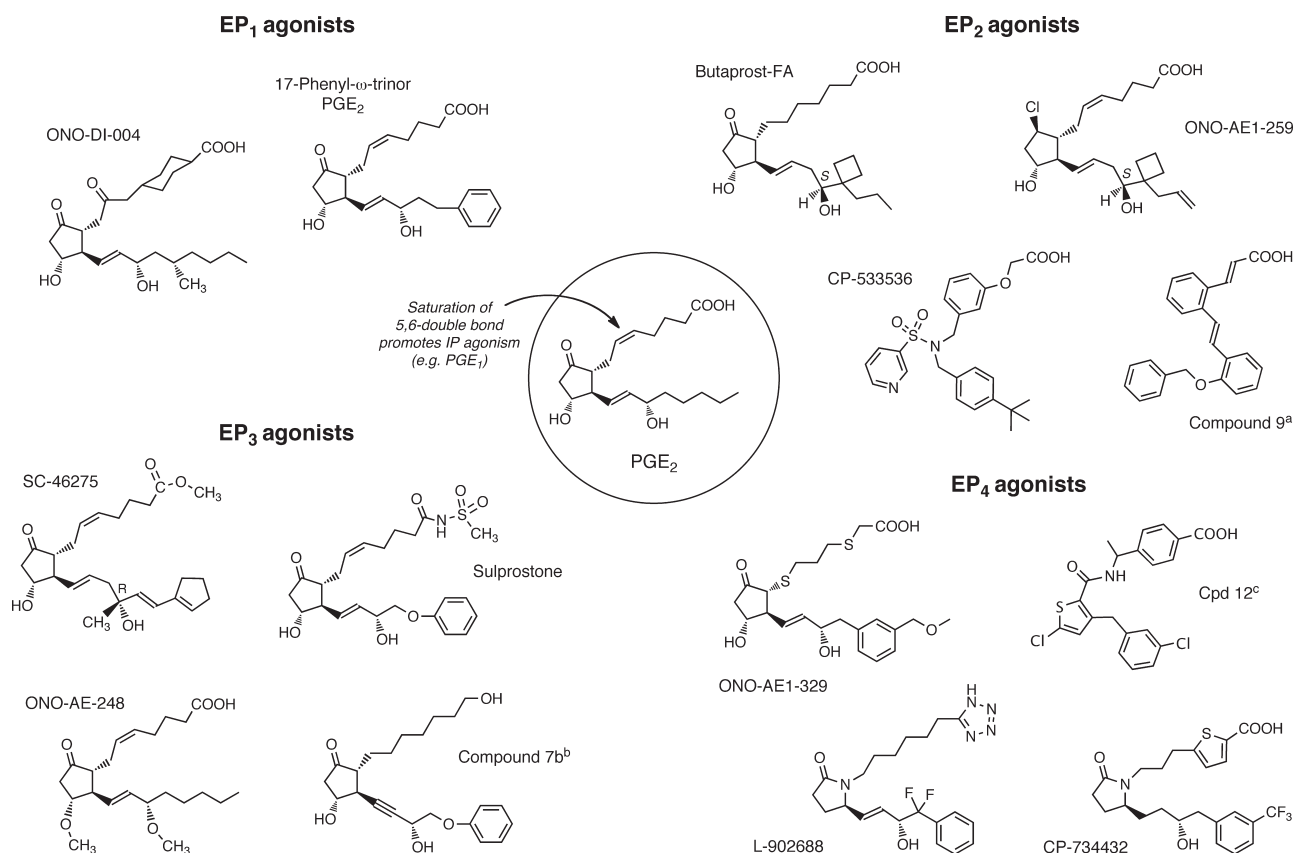


FIG. 4. Structures of agonists for prostanoid EP receptor subtypes. PGE<sub>2</sub>, the most active natural agonist, is shown in the circle. 17-Phenyl PGE<sub>2</sub> and sulprostone have modest EP<sub>1</sub>/EP<sub>3</sub> and EP<sub>3</sub>/EP<sub>1</sub> selectivities, respectively. CP-533536 and compound 9 are EP<sub>2</sub> agonists with nonprostanooid structures. <sup>a</sup>, Belley et al. (2005). <sup>b</sup>, Shimazaki et al. (2000). <sup>c</sup>, Blovin et al. (2010).

17*S*,20-dimethyl (methyl ester) analog [OU-1308 (ornoprostil)] (Kobayashi et al., 1991). 6*α*-Carba analogs of prostacyclin, such as carbacyclin and iloprost, are unexpectedly potent EP<sub>1</sub> agonists (Dong and Jones, 1982; Dong et al., 1986; Lawrence et al., 1992), iloprost showing partial agonism in some systems (Dong and Jones, 1982; Dong et al., 1986; Boie et al., 1997). Functional studies with rat and human recombinant EP<sub>1</sub> receptors in either reporter gene (Durocher et al., 2000) or aequorin-based Ca<sup>2+</sup> flux assays (Boie et al., 1997; Ungrin et al., 2001) have confirmed and expanded these structure-activity relationship data.

The first EP<sub>1</sub> receptor antagonist was 8-chloro-dibenzo[*Z*][*b*,*f*][1,4]oxazepine-10(11*H*)-carboxylic acid, 2-acetylhydrazide (SC-19220), which is a dibenzoxazepine hydrazide (Sanner, 1969). It has low affinity (pA<sub>2</sub> = 5.5), but proved useful in the early characterization of EP receptor pharmacology. Modification of SC-19220, notably removal of the acetyl group, led to the thioether SC-51222 (Fig. 5), which was much more potent than the corresponding sulfone (Hallinan et al., 1994). Within the Searle series, SC-51322 has become the agent of choice for EP<sub>1</sub> receptor pharmacology studies (Fig. 5).

An alternative early EP<sub>1</sub> antagonist was AH-6809 (Fig. 5). Over a 0.1 to 10 μM concentration range, it is

selective for EP<sub>1</sub> receptors (pA<sub>2</sub>, 7.4) (Coleman et al., 1987; Eglen and Whiting, 1988; Lawrence et al., 1992) and does not block EP<sub>3</sub> receptors (Lawrence et al., 1992; Racké et al., 1992; Qian et al., 1994). AH-6809 was later reported to antagonize the human EP<sub>2</sub> receptor (Woodward et al., 1995b) and is now probably more useful for this purpose, given the diverse array of potent and selective EP<sub>1</sub> antagonists currently available.

EP<sub>1</sub> antagonists designed by Ono Pharmaceuticals show an interesting progression from the TP antagonist 11*α*-carba-12-(2'*S*-hydroxy-3'-phenylpropylamino)-9*α*,11*α*-isopropylideno-*ω*-octanor-prost-5*Z*-enoic acid (ONO-11120) (Katsura et al., 1983) to a related pinane analog (ONO-NT-012; Minami et al., 1995) showing EP<sub>1</sub>, FP, and TP antagonism (and EP<sub>3</sub> agonism) to a bicyclo[2.2.2]octane analog (ONO-8711) showing EP<sub>1</sub>/EP<sub>3</sub> antagonism and ultimately to the nonprostanoids ONO-8713 (Fig. 5), with high selectivity for the EP<sub>1</sub> receptor. ONO-8711 and ONO-8713 possess *K*<sub>d</sub> values for mouse recombinant EP<sub>1</sub> receptors of 1.7 and 0.3 nM, respectively (Watanabe et al., 1999, 2000; Naganawa et al., 2006). Small modifications may dramatically increase EP<sub>3</sub> antagonist affinity. This series has also been widely employed to elucidate the therapeutic utility of EP<sub>1</sub> antagonists.



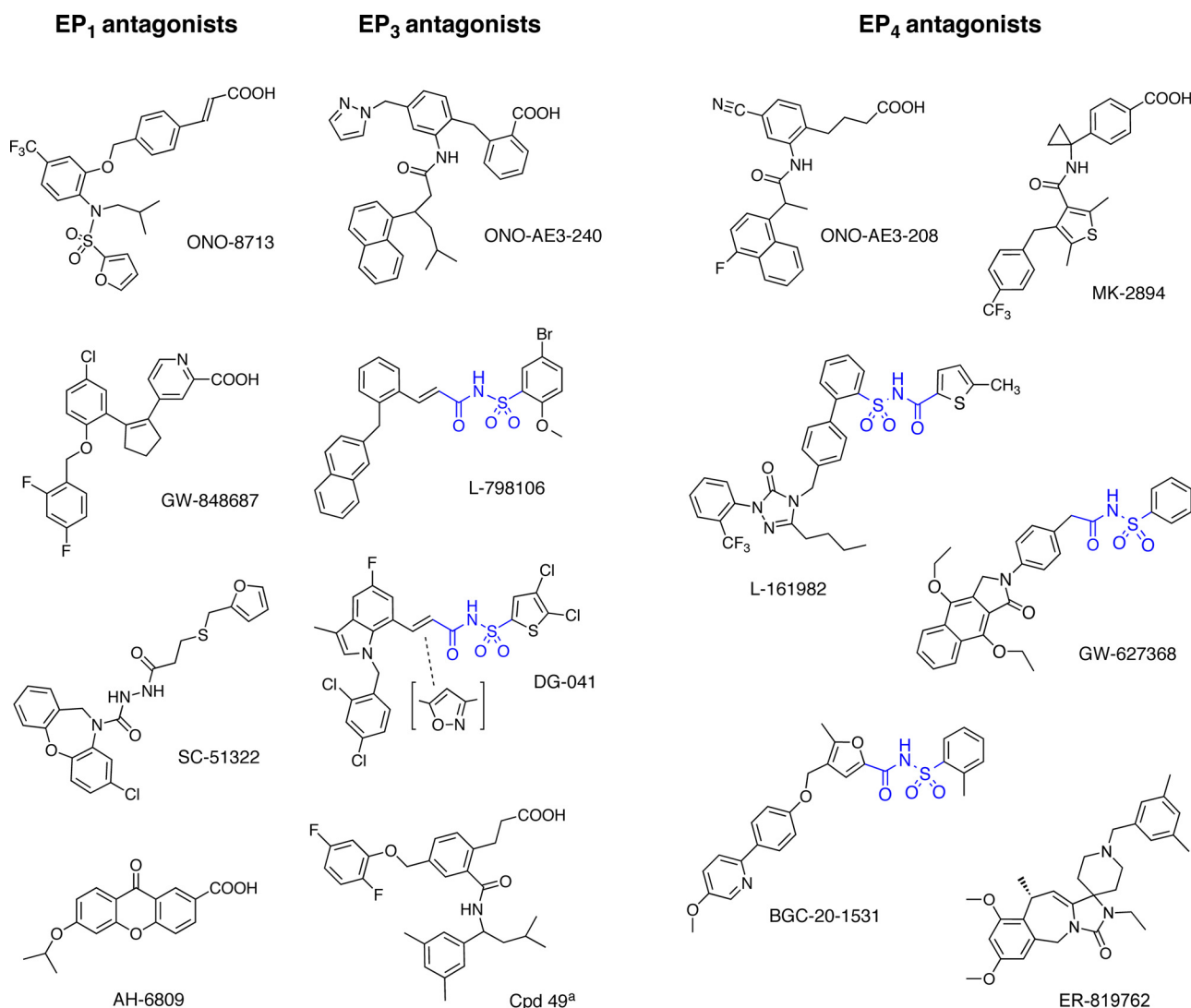


FIG. 5. Structures of representative EP<sub>1</sub>, EP<sub>3</sub>, and EP<sub>4</sub> receptor antagonists. Heterocycle substitution in DG-041 maintains high EP<sub>3</sub> affinity (Hategan et al., 2009).<sup>a</sup> Asada et al. (2010). ONO-AE3-240, T. Maruyama, personal communication.

Pharmacophores possessing potent EP<sub>1</sub> antagonism with good CNS penetration have emerged over the last 10 years. The series reported by Merck (Ruel et al., 1999) contains a tricyclic system similar to the Searle series. A large number of 1,2-diaryl-thiophene/cyclopentene analogs have been prepared as highly potent EP<sub>1</sub> antagonists, including 1-(5-{3-[2-(benzyloxy)-5-chlorophenyl]-2-thienyl}pyridin-3-yl)-2,2,2-trifluoroethane-1,1-diol (MF-266-1) (Ducharme et al., 2005; Clark et al., 2008) and GW-848687 (Giblin et al., 2007), shown in Fig. 5. Nonacidic analogs of GW-848687 (e.g., pyridylmethyl-amides) have been reported (Hall et al., 2007b,c).

*e. Therapeutics.* Very few potential medical uses for EP<sub>1</sub> agonists have been presented to date. Moreover, the prospect of major unwanted side effects would be anticipated. Probably the most promising clinical application is for EP<sub>1</sub> agonist control of impulsive behavior in psychiatric patients (Matsuoka et al., 2005). EP<sub>1</sub> antagonists are quite a different matter and several therapeutic applications have been put forward. Indeed, EP<sub>1</sub>

receptors have been described as the downstream effectors of COX-2-induced neurotoxicity (Kawano et al., 2006). These proposed therapeutic applications are listed in Table 4, with only a brief reference to antinociceptive and anti-inflammatory activities, because these have been extensively reviewed (Jones et al., 2009). Despite considerable effort devoted to the design, synthesis, and testing of EP<sub>1</sub> antagonists, no convincing evidence of clinical efficacy seems to have emerged. Clinical success seems limited to acid-induced visceral pain hypersensitivity (Sarkar et al., 2003), which arguably portends little for other indications.

## 2. EP<sub>2</sub> Receptors.

*a. Second messenger signaling.* EP<sub>2</sub> receptors are G<sub>s</sub>-coupled and mediate increases in cAMP (Regan et al., 1994b). A positive feedback loop whereby cAMP signaling enhances EP<sub>2</sub> receptor expression has been suggested (Sagana et al., 2009). Changes in cAMP levels produce pleiotropic effects by activating cAMP-binding proteins. These include PKAs, Epacs, and cAMP re-

TABLE 4  
*Potential therapeutic application of EP<sub>1</sub> antagonists*

EP <sub>1</sub> Antagonists	Route	Dose	Species	Experimental Model	Indication	Reference
ONO-8711	Intravenous	1 and 3 mg/kg	Rat	Acetic acid-induced inflammation	Bladder inflammation	Ikeda et al., 2006
ONO-8711	Dietary	400 or 800 ppm	Rat	PhIP-induced breast cancer	Breast cancer	Kawamori et al., 2001a,b
ONO-8711	Dietary	250, 500, 1000 ppm	Mouse	Azoxymethane-induced aberrant crypt foci, Min mice	Colon cancer	Watanabe et al., 1999
ONO-8713	Oral	20 mg/kg	Mouse	DNFB contact sensitivity	Contact dermatitis	Nagamachi et al., 2007
ONO-8713	Topical	50 $\mu$ g	Mouse	UVB-induced inflammation and tumor formation	Sunburn, skin cancer	Tober et al., 2006
ONO-8713	Dietary	250, 500 1000 ppm	Mouse	Azoxymethane-induced aberrant crypt foci	Colon cancer	Watanabe et al., 2000
ONO-8713	Intraventricular	0–1–10 nM	Mouse	NMDA excitotoxicity	Stroke	Ahmad et al., 2006a
ONO-8713	Intraperitoneal	10 mg/kg	Mouse	Cerebral artery occlusion	Stroke	Ahmad et al., 2006a
ONO-8713	Intrathecal	1 or 10 nmol	Mouse	Neurochemical allodynia	Pain	Tsukamoto et al., 2010
PF-2907617-02	Intravenous	1 mg/kg	Rat	Balloon catheter-induced bladder outlet obstruction	Overactive bladder syndrome	Lee et al., 2007a,b
SC-51089	Intraperitoneal	25 mg/kg	Mice	KMG4 tumor xenografts on SCID mice	Glioma	Matsuo et al., 2004
SC-51089	Intraperitoneal	10 mg/kg	Rat	Lithium-pilocarpine status epilepticus	Epilepsy	Pekcec et al., 2009
SC-51322	Oral	10 mg/kg	Rat	Spontaneously hypertensive rat	Hypertension	Guan et al., 2007

PhIP, 2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine; SC-51089, dibenz(*b,f*)(1,4)oxazepine-10(11*H*)-carboxylic acid, 8-chloro-, 2-(1-oxo-3-(4-pyridinyl)propyl)hydrazide, monohydrochloride.

sponse element-binding protein regulation of gene transcription. The nature of the cellular responses to cAMP is also dependent on compartmentalization; in fact, the EP<sub>2</sub> receptor seems to be excluded from caveolin-rich membrane fractions (Ostrom et al., 2001). The potential repertoire of EP<sub>2</sub>-mediated responses via cAMP-Epac-PKA-cAMP response element-binding protein has yet to be elucidated for many cells. The potential diversity may include ion channel function, [Ca<sup>2+</sup>] signaling, ion transport, exocytosis, cell adhesion, and gap junction formation (Holz et al., 2006). In addition, cAMP can influence several transcription factors by both PKA dependent and independent mechanisms (Sands and Palmer, 2008). Beyond G<sub>s</sub>, it is probably better to describe cell signaling on a “cell-to-cell” basis. Finally, EP<sub>2</sub> receptors inhibit the formyl-Met-Leu-Phe-induced phospholipase D pathway activation of neutrophils (Burelout et al., 2004) and cause Th1 cell differentiation (Yao et al., 2009), which are dependent on PI3K rather than cAMP.

*b. Distribution and biological functions.* EP<sub>2</sub> receptors seem widely distributed, according to functional studies on isolated tissues, where they almost invariably produce relaxation (Coleman et al., 1994b). The earliest studies on mRNA expression suggested relatively low abundance and an uncertain distribution pattern (Regan et al., 1994b; Narumiya et al., 1999; Smock et al., 1999). In the past decade, a number of studies have indicated widespread distribution and important functions.

In the absence of potent and selective antagonists, gene deletion studies are of essential value in understanding the physiological and pathological roles of the EP<sub>2</sub> receptor in living animals. In isolated tissues and cell culture, bioavailability is not an issue and therefore the EP<sub>2</sub>/EP<sub>1</sub> antagonist AH-6809 (Fig. 5) may be used despite its low potency (Woodward et al., 1995b; Jones et al., 2009). In conjunction with selective agonists, the role

of EP<sub>2</sub> receptors is now better understood in many biological systems. EP<sub>2</sub> receptors exert many inhibitory functions. In the general context of PG release and activity, the EP<sub>2</sub> receptor could be considered to provide a major regulatory component in many instances.

PGE<sub>2</sub> has long been known to be a bronchodilator with potential for treating asthma (Wasserman, 1981; Gardiner, 1986). The prospects improved with the discovery of selective EP<sub>2</sub> agonists (Gardiner, 1986; Nials et al., 1993). PGE<sub>2</sub>-induced bronchodilation has been confirmed as EP<sub>2</sub> receptor-mediated relaxation in isolated human bronchial preparations (Norel et al., 1999) and mouse airways (Sheller et al., 2000; Tilley et al., 2003; Hartney et al., 2006). EP<sub>4</sub> receptors do not seem to mediate PGE<sub>2</sub>-induced relaxation of human bronchi (Norel et al., 1999). EP<sub>2</sub> receptors also mediate substance P- and ATP-induced airway relaxation (Fortner et al., 2001). EP<sub>2</sub> receptor activation on human airway smooth muscle cells may indirectly produce anti-inflammatory effects. IL-1 $\beta$  releases granulocyte macrophage–colony-stimulating factor from human airway smooth muscle cells, which is inhibited by EP<sub>2</sub> receptor stimulation (Clarke et al., 2004). Thus, the survival of infiltrating leukocytes by granulocyte macrophage–colony-stimulating factor would be abrogated by the action of PGE<sub>2</sub> at EP<sub>2</sub> receptors.

The tocolytic effect of EP<sub>2</sub> agonists has also received attention. Suppression of spontaneous uterine activity has been reported with selective EP<sub>2</sub> receptor agonists: 19(*R*)-OH PGE<sub>2</sub> in rabbit (Spilman et al., 1977; Woodward et al., 1993a) and butaprost in human preparations (Senior et al., 1991; Duckworth et al., 2002). 19(*R*)-OH PGE<sub>2</sub> increased uterine motility in a second-trimester pregnant monkey (Spilman et al., 1977), but this effect did not transition into the human pregnant myometrium, where EP<sub>2</sub> receptor stimulation produced tocolysis (Duckworth et al., 2002). Temporal and re-

gional changes in EP<sub>2</sub> receptor expression have also been implicated in pregnancy maintenance and labor-associated events. EP<sub>2</sub> receptors decline toward term gestation (Brodth-Eppley and Myatt, 1999; Leonhardt et al., 2003), although they remain unaltered (Brodth-Eppley and Myatt, 1999; Astle et al., 2005; Sooranna et al., 2005) or increase during parturition (Grigsby et al., 2006). EP<sub>2</sub> mRNA and nuclear EP receptors were most abundant in lower compared with upper segment tissues (Astle et al., 2005; Grigsby et al., 2006). This corresponds with a caudal decline in contractile responsiveness to PGE<sub>2</sub>, suggesting that EP<sub>2</sub> receptors (Wikland et al., 1984) facilitate uterine distension for delivery of the fetus during labor. Rabbit oviductal motility was also suppressed by 19(R)-OH PGE<sub>2</sub> (Spilman et al., 1977). Should this effect occur in humans, with a resultant slowdown in Fallopian tube movement, it could disrupt the timing for implantation. It could even result in an ectopic pregnancy, an unwanted event made more likely by EP<sub>2</sub> receptor signaling as a contributory factor for fertilization and implantation (Lim and Dey, 1997; Hizaki et al., 1999; Tamba et al., 2008).

EP<sub>2</sub> receptors have long been known to relax vascular smooth muscle and produce a vasodepressor response (Armstrong et al., 1976; Audoly et al., 1999). The renin-angiotensin system also regulates systemic blood pressure, and EP<sub>2</sub> receptors are involved. In addition to controlling electrolyte and water homeostasis, the kidney also regulates blood pressure, and EP<sub>2</sub> and EP<sub>4</sub> receptors participate in PGE<sub>2</sub>-induced renin release (Schweda et al., 2004). At the juxtaglomerular cell level, EP<sub>2</sub> and EP<sub>4</sub> receptors were found to produce renin exocytosis (Friis et al., 2005). By measuring afferent arteriolar diameter, it was concluded that EP<sub>2</sub> receptors partially mediate PGE<sub>2</sub>-induced vasodilatation (Imig et al., 2002). It has been claimed that EP<sub>2</sub> receptors do not regulate overall renal hemodynamics, according to studies involving direct injection into mouse renal arteries and ultrasonic flowmetry to measure blood flow (Audoly et al., 2001). Intramedullary PGE<sub>2</sub> infusion resulted in EP<sub>2</sub>-mediated renal Na<sup>+</sup> excretion (Chen et al., 2008). In mice lacking EP<sub>2</sub> receptors, salt-sensitive hypertension occurs (Kennedy et al., 1999). These results suggest that EP<sub>2</sub> receptors produce natriuresis and promote normotension in a high-salt dietary regimen. Protection of renal cystic epithelial cells from apoptosis has implicated the EP<sub>2</sub> receptor in polycystic kidney disease (Elberg et al., 2007).

PGE<sub>2</sub> has been proposed to influence a number of CNS functions by activating EP<sub>2</sub> receptors. These range from activity-dependent synaptic plasticity against oxidative stress and acute excitotoxicity to a role in the development of COX-2-induced neurotoxicity. Evidence for EP<sub>2</sub> and EP<sub>3</sub> receptor involvement in long-term potentiation and long-term depression has been advanced;  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid receptor trafficking in the postsynaptic membrane was impli-

cated as an underlying mechanism (Andreasson, 2010). The behavioral phenotype in EP<sub>2</sub> receptor knockout mice was associated with a deficit in hippocampal long-term depression (Savonenko et al., 2009). EP<sub>2</sub> receptors have also been implicated in PGE<sub>2</sub> pain transmission (Ahmadi et al., 2002; Harvey et al., 2004; Reinold et al., 2005), the mechanism involving blockade of inhibitory glycine receptors. Calcitonin-gene related peptide (CGRP) release from trigeminal neurons may be evoked by EP<sub>2</sub>, DP<sub>1</sub>, and IP receptors (Jenkins et al., 2001).

Because both EP<sub>2</sub> and EP<sub>4</sub> receptors are G<sub>s</sub>-coupled G-protein-coupled receptors, it is not surprising that they share certain neurological functions. These include modification of tetrodotoxin-resistant Na<sup>+</sup> currents in neonatal rat nodose ganglia (Matsumoto et al., 2005) and protection against oxidative stress and amyloid  $\beta$ -peptide neurotoxicity (Echeverria et al., 2005). Nevertheless, there are several instances in which EP<sub>2</sub> receptors are the singular cognate receptor mediating PGE<sub>2</sub>-mediated events.

EP<sub>2</sub> receptor activation protects neurons against NMDA-receptor-mediated cytotoxicity, according to in vitro (Akaike et al., 1994; McCullough et al., 2004; Liu et al., 2005; Ahmad et al., 2006b) and living animal studies (Ahmad et al., 2006). EP<sub>2</sub> receptors have actually been claimed to exacerbate NMDA-mediated cytotoxicity in the very same cells, cultured rat cortical neurons (Takadera and Ohyashiki, 2006), in which EP<sub>2</sub>-mediated neuroprotection was originally described (Akaike et al., 1994). In hippocampal neurons, caspase-dependent apoptosis seems to be produced by PGE<sub>2</sub> acting through EP<sub>2</sub> receptors (Takadera et al., 2004). Nevertheless, the majority of studies on neuronal cytotoxicity suggest that EP<sub>2</sub> receptors are neuroprotective. In neuronal cells, EP<sub>2</sub> signaling affords significant neuroprotection by a cAMP-PKA mechanism, whereas microglial EP<sub>2</sub> receptor activation may lead to secondary neurotoxicity by up-regulating proinflammatory genes (Andreasson, 2010). Likewise, EP<sub>2</sub> receptors may contribute to either neuroprotection or neurotoxicity by inducing brain-derived neurotrophic factor release from microglial cells and astrocytes (Hutchinson et al., 2009). In astrocytes, it was reported that EP<sub>2</sub> receptor stimulation elicits Ca<sup>2+</sup> release from intracellular stores (Di Cesare et al., 2006).

A proinflammatory neurotoxic function for EP<sub>2</sub> receptors has emerged during the past few years, notably with respect to innate immunity. LPS has been used to model innate immunity, because binding to CD14 and toll-like receptor 4 up-regulates pro-inflammatory genes, including COX-2 (Andreasson, 2010). EP<sub>2</sub> receptors are highly inducible in the cerebral cortex and hippocampus (Zhang and Rivest, 1999). In hippocampal slice preparations, EP<sub>2</sub> receptors exacerbate LPS-induced neurotoxicity (Wu et al., 2007). EP<sub>2</sub>, and EP<sub>1</sub>, receptors are very important in Toll-like receptor 4-induced depletion of intermediate progenitor cells destined for maturation in the hippocampal subgranular



zone (Keene et al., 2009). In terms of hippocampal neurotransmission, postsynaptically synthesized PGE<sub>2</sub> modulates transmission via presynaptic EP<sub>2</sub> receptors (Sang et al., 2005). Activated microglial EP<sub>2</sub> receptors seem to play a central role in the generation of reactive oxygen species and secondary neurotoxicity (Andreasson, 2010). LPS-induced cerebral oxidative damage was selectively abolished in EP<sub>2</sub> gene deletion studies (Montine et al., 2002). Additional, mostly deleterious, EP<sub>2</sub> effects on microglia include  $\alpha$ -synuclein neurotoxicity (Jin et al., 2007), regulation of amyloid precursor protein (Pooler et al., 2004; Liang et al., 2005a), and inhibition of ATP-induced microglia migration (Nagano et al., 2008).

Neuroprotection has also attracted attention in vision research because sight is entirely dependent on optimal ocular and central neurotransmission. Experiments have been largely designed to assess vision-sparing potential in glaucoma (Woodward, 2000) and retinal disease (Andrade da Costa et al., 2009; Mori et al., 2009; Osborne et al., 2009). The underlying EP<sub>2</sub>-regulated signaling mechanisms involved in ocular neuroprotection have received little attention but are probably likely similar to those reported for CNS neurons. The link between sight-threatening ocular diseases and immune regulation is tenuous at best, and no attention has been paid to innate immunity.

Generally EP<sub>2</sub> receptors tend to play anti-inflammatory roles, which is in contrast to their participations in innate immunity in the CNS. EP<sub>2</sub> receptors inhibit T-cell proliferation according to MLR studies (Nataraj et al., 2001), regulate antigen-presenting cell function (Nataraj et al., 2001), inhibit TNF $\alpha$  release from bone marrow-derived dendritic cells (Vassiliou et al., 2003), inhibit major histocompatibility complex class II expression in dendritic cells (Harizi et al., 2003), suppress IFN $\alpha$  release by natural killer cells (Walker and Rondono, 2004), are particularly effective in inhibiting Th1 and Th2 polarized antigen-specific T-cell responses (Okano et al., 2006), and augment the signaling and function associated with the anti-inflammatory cytokine IL-10 (Cheon et al., 2006). Several immune regulatory functions assigned to EP<sub>2</sub> receptors are shared with EP<sub>4</sub>. These include Th1 differentiation (Yao et al., 2009), Th2 polarization (Kubo et al., 2004), and Th17 differentiation (Boniface et al., 2009; Napolitani et al., 2009; Narumiya, 2009; Yao et al., 2009). It should also be noted that numerous immunomodulatory effects of PGE<sub>2</sub> are reported but not pharmacologically defined. Studies in living animals seem somewhat consistent with findings on immune competent cells: EP<sub>2</sub>(-/-)mice treated with an EP<sub>4</sub> antagonist exhibited a therapeutic effect in the collagen-induced arthritis model (Narumiya, 2009), which is consistent with in vitro T-cell studies.

In polymorphonuclear leukocytes and monocytes/macrophages, EP<sub>2</sub> receptors largely exert down-regulatory functions. EP<sub>2</sub> receptors inhibit neutrophil chemotaxis, superoxide generation, LTB<sub>4</sub> release, and

aggregation (Nials et al., 1993; Wheeldon and Vardey, 1993; Talpain et al., 1995; Yamane et al., 2000). LPS-stimulated murine peritoneal neutrophils also secrete cytokines, and EP<sub>2</sub> receptor activation augments IL-6 and granulocyte cell-stimulating factor release (Yamane et al., 2000; Sugimoto et al., 2005) but suppresses TNF $\alpha$  production to some extent (Yamane et al., 2000). Efferocytosis, ingestion of apoptotic cells by phagocytes, triggers the release of cytokines, NO, and PGE<sub>2</sub>. Although this process may recover tissue homeostasis after injurious stimuli, it renders the lung, in particular, susceptible to secondary infection, because alveolar macrophages are impaired by apoptotic cells. PGE<sub>2</sub>, acting via EP<sub>2</sub> receptors, mediates efferocytosis-induced inhibition of pulmonary macrophage antibacterial function (Medeiros et al., 2009). Such an effect may be mitigated by EP<sub>2</sub>-mediated monocyte/macrophage survival against free radical damage by peroxynitrite, a superoxide-nitric oxide coupling product that is an extremely reactive free radical (Tommasini et al., 2008). More pertinent to asthma and other type 1 allergies, EP<sub>2</sub> receptors inhibit lung mast cell degranulation (Kay et al., 2006) and eosinophil trafficking (Sturm et al., 2008).

PGE<sub>2</sub> has been implicated in osteoclast, chondrocyte, and synoviocyte function. EP<sub>2</sub> and EP<sub>4</sub> receptors often mediate similar effects, but responses to EP<sub>4</sub> stimulation are typically more prominent. Small animal cell lines are used extensively, but results are contradictory (Graham et al., 2009). EP<sub>2</sub> effects include osteoblast differentiation, osteoclast-induced bone resorption, and bone anabolic activity in living animal studies (Graham et al., 2009). It is also pertinent to note that PGE<sub>2</sub> strongly inhibits human osteoclast formation (Take et al., 2005). In human and rat chondrocytes, EP<sub>2</sub> receptors have quite opposite effects on proteoglycan accumulation: suppression (Li et al., 2009b) and enhancement (Miyamoto et al., 2003), respectively. EP<sub>2</sub> receptors have been identified in synovial fibroblasts obtained from rheumatoid arthritis tissue specimens (Kojima et al., 2009).

Studies on EP<sub>2</sub> receptor function in fibroblasts usually involves fetal or patient-derived lung fibroblasts. EP<sub>2</sub> receptors have been shown to produce diverse effects that may limit fibrosis and scar formation. It is noteworthy that EP<sub>2</sub> receptors inhibit the transition of human lung fibroblasts to myofibroblasts, which are the hallmark of pulmonary fibrotic disease (Kolodtsick et al., 2003). Furthermore, PGE<sub>2</sub> inhibits fibroblast proliferation and collagen expression in patient-derived lung fibroblasts (Huang et al., 2007b) and migration (White et al., 2005) via EP<sub>2</sub> receptor activation. It is pertinent that bleomycin, which produces fibrogenesis, produces a loss of EP<sub>2</sub> expression in pulmonary fibroblasts with a resultant loss of the inhibitory effect of PGE<sub>2</sub> on collagen biosynthesis and proliferation (Moore et al., 2005). Finally, it is of interest that EP<sub>2</sub> receptors protect human lung fibroblasts from cigarette smoke-induced apoptosis

(Sugiura et al., 2007). Tissue destruction associated with periodontitis and healing may also involve EP<sub>2</sub> receptors expressed by fibroblasts (Noguchi et al., 2002; Weinberg et al., 2009). In view of the importance of fibroblasts in cutaneous wound healing and scar formation, dermal fibroblasts have also received attention. Maintenance of the migratory phenotype may be important for remodeling and regenerative repair, an effect achievable by EP<sub>2</sub> receptor activation (Parekh et al., 2007). Reduced EP<sub>2</sub> expression causes increased collagen synthesis in keloid fibroblasts (Hayashi et al., 2006).

The role of EP<sub>2</sub> receptors in skin tumor development has been the subject of several investigations. EP<sub>2</sub> receptors are claimed to induce COX-2 (Ansari et al., 2007) and to participate in tumor formation (Sung et al., 2006; Chun et al., 2009). Loss of EP<sub>2</sub> receptors from keratinocytes has been suggested as a mechanism for neoplastic progression resulting from a more invasive phenotype, although EP<sub>2</sub> receptors protect against UV-induced carcinogenesis (Konger et al., 2002; Brouxhon et al., 2007). The immune regulatory effects associated with EP<sub>2</sub> receptors, which may be beneficial in relieving cutaneous hypersensitivity, can result in diminished antitumor immune responses by virtue of reduced cytotoxic T-cell responses and inhibition of dendritic cell differentiation (Yang et al., 2003).

It seems to be generally accepted that COX inhibitors are of value in treating colorectal cancer (Clevers, 2006). A significant role for PGE<sub>2</sub> in producing a signaling cascade involving phosphorylation of glycogen synthase kinase 3,  $\beta$ -catenin translocation to the nucleus, and resultant Tcf/Lef transcription and COX-2 up-regulation in the development of chronic inflammation and colon cancer has been elucidated (Fujino et al., 2002; Regan, 2003) and subsequently confirmed (Castellone et al., 2005). Because EP<sub>2</sub> receptors are claimed to have a central role in colon cancer cells (Sonoshita et al., 2001; Seno et al., 2002), the role of EP<sub>2</sub> receptors in other carcinomas has been the subject of intense activity.

The EP<sub>2</sub>-dependent angiogenesis associated with mouse intestinal tumors (Seno et al., 2002) has been further studied in pulmonary endothelial cells, aortic rings, and the cornea (Kamiyama et al., 2006). It was concluded that EP<sub>2</sub> receptors are a major factor in endothelial cell motility (Kamiyama et al., 2006). EP<sub>2</sub> receptors, in addition to EP<sub>4</sub> receptors, produce angiogenesis in prostate cancer (Jain et al., 2008). EP<sub>2</sub> and EP<sub>4</sub> have also been implicated in VEGF expression and increased invasiveness of ovarian carcinoma cells by stimulating tumor-associated matrix metalloproteases (Spinella et al., 2004). EP<sub>2</sub> receptors may play a role in breast cancer by inducing aromatase (Brueggemeier et al., 2001; Subbaramaiah et al., 2008) and VEGF induction and hyperplasia in mammary tumor cells (Chang et al., 2005a,b). EP<sub>2</sub> receptor deficiency decreased the growth, angiogenesis, and metastasis of mammary tumors in mice; increased EP<sub>2</sub> receptor expression by

TGF $\beta$  increased mammary epithelial cell invasion, growth, and resistance to TGF $\beta$ -mediated cytostasis (Tian and Schiemann, 2010). Proliferation of squamous cell carcinoma has been ascribed to EP<sub>2</sub> receptors (Donnini et al., 2007; Yu et al., 2008a, 2009). PPAR $\gamma$  ligands were shown to inhibit lung carcinoma cell proliferation by suppressing EP<sub>2</sub> receptor expression (Han and Roman, 2004). In complete contrast, EP<sub>2</sub> and EP<sub>4</sub> receptors inhibit human gastric carcinoma cell lines (Okuyama et al., 2002).

*c. Gene deletion studies.* No less than three different lines of EP<sub>2</sub>( $-/-$ ) mice have been generated (Hizaki et al., 1999; Kennedy et al., 1999; Tilley et al., 1999). EP<sub>2</sub> receptor-deficient mice exhibited reduced fertility; they became pregnant and delivered at term, but the litter sizes were reduced (Hizaki et al., 1999; Kennedy et al., 1999; Tilley et al., 1999). This seems to be due to failure of the released ovum to be fertilized by virtue of incomplete expansion of the cumulus (Hizaki et al., 1999; Tilley et al., 1999); cumulus cells are needed for oocyte maturation-associated reduction in TNF $\alpha$ -stimulated gene 6 (*TSG-6*) expression, which seems to be the EP<sub>2</sub> receptor-mediated effect (Ochsner et al., 2003).

EP<sub>2</sub> receptor deletion also affects blood pressure. Resting blood pressure for mice on a normal diet is reduced but may be restored by providing excess salt in the diet (Tilley et al., 1999). Neither plasma renin activity nor renin mRNA were elevated in EP<sub>2</sub>( $-/-$ ) animals (Tilley et al., 1999). The vasodilator responses to PGE<sub>2</sub>, per se, seems to be EP<sub>2</sub> receptor-mediated, and PGE<sub>2</sub> produces considerable hypertension in EP<sub>2</sub>( $-/-$ ) mice (Kennedy et al., 1999). In one study, a high-salt diet resulted in a rapid and sustained increase in blood pressure (Kennedy et al., 1999). The degree of salt-sensitive hypertension recorded in the two studies was different and, in contrast to Tilley et al. (1999), Kennedy et al. (1999) reported an elevation in resting blood pressure. These findings are not readily explained; they may reflect different genetic backgrounds of the mice used in each study. There is also incongruence with respect to renal hemodynamics. Deletion of EP<sub>2</sub> receptors had little effect on PGE<sub>2</sub>-induced renal vasodilation and did not alter resting renal blood flow (Audoly et al., 2001). Quite different and unexpected results were obtained by monitoring afferent renal arteriolar caliber. The vasodilator response to PGE<sub>2</sub>, as measured by arteriolar diameter, in wild-type mice was not merely abolished by EP<sub>2</sub> receptor deletion but was converted to a vasoconstrictor response (Imig et al., 2002). The precise reason for this apparent discrepancy is not clear. It may reflect different technologies: ultrasonic flowmeter (Audoly et al., 2001) versus measurement of arteriolar diameter by transillumination videomicroscopy (Imig et al., 2002). EP<sub>2</sub> receptors have been shown also to be involved in natriuresis, according to gene deletion studies. Thus, a high-salt diet increases PGE<sub>2</sub> biosynthesis in the renal

medulla, which promotes EP<sub>2</sub> receptor-mediated sodium excretion (Chen et al., 2008).

The CNS has been a major focus of EP<sub>2</sub> receptor deficiency studies, where the neuroprotective properties of EP<sub>2</sub> receptors have been confirmed and extended. Thus, gene deletion resulted in a greater infarct volume in the cerebral artery occlusion-reperfusion model of transient forebrain ischemia (McCullough et al., 2004). EP<sub>2</sub> receptor deletion reduced oxidative damage and amyloid burden in a model of Alzheimer's disease, however (Liang et al., 2005a). In EP<sub>2</sub>-deficient mice, there is altered long-term synaptic plasticity in the hippocampus and impaired spatial learning (Yang et al., 2009). In a separate study using the same test (i.e., the Morris water maze to test hippocampal dependent spatial memory), EP<sub>2</sub>(-/-) mice demonstrated no deficits in spatial memory (Savonenko et al., 2009). In studies of EP<sub>2</sub> receptor involvement behavior and cognition, there were cognitive deficits in tests for fear and social memory (Savonenko et al., 2009). EP<sub>2</sub>(-/-) mice also exhibited impaired prepulse inhibition and heightened anxiety. The complex behavioral phenotype observed was attributed to a long-term depression deficit in the hippocampus (Savonenko et al., 2009). It is noteworthy that mice lacking EP<sub>2</sub> receptors completely lacked spinal hyperalgesia in response to PGE<sub>2</sub> (Reinold et al., 2005). This was identified by electrophysiology as diminished synaptic inhibition of excitatory dorsal horn neurons.

Neuronal cytotoxicity is intimately associated with innate immunity. LPS, which stimulates innate immunity by binding to CD14 and activating toll-like receptor 4, produces cerebral oxidative damage that was almost abolished in EP<sub>2</sub> receptor-deficient mice (Montine et al., 2002). Innate immunity-mediated neurodegeneration produced by LPS was subsequently determined to be critical to EP<sub>2</sub> receptors expressed by microglia (Shie et al., 2005). COX-2 and inducible nitric-oxide synthase are also implicated in LPS-induced neurodegeneration. The roles of EP<sub>2</sub> receptors in acquired immunity are arguably worthy of further study, notably because excessive concentrations of PGE<sub>2</sub> have been used in some key human cell studies. An important role for the EP<sub>2</sub> receptor in inhibition of dendritic cell differentiation and function has been claimed (Yang et al., 2003). In the MC26 tumor model, there was improved dendritic cell function and number in tumor-bearing EP<sub>2</sub>(-/-) mice (Yang et al., 2003). This reflects EP<sub>2</sub> receptor-mediated tumor immunosuppression and inhibition of the host reaction to tumor progression. In a different setting, 7,12-dimethylbenz(a)anthracene/12-O-tetradecanoylphorbol-13-acetate-induced skin tumors, EP<sub>2</sub>(-/-) mice showed suppressed skin tumor development associated with decreased proliferation, angiogenesis, inflammation, and cell survival (Sung et al., 2005). The link between COX-2 and EP<sub>2</sub> receptors is also manifest in a study on mammary hyperplasia, which was reduced in EP<sub>2</sub>(-/-) mice (Chang et al., 2005a).

Bone deposition and resorption has been a popular avenue of study using EP<sub>2</sub>-deficient mice. In cocultures of spleen and calvarial osteoblasts, the response to PGE<sub>2</sub>, or even PTH, was greatly reduced when both of these cell types were obtained from EP<sub>2</sub>(-/-) mice; the data in total suggest that EP<sub>2</sub> receptors activate osteoblastic cells to stimulate osteoclast formation (Li et al., 2000). Osteoclast formation from cells of an osteoblastic lineage requires up-regulation of RANKL and macrophage colony-stimulating factor. PGE<sub>2</sub>-induced increases in RANKL expression were reduced in cells derived from both EP<sub>2</sub>(-/-) and EP<sub>4</sub>(-/-) mice (Li et al., 2002); this was the major effect observed. A biphasic effect of PGE<sub>2</sub> on osteoclast formation was apparent, the secondary stimulatory effect being EP<sub>2</sub> receptor-mediated (Ono et al., 2005). Finally, in EP<sub>2</sub>(-/-) mice, bone exhibited weak biomechanical properties compared with that obtained from wild-type control mice (Akhter et al., 2001).

*d. Agonists and antagonists.* Misoprostol (Collins et al., 1985), in which the 15-hydroxyl group of PGE<sub>1</sub> is displaced to C16, shows modest selectivity for EP<sub>2</sub> and EP<sub>3</sub> receptors, whereas the related butaprost (TR4979) is EP<sub>2</sub>-selective (Gardiner, 1986). However, tissue-dependent hydrolysis of its C1 ester is required to obtain full bioactivity (*K*<sub>i</sub>: butaprost, 3500 nM; butaprost-free acid, 91 nM for human rc-EP<sub>2</sub> receptor; Abramovitz et al., 2000). There has been some confusion over the configuration at C16 of butaprost used in the original reports (see <http://www.caymanchem.com/app/template/Product.vm/catalog/10006045> for resolution of this issue). Commercially available butaprost-FA (Fig. 4) is the more active (racemic) 16*S* epimer. The "2-series" analog of butaprost-FA (CAY-10399) also shows high EP<sub>2</sub>/EP<sub>4</sub> selectivity and is devoid of the IP agonism present in butaprost-FA (Tani et al., 2002a,b). Retaining the 16*S* configuration and replacing the 9-ketone with a β-chloro group generated highly potent and selective EP<sub>2</sub> agonists (Tani et al., 2002a,b). One of these, ONO-AE1-259 (Fig. 4), has proved valuable in characterizing inhibitory EP<sub>2</sub> systems (Cao et al., 2002; Clarke et al., 2004) and has proved useful when both EP<sub>4</sub> and IP receptor systems are also present (Jones and Chan, 2005).

Acyclic derivatives of PGE<sub>1</sub> with a *N*<sup>8</sup>-methylsulfone structure exhibited weak, but selective, EP<sub>2</sub> agonism (Jones et al., 1977). CP-533536 (Fig. 4) is a related nonprostanoid with subnanomolar EC<sub>50</sub> for the rat rc-EP<sub>2</sub> receptor (Li et al., 2003; Cameron et al., 2009). EP<sub>2</sub> full agonism was also found for another nonprostanoid (Fig. 4, compound 9; from Belley et al., 2005) during screening for EP<sub>3</sub> antagonism.

There are no potent and selective EP<sub>2</sub> receptor antagonists available. AH-6809 (Fig. 5) remains the most useful compound in this class (Woodward et al., 1995b), despite significant activity at EP<sub>1</sub> receptors (Coleman et al., 1994b). Presumably, there has been inadequate ra-



tionale and commercial incentive to design potent and selective EP<sub>2</sub> antagonists. Potential uses, such as for innate immunity and secondary pulmonary infection, would likely be effectively treated by COX-2 inhibitors.

*e. Therapeutics.* The therapeutic potential of EP<sub>2</sub> agonists has been under consideration for more than 2 decades; so far, however, no clinically useful drugs have emerged. Indications for EP<sub>2</sub> agonist therapy involving relief of smooth muscle spasm have been known for at least 2 decades. These include tocolysis, for the prevention of preterm labor and dysmenorrhoea (Senior et al., 1993; Duckworth et al., 2002) and bronchodilatation for asthma treatment (Gardiner, 1986; Nials et al., 1993), which would be analogous to  $\beta_2$ -adrenoceptor therapy for relieving bronchospasm. An additional dimension to EP<sub>2</sub> agonist therapy is the inhibitory effect on eosinophil infiltration (Sturm et al., 2008) and lung mast cell degranulation (Kay et al., 2006). AH 13205 was not a clinical success and caused airway irritation in human volunteers (Nials et al., 1993). Although highly selective, AH 13205 is not a potent EP<sub>2</sub> agonist (Regan et al., 1994b). Perhaps asthma therapy should be revisited using the more potent compounds that have been discovered during the past few years.

Bone anabolic therapy with both EP<sub>2</sub> and other prostanoid agonists has been studied for several years (Hartke and Lundy, 2001; Graham et al., 2009). EP<sub>2</sub> agonists, even within the prostanoid class of drugs, are not the only options for treating osteoporosis and enhancement of fracture healing. Likewise, several prostanoid receptors mediate ocular hypotension. EP<sub>2</sub> agonists are, however, particularly efficacious at lowering intraocular pressure, and even butaprost can restore laser-induced ocular hypertension to an ocular normotensive state (Nilsson et al., 2006). A novel series of EP<sub>2</sub> agonists of a nonprostanoid structure have been reported to be extraordinarily potent, efficacious, and long-acting (Coleman and Middlemiss, 2009). The potential therapeutic utility of EP<sub>2</sub> agonists is summarized in Table 5.

### 3. EP<sub>3</sub> Receptors.

*a. Second messenger signaling.* Pharmacological characterization of EP<sub>3</sub> receptors revealed smooth muscle contractility (Coleman et al., 1994b) and pertussis toxin sensitivity (Sonnenburg et al., 1990), which pre-

dicted a more promiscuous G protein-coupling repertoire. This has been proven to be the case to some extent. The major signaling pathway for EP<sub>3</sub> receptors is G<sub>i</sub>-induced adenylate cyclase inhibition (Narumiya et al., 1999). Numerous alternatively spliced EP<sub>3</sub> variants have been identified (Sugimoto et al., 1993; Breyer et al., 1994; Regan et al., 1994a; Takeuchi et al., 1994; Kotani et al., 1995; Schmid et al., 1995; Kotelevets et al., 2007). EP<sub>3</sub> mRNA splicing variants are reported to subserve diverse receptor functions. The EP<sub>3</sub> receptor carboxyl terminus is essential for G protein activation (Irie et al., 1994) and alternative splicing variants thereof determine agonist and constitutive G<sub>i</sub> activity (Hasegawa et al., 1996; Negishi et al., 1996; Hizaki et al., 1997; Jin et al., 1997). Moreover, these alternative splicing variants confer a wide repertoire of signaling pathways. In addition to decreasing cAMP levels, PI turnover, and increased intracellular Ca<sup>2+</sup> have been reported for EP<sub>3</sub> isoforms (Namba et al., 1993; An et al., 1994; Takeuchi et al., 1994; Schmid et al., 1995; Yamaoka et al., 2009). EP<sub>3</sub> isoforms also activate Rho via pertussis toxin-sensitive G protein(s) (Katoh et al., 1996; Hasegawa et al., 1997), via G<sub>12</sub> and possibly G<sub>13</sub> (Hasegawa et al., 1997; Macias-Perez et al., 2008). EP<sub>3</sub> receptor stimulation produces neurite retraction via small GTPase Rho (Katoh et al., 1996). A role for Rho kinase in EP<sub>3</sub> receptor-induced smooth muscle contraction was indicated by agreement between the potencies of Rho kinase inhibitors for suppression of sulprostone-induced contraction in guinea pig aorta and their reported potencies on the isolated enzyme system (Shum et al., 2003). However, in the case of "silent" EP<sub>3</sub> contraction/synergism in the rat femoral artery preparation, Rho kinase inhibition suppressed both the priming agent response and the enhanced response to sulprostone, implicating Rho kinase as a common late-stage transduction process (Hung et al., 2006). Synergism between agonists on smooth-muscle EP<sub>3</sub> agonist-dependent G<sub>s</sub> activity also occurs (Negishi et al., 1996), which seems to depend on interaction between the arginine residue in the seventh transmembrane-spanning segment and the carboxylate anion of the ligand (Negishi et al., 1995), as reported for the EP<sub>3</sub>D isoform. The differential function of EP<sub>3</sub> mRNA splicing variants even extends to receptor internalization (Bilson

TABLE 5  
Potential therapeutic application of EP<sub>2</sub> agonists

Agonist	Route	Dose	Species	Experimental Model	Indication	Reference
AH 13205	Aerosol	0.1–3 nM	Guinea pigs	Histamine induced bronchoconstriction	Asthma	Nials et al., 1993
Butaprost	Tissue culture vs. oxytocin	10 <sup>-6</sup> M infusion	Human	Isolated human uterine specimens from pregnant and nonpregnant donors	Prevention of preterm labor; dysmenorrhoea	Duckworth et al., 2002
Butaprost	Topical (ocular)	0.1%	Monkey	Laser-induced glaucomatous monkey	Glaucoma	Nilsson et al., 2006
CP 533536	Local implant	10, 50, 100 mg/ml	Dog	Canine ulnar critical defect	Fracture healing	Paralkar et al., 2003
CP 533536	Topical (ocular)	0.01–0.1%	Dog, monkey	Laser-induced glaucomatous monkey	Glaucoma	Woodward and Chen, 2007

et al., 2004) and membrane targeting (Hasegawa et al., 2000). No consistent nomenclature for alternative splicing variants of EP<sub>3</sub> or other prostanoid receptors has been agreed upon.

*b. Distribution and biological functions.* In situ hybridization, Northern blotting, and functional studies have revealed a wide distribution for EP<sub>3</sub> receptors. They seem to play important roles in the CNS, cardiovascular system, reproductive system, kidney, and urinary bladder. In common with most other prostanoid receptors, EP<sub>3</sub> receptors have been implicated in cancer and inflammation/immune regulation.

EP<sub>3</sub> receptors are widely expressed in the CNS and specifically localized to neurons (Sugimoto et al., 1994b). EP<sub>3</sub> mRNA was most abundant in neurons in the sensory ganglia (Sugimoto et al., 1994b). This would be consistent with studies on acute herpetic pain, where PGE<sub>2</sub> content is increased in the dorsal root ganglia and analgesia was produced by an EP<sub>3</sub> antagonist and in EP<sub>3</sub>(-/-) mice (Takasaki et al., 2005). Gene deletion studies have also revealed a role for EP<sub>3</sub> receptors in PGE<sub>2</sub>-induced hyperalgesia (Minami et al., 2001), LPS enhanced acetic acid-induced writhing (Ueno et al., 2001), and allodynia produced by intrathecal administration of HIV-1 glycoprotein gp120 (Minami et al., 2003). A mechanistic study on sensory ganglia demonstrated that EP<sub>3</sub> receptors attenuate desensitization of B2 receptors, thereby restoring the response to bradykinin (Kozaki et al., 2007). EP<sub>3</sub> receptors located in higher centers have been implicated in mediating hyperalgesia (Oka et al., 1994; Hosoi et al., 1997; Oliva et al., 2006).

Aspirin has been used to reduce fever for over a century, now a detailed insight into the pharmacology is available. EP<sub>3</sub> receptors play a major role in hyperpyrexia. An impaired febrile response in EP<sub>3</sub>(-/-) mice was reported (Ushikubi et al., 1998). EP<sub>3</sub> receptors in the median preoptic nucleus have been demonstrated as critical for sickness-induced fever (Lazarus et al., 2007; Furuyashiki and Narumiya, 2009). Studies on EP receptor-specific ligands also implicate EP<sub>3</sub> receptors in producing fever but, somewhat surprisingly, also implicate EP<sub>1</sub> receptors (Oka et al., 2003b). Subsequent studies have focused on EP<sub>3</sub> receptors. EP<sub>3</sub> receptors have been proposed to mediate brown adipose tissue thermogenesis, although supportive pharmacological evidence is lacking (Yoshida et al., 2003). The preoptic area expresses EP<sub>3</sub> receptors that provide direct pyrogenic input to two hyperpyrexia generating sympathoexcitatory brain regions, the dorsomedial hypothalamus (Nakamura et al., 2005), and the rostral raphe pallidus nucleus (Nakamura et al., 2009). It has been suggested that EP<sub>3</sub> receptors cause a decrease in preoptic GABA<sub>A</sub> expression as a mechanism for PGE<sub>2</sub>-induced fever (Tsuchiya et al., 2008).

The abundance (Sugimoto et al., 1992) and widespread distribution of EP<sub>3</sub> receptors in the brain (Sug-

imoto et al., 1994b) point to multiple CNS functions and even opportunities for novel therapies. This opportunity does not seem to have been pursued to its fullest extent. Nevertheless, some progress has been made. EP<sub>3</sub> receptor mRNA was found to be associated with monoaminergic neurons in the brainstem, and EP<sub>3</sub> receptors were postulated as performing a modulatory function (Narumiya et al., 1999). In mouse cerebral cortex slices, EP<sub>3</sub> and histamine H<sub>3</sub> receptors located presynaptically inhibited norepinephrine in a partially exclusive manner (Schlicker and Marr, 1997), but beyond this EP<sub>3</sub>, modulation of monoamine neurotransmitter release does not seem to have been diligently pursued. One study points to PGE<sub>2</sub>-activated sympathetic nerve activity in the brain stem, with resultant tachycardia and hypertension (Ariumi et al., 2002). Behavioral suppression produced by IL-1 $\beta$  on naloxone-induced withdrawal jumping in morphine-dependent mice (Nakagawa et al., 1995) and  $\Delta^8$ -tetrahydrocannabinol on lever-pressing behavior (Yamaguchi et al., 2004) both seem to involve PGE<sub>2</sub> acting through EP<sub>3</sub> receptors. Finally, reduced brain injury has been reported after cerebral ischemia in mice lacking EP<sub>3</sub> receptors (Saleem et al., 2009b) and ischemic excitotoxicity was reduced by the EP<sub>3</sub> antagonist ONO-AE3-240 (Fig. 5) (Ikeda-Matsuo et al., 2010).

The major foci of cardiovascular EP<sub>3</sub> receptor research have been myocardial injury and platelets. The cardiovascular system, however, may provide a large-scale operational model of EP<sub>3</sub> receptor influence on monoaminergic neuronal function in the CNS. This would be provided by the pithed rat model, where presynaptic EP<sub>3</sub> receptors inhibit the release of catecholamines and the resultant vasopressor response (Malinowska et al., 1994). Platelet aggregation studies have largely concentrated on TP and IP receptors, but EP<sub>3</sub> receptors seem to have a more subtle role in platelet aggregation. PGE<sub>2</sub> exerts a dual action on platelets, inhibition at high doses, and potentiation of the effect of proaggregatory agents (Armstrong, 1996). This potentiating effect was ascribed to EP<sub>3</sub> receptors (Matthews and Jones, 1993), which has been amply confirmed (Fabre et al., 2001; Ma et al., 2001; Gross et al., 2007; Singh et al., 2009). EP<sub>3</sub> agonists have been shown to reduce infarct size and reduce myocardial injury (Zacharowski et al., 1999; Hohlfeld et al., 2000), with supportive evidence from cardiospecific EP<sub>3</sub> receptor overexpression (Martin et al., 2005). In the kidney, an EP<sub>3</sub> vasoconstrictor effect has been observed for the intralobular arteries (van Rodijnen et al., 2007).

In 2008, a series of articles implicated EP<sub>3</sub> receptors in bladder micturition. First, a report on increased bladder capacity in EP<sub>3</sub>(-/-) mice was reported (McCafferty et al., 2008). Infusion of the EP<sub>3</sub> agonist GR 63799X into the bladder of wild-type mice reduced bladder capacity, implicating EP<sub>3</sub> receptors as a contributing factor to overactive bladder (McCafferty et al., 2008). At the same time, two studies on peripheral and central neuronal

control of bladder function were published. The EP<sub>3</sub> antagonist DG-041 (Fig. 5) selectively inhibited responses of mechanosensitive afferent nerves to urinary bladder distension, inhibited the visceromotor response to bladder distension, and reduced the frequency of rhythmic bladder motility (Su et al., 2008a). A second study demonstrated that intrathecal or intracerebroventricular administration of potent EP<sub>3</sub> antagonists reduced the frequency of bladder contractions, albeit not their amplitude (Su et al., 2008b). The visceromotor reflex response was more effectively inhibited by intrathecal dosing, suggesting that bladder nociception primarily involves spinal EP<sub>3</sub> receptors (Su et al., 2008b). The potential utility of EP<sub>3</sub> antagonists for treating detrusor hyperactivity and pain associated with bladder disorders may be offset by a study showing that EP<sub>3</sub> receptors cause hypercontractility in obstructed urethra (Ankem et al., 2005).

EP<sub>3</sub> receptor mRNA has been found in parietal and chief cells of the gastric fundic epithelium (Narumiya et al., 1999). This is consistent with EP<sub>3</sub>-mediated inhibition of gastric acid secretion (Bunce et al., 1991; Perkins et al., 1991; Savage et al., 1993; Yokotani et al., 1996; Kato et al., 2005; Dey et al., 2006) in rats. This EP<sub>3</sub> inhibitory mechanism seems widely held to be true for humans, although actual functional evidence cannot be found. EP<sub>3</sub> receptors, however, have been shown to be expressed throughout the human gastric epithelium (Takafuji et al., 2002). It was proposed that EP<sub>3</sub> receptors may control gastrointestinal smooth muscle contraction because expression was found in longitudinal smooth muscle and in neurons of the myenteric ganglia (Narumiya et al., 1999). EP<sub>3</sub> receptors increase slow-wave peristaltic frequency in mice (Forrest et al., 2009). The presence of EP<sub>3</sub> receptors in the duodenum seems essential for HCO<sub>3</sub><sup>-</sup> secretion to counteract acid-induced mucosal damage (Takeuchi et al., 1999). Species differences may occur in the gastrointestinal functions of prostanoïd receptors (Dey et al., 2006).

In cancer, EP<sub>3</sub> receptors have been found to exert both facilitatory and inhibitory effects. In human colon cancer specimens, EP<sub>3</sub> receptor mRNA was reduced by 28% compared with the normal colon mucosa (Shoji et al., 2004). Studies in the human colon cancer cell line HCA-7 showed that a selective agonist decreased viable cell numbers by 30%, providing supportive evidence for EP<sub>3</sub> receptor down-regulation playing a permissive role in colon carcinogenesis (Shoji et al., 2004). In a detailed study on individual mRNA splicing variants of the EP<sub>3</sub> receptor, overexpression of each individual isoform decreased tumorigenic potential in cell lines and stably transfected human embryonic kidney 293 or HCT 116 cells exhibited decreased tumor growth in vivo (Macias-Perez et al., 2008). Analysis of second-messenger signaling revealed a G<sub>12</sub>-RhoA pathway for all three variants (Macias-Perez et al., 2008). EP<sub>3</sub> receptors decreased aromatase activity in human adipose stromal cell lines,

pointing to an inhibitory role in breast cancer (Richards and Brueggemeier, 2003). A role for EP<sub>3</sub> receptors in tumor development is supported by other studies. In a model of tumor-stromal angiogenesis, tumor growth and angiogenesis were inhibited in EP<sub>3</sub>(-/-) mice and by a selective antagonist (Amano et al., 2003). A role for EP<sub>3</sub> receptors in up-regulating VEGF (Amano et al., 2003) has been confirmed with respect VEGF and VEGF receptor expression (Taniguchi et al., 2008; Amano et al., 2009). In addition, a contributory role for EP<sub>3</sub> receptors has been suggested in oral squamous cancer cell growth (Hoshikawa et al., 2009).

Studies on edema formation have yielded tissue-specific results. In edema formation in rat and mouse skin, EP<sub>3</sub> receptor stimulation potently inhibited the response to zymosan-activated serum and platelet-activating factor (Ahluwalia and Perretti, 1994). PGE<sub>2</sub>-induced mouse paw edema was EP<sub>3</sub> receptor-mediated (Claudino et al., 2006). In the rat adjuvant arthritis model, synoviocytes expressed the EP<sub>3</sub>B isoform, which mediated FL-6 release (Kurihara et al., 2001). EP<sub>3</sub> antagonists are also reported to be active in standard models of pain and inflammation (Jones et al., 2009).

Inflammatory cell studies seem largely restricted to T cells and mast cells. Prolactin stimulates Ig and cytokine release from T cells and PGE<sub>2</sub> enhances prolactin transcription via EP<sub>3</sub> and EP<sub>4</sub> receptors (Gerlo et al., 2004). EP<sub>3</sub> receptors also regulate expression and release of matrix metalloproteinase-9 in early T cells (Zeng et al., 1996). Both of these studies relied on cultured cell lines. The influence of EP<sub>3</sub> receptors on mast-cell function seem of greater pathophysiological significance. The first studies employed mouse bone marrow derived mast cells, which expressed all four EP receptor subtypes (Nguyen et al., 2002). However, only EP<sub>3</sub> receptors were involved in potentiation of antigen-induced degranulation and IL-6 production (Nguyen et al., 2002). The importance of EP<sub>3</sub> receptors in allergic airway inflammation was demonstrated by 1) suppression of eosinophil infiltration and antigen-induced mediator release by the EP<sub>3</sub> agonist ONO-AE-248 (Fig. 4) and 2) development of a more prominent eosinophil and mononuclear cell infiltrate and increased cytokine release in EP<sub>3</sub>(-/-) mice (Kunikata et al., 2005). In dermal mast cells, EP<sub>3</sub> receptor-induced degranulation occurred only in older mice, suggesting reprogramming with age (Nguyen et al., 2005). Studies in human mast cells provide at least partial confirmation in that PGE<sub>2</sub> potentiates IgE-mediated histamine release via EP<sub>3</sub> receptors but also EP<sub>1</sub> receptors (Wang and Lau, 2006). Studies on the recruitment of mast cell progenitors showed that EP<sub>3</sub> receptors are also chemoattractant for mast cells (Weller et al., 2007).

Beyond EP<sub>3</sub> receptor-mediated down-regulation of cutaneous mast cell function and a potential role in type 1 allergies such as urticaria, EP<sub>3</sub> receptors have also been implicated in keratinocytes and indirectly in atopic der-



matitis. This is based on increased neutrophin 4 expression in keratinocytes, a factor that may participate in the hyperinnervation that occurs in atopic dermatitis (Kanda et al., 2005). Although the choice of pharmacological agents chosen for these studies was not ideal for unambiguously distinguishing EP<sub>3</sub> from EP<sub>1</sub> involvement, the use of antisense oligonucleotides provided strong evidence for responses exclusively mediated by EP<sub>3</sub> receptors (Kanda et al., 2005). In addition, EP<sub>3</sub> receptors were shown to inhibit the growth of human primary keratinocytes (Konger et al., 2005b). Three EP<sub>3</sub> mRNA splicing variants were expressed in the intact human epidermis and keratinocytes. Predominant expression in the proliferative zone of the intact epidermis further supported involvement in regulating proliferation (Konger et al., 2005b).

Very diverse activities have been attributed to EP<sub>3</sub> receptors in the lung. EP<sub>3</sub> agonists constrict the human pulmonary artery (Qian et al., 1994). PGE<sub>2</sub>-induced sensory nerve activation, as assessed by depolarization of vagus nerves, was found to be EP<sub>3</sub> receptor-mediated (Maher et al., 2009). Apnea in human neonates has been related to increased PGE<sub>2</sub> release and EP<sub>3</sub>-mediated modulation of respiratory neurons in the brainstem (Hofstetter et al., 2007). EP<sub>3</sub> receptors may not only participate in infection (sepsis)-induced apnea (Hofstetter et al., 2007) but also may contribute to mortality caused by infection with *Streptococcus pneumoniae*, according to EP<sub>3</sub> gene deletion studies (Aronoff et al., 2009). The EP<sub>3</sub> agonist GR 63799X induced S-phase arrest and inhibited fibroblast growth (Sanchez and Moreno, 2006), which is possibly relevant to fibrotic disease of the lungs and fibrosis in general.

Prostaglandins are very important in uterine function and reproduction. EP<sub>3</sub> receptors contract the myometrium (Senior et al., 1991). Two EP<sub>3</sub> receptor isoforms were found to be expressed in the human uterus, which signaled via G<sub>i</sub> and MAP kinase (Kotani et al., 2000). EP<sub>3</sub> receptors have also been implicated in cervical ripening and misoprostol is used clinically for labor induction and cervical ripening (Sanchez-Ramos et al., 1997).

*c. Gene deletion studies.* Two independent lines of EP<sub>3</sub>(-/-) mice have been generated (Fleming et al., 1998; Ushikubi et al., 1998). The progeny of EP<sub>3</sub>(-/-) mice were normal in all respects. Mice were born at the anticipated Mendelian frequency, survived normally, and were observed to be normal. Histological examination of tissues from EP<sub>3</sub> receptor-deficient mice revealed no pathological changes (Fleming et al., 1998). Fertility and reproduction were normal in all respects (Fleming et al., 1998; Ushikubi et al., 1998). Nonsteroidal anti-inflammatory drugs have been known as antipyretics, and the generation of prostanoid receptor knockout mice has enabled further insight into the pharmacology of febrile responses. These investigations were guided by the original proposal that PGE<sub>1</sub> acted as a central mediator of fever (Milton and Wendlandt, 1970). Mice lack-

ing EP<sub>3</sub> receptors failed to mount a febrile response to PGE<sub>2</sub> or to IL-1 $\beta$  or LPS (Ushikubi et al., 1998). In contrast, PGE<sub>2</sub> induced fever in EP<sub>1</sub>(-/-), EP<sub>2</sub>(-/-), and EP<sub>4</sub>(-/-) mice (Ushikubi et al., 1998). After these studies on body temperature using a rectal probe (Ushikubi et al., 1998), febrile responses and thermoregulation in EP<sub>1</sub>(-/-) and EP<sub>3</sub>(-/-) mice were studied by telemetric measurement of core temperature (Oka et al., 2003). In this more systematic study, the EP<sub>1</sub> receptor was implicated in the febrile response. The secondary phase of the hyperthermia to LPS was blunted in EP<sub>1</sub>(-/-) mice, whereas only the initial hyperthermic phase was affected in EP<sub>3</sub>(-/-) mice (Oka et al., 2003a). Agonist studies also implicated both EP<sub>1</sub> and EP<sub>3</sub> involvement in hyperthermia (Oka et al., 2003b).

EP<sub>3</sub> receptors have also been implicated in urine production, osmolality, and bladder function. EP<sub>3</sub> receptors are not essential for regulation of urinary osmolality, but in animals treated with indomethacin, urinary osmolality was increased in EP<sub>3</sub>(+/+) but not EP(-/-) mice (Fleming et al., 1998). Enhanced bladder capacity and PGE<sub>2</sub>-induced bladder hyperactivity were reduced in mice lacking EP<sub>3</sub> receptors (McCafferty et al., 2008). Thus, EP<sub>3</sub> receptors may contribute to overactive bladder disorders. EP<sub>3</sub> receptor deletion studies have shown that the EP<sub>3</sub> receptor is essential for maintaining duodenal HCO<sub>3</sub><sup>-</sup> secretion and mucosal integrity (Takeuchi et al., 1999).

The majority of studies have implicated EP<sub>3</sub> receptors in inflammation, pain, and associated events. Mice lacking EP<sub>3</sub> receptors develop more pronounced allergic airway inflammation than wild-type or other knockout mice (Kunikata et al., 2005). Allergic inflammation was suppressed by an EP<sub>3</sub> agonist (Kunikata et al., 2005). In complete contrast, EP<sub>3</sub> receptor deletion has equally revealed proinflammatory roles. Systematic study of all EP knockout mice identified only the EP<sub>3</sub> receptor as responsible for PGE<sub>2</sub>-induced mast cell activation and associated proinflammatory signaling pathways (Nguyen et al., 2002). Arachidonic acid-induced cutaneous microvascular exudation and edema was attenuated in EP<sub>3</sub>(-/-) but not EP<sub>1</sub>(-/-), EP<sub>2</sub>(-/-), or EP<sub>4</sub>(-/-) mice (Goulet et al., 2004). Exudate formation is also partially mediated by EP<sub>3</sub> receptors in the carrageenin-induced pleurisy model (Yuhki et al., 2004). Additional studies on the lung have found that EP<sub>3</sub> receptor deletion protects against severe *S. pneumoniae* infection (Aronoff et al., 2009) and virtually abolishes PGE<sub>2</sub>-induced depolarization of isolated vagus nerves (Maher et al., 2009). EP<sub>3</sub> receptors have also been implicated in angiogenesis. Thus, in full-thickness skin wounds, EP<sub>3</sub> receptor deletion delayed wound closure, re-epithelialization, and angiogenesis (Kamoshita et al., 2006). Accordingly, CD31 and VEGF expression were reduced (Kamoshita et al., 2006). COX-2 and EP<sub>3</sub> receptors have also been found to be involved in acute herpetic pain: in contrast to EP<sub>3</sub>(-/-) mice, the allodynia and hyperalge-

sia were not altered in EP<sub>1</sub>(-/-), IP(-/-), or TP(-/-) mice (Takasaki et al., 2005). Tumor progression was also attenuated in EP<sub>3</sub>-deficient mice (Shoji et al., 2005).

As a possible connection between inflammatory signaling and obesity, effects observed in EP<sub>3</sub> receptor-deficient mice are of interest. The body weight illustration and histogram data are quite striking, the EP<sub>3</sub>-deficient mice being clearly obese compared with wild-type littermate control mice (Sanchez-Alavez et al., 2007). Adult EP<sub>3</sub>(-/-) mice were feeding more frequently during the day and developed an obese phenotype on a normal fat diet. Increased locomotor activity did not affect the obesity. Obesity was accompanied by elevated leptin and insulin levels (Sanchez-Alavez et al., 2007).

The presence of functional EP<sub>3</sub> receptors that potentiate platelet aggregation and inhibit adenylate cyclase has been recognized for some time (Matthews and Jones, 1993). Consistent with these findings, mice lacking EP<sub>3</sub> receptors show an increased bleeding tendency and decreased susceptibility to thromboembolism (Ma et al., 2001). Finally, few studies on prostanoid receptor overexpression have been reported. In transgenic mice with cardiospecific EP<sub>3</sub> receptor overexpression, increased calcineurin and NFAT activity with cardiac hypertrophy were found (Meyer-Kirchrath et al., 2009).

*d. Agonists and antagonists.* Phenoxy substitution (particularly *p*-halo-phenoxy) at C16 on the PGE template modestly promotes EP<sub>1</sub>, EP<sub>3</sub>, and FP agonism but markedly accentuates TP agonism. 16-*p*-Chlorophenoxy- $\omega$ -tetranor PGE<sub>2</sub> (ICI-80205) is a good example of these trends (Jones et al., 1982; Lawrence et al., 1992). 11-Deoxy-16-phenoxy PGE<sub>1</sub> (MB-28767) shows reasonably good EP<sub>3</sub>-versus-EP<sub>1</sub> selectivity but is still a moderately potent TP agonist (Banerjee et al., 1985; Lawrence and Jones, 1992; Lawrence et al., 1992; Boie et al., 1997). Sulprostone, in which the C1 carboxylate is converted to the acidic methylsulfonamide (Schillinger et al., 1979), has modest EP<sub>3</sub>-versus-EP<sub>1</sub> selectivity and minimal TP agonism (Coleman et al., 1987; Coleman and Sheldrick, 1989). The combination of sulprostone and 17-phenyl PGE<sub>2</sub> (Lawrence et al., 1992) has often been used to discriminate EP<sub>1</sub> and EP<sub>3</sub> receptors but is clearly not ideal. In a comprehensive structure-activity relationship study using guinea pig vas deferens and binding to mouse recombinant EP receptor subtypes, Shimazaki et al. (2000) showed that replacement of the C1-carboxylate in 13,14-didehydro-16-phenoxy PGE<sub>1</sub> by a primary alcohol group (Fig. 4, compound 7b) results in only modest loss of agonist potency coupled with high EP<sub>3</sub> selectivity; the C1-methyl ketone analog showed a similar profile.

Other routes to EP<sub>3</sub>-selective agonism have been explored. Deriving from misoprostol, SC-46275 (Fig. 4) is a highly potent and selective EP<sub>3</sub> agonist (Savage et al., 1993) used in a limited number of investigations (Jones et al., 1998); the orientation of the 16-hydroxyl is opposite that of ONO-AE-259. Again, de-esterification may

be required for full bioactivity. Finally, methylation of both 11- and 15-hydroxyls in PGE<sub>2</sub> (ONO-AE-248; Fig. 4) imparts high EP<sub>3</sub> selectivity (Okada et al., 2000; Suzawa et al., 2000), but potency is only modest (Jones et al., 2008).

The development of EP<sub>3</sub> antagonists has recently been reviewed (Jones et al., 2009) and is therefore summarized only briefly. The acryloylsulfonamides L-798106 and L-826266 emerged from a combinatorial approach (Fig. 5) (Gallant et al., 2002; Belley et al., 2005). They are both highly lipophilic with slow onsets on certain isolated smooth muscle preparations (Jones et al., 2008). The related EP<sub>3</sub> antagonist DG-041 (Heptinstall et al., 2008; Singh et al., 2009) is even more lipophilic (log *P* = 7.67); affinity is maintained when the acryloyl unit is replaced by a heterocycle (Fig. 5, Hategan et al., 2009). More water-soluble antagonists are present within the DeCode Genetics series (O'Connell et al., 2009); one of these showed faster block than L-798106/L-826266 (Jones et al., 2011). An EP<sub>3</sub> antagonist of a different class is represented by compound 49 (Fig. 5) (Asada et al., 2010); it showed good *in vivo* activity against PGE<sub>2</sub>-induced contraction of pregnant rat uterus.

*e. Therapeutics.* EP<sub>3</sub> receptor pharmacology has successfully resulted in small-molecule therapeutics in current use, both of which contain misoprostol (Collins et al., 1985). Misoprostol is used for cervical ripening and labor induction (Sanchez-Ramos et al., 1997; Woodward and Chen, 2004). Misoprostol is effective gastric ulcer therapy (Rachmilewitz et al., 1986) and is combined with a nonsteroidal anti-inflammatory agent to ameliorate gastric irritation and ulceration and is commercially available as Arthrotec (G. D. Searle, Peapack, NJ) (Woodward and Chen, 2004). Misoprostol is a potent EP<sub>3</sub> agonist but does not really possess adequate receptor selectivity for modern therapeutic application. Given the complexity of PG-mediated effects (e.g., EP<sub>3</sub> receptors and EP<sub>2</sub>/EP<sub>4</sub> receptors having opposing effects on cAMP), a highly selective agent may be preferable for new uses. A decade of recent EP<sub>3</sub> research has witnessed diverse findings. EP<sub>3</sub> agonists protect against ischemic myocardial injury and reduced infarct size (Hohlfeld et al., 1997, 2000; Zacharowski et al., 1999), culminating in a study on transgenic mice with cardiospecific EP<sub>3</sub> overexpression (Martin et al., 2005). Reduction in tumor development may be achieved with EP<sub>3</sub> agonists (Amano et al., 2003; Macias-Perez et al., 2008), and down-regulation of EP<sub>3</sub> receptors may enhance colon carcinogenesis in later stages (Shoji et al., 2004). Suppression of allergic inflammation by EP<sub>3</sub> receptor activation has also been demonstrated (Kunikata et al., 2005). Finally, EP<sub>3</sub> agonists may offer treatment for opiate withdrawal syndrome (Nakagawa et al., 1995). The potential of the EP<sub>3</sub> agonist-based therapeutic options (Table 6) must be viewed from the standpoint that EP<sub>3</sub> receptor stimulation may result in many patho-

TABLE 6  
*Potential therapeutic application of EP<sub>3</sub> agonists*

Agonist	Route	Dose	Species	Experimental Model	Indication	Reference
MB 28767	Intravenous	2 pmol · kg <sup>-1</sup> · min <sup>-1</sup>	Pig	Occlusion/reperfusion	Cardiac infarction	Hohlfeld et al., 2000
MB 28767	Intracisternal	1 fg–30 ng per animal	Mouse	Naloxone-induced withdrawal jumping	Opiate withdrawal syndrome	Nakagawa et al., 1995
ONO-AE-248	Subcutaneous	10 µg/kg	Mouse	Ovalbumin induced asthma	Asthma	Kunikata et al., 2005
TEI-3356	Intravenous	1 µg · kg <sup>-1</sup> · h <sup>-1</sup>	Rat	Occlusion/reperfusion	Cardiac infarction	Zacharowski et al., 1999

TEI-3356, 5-(7-hydroxy-6-(4-hydroxy-4-methyl-1-octenyl)bicyclo(3.3.0)oct-2-en-3-yl)pentanoic acid.

physiological events, as indicated by the potential uses of EP<sub>3</sub> antagonists.

Potential therapeutic uses of EP<sub>3</sub> receptor antagonists, identified and confirmed in animal models with compounds, are summarized in Table 7. These include pain and inflammation (Jones et al., 2009), type 1 allergy (Nguyen et al., 2002), lung infection (Aronoff et al., 2009), cough (Maher et al., 2009), overactive bladder (McCafferty et al., 2008; Su et al., 2008), hyperpyrexia (Ushikubi et al., 1998), and cancer (Amano et al., 2003). Most recently, EP<sub>3</sub> antagonists have emerged as a new target for antiplatelet agents in atherothrombotic disease, without prolonged bleeding (Heptinstall et al., 2008; Singh et al., 2009).

#### 4. EP<sub>4</sub> Receptors.

*a. Second messenger signaling.* EP<sub>4</sub> receptors are widely distributed (Narumiya et al., 1999). The pharmacologically defined EP<sub>4</sub> receptor was originally designated EP<sub>2</sub> (Honda et al., 1993; Bastien et al., 1994) until the authentic EP<sub>2</sub> receptor was cloned (Regan et al., 1994b). Studies on second-messenger signaling demonstrated functional coupling to cAMP via G<sub>s</sub> (Narumiya et al., 1999). Certain EP<sub>4</sub> receptor-mediated effects seem to exclusively employ the cAMP-PKA pathway (Southall and Vasko, 2001; Gray et al., 2004; Ziemann et al., 2006; Boniface et al., 2009). Direct comparison of EP<sub>4</sub> and EP<sub>2</sub> receptor signaling demonstrated that the functional coupling to cAMP seems less efficient for EP<sub>4</sub> compared with the EP<sub>2</sub> subtype (Fujino et al., 2002, 2005). The prospect of a second EP<sub>4</sub> signaling pathway was realized when a PI3K signaling pathway was discovered coupled to the pertussis toxin-sensitive G protein G<sub>i</sub> (Fujino et al., 2002, 2003, 2005; Fujino and Regan, 2005). This also provides a mechanism for limiting the cAMP response to EP<sub>4</sub> stimulation (Fujino and Regan, 2006). It should be noted that several cAMP-independent signaling cascades have been reported for EP<sub>4</sub> receptor activation

(Fiebich et al., 2001; Pozzi et al., 2004; Mendez and LaPointe, 2005; Frias et al., 2007; George et al., 2007; Rao et al., 2007). Evidence for the EP<sub>4</sub> signaling cascade PI3K-ERK-early growth response-1 (Fujino et al., 2003) seems operative in cell growth (Pozzi et al., 2004; Mendez and LaPointe, 2005; Frias et al., 2007; Rao et al., 2007). Serum-deprived apoptosis in Jurkat cells was reduced by EP<sub>4</sub> receptors through PI3K and cAMP pathways may be operative, as in EP<sub>4</sub>-mediated inhibition of apoptosis (Leone et al., 2007).

Epac signaling has received little attention from the standpoint of prostanoid receptor signal transduction. A recent study has indicated both PKA and Epac 1 signaling in rheumatoid synovial fibroblasts (Kojima et al., 2009) for both EP<sub>2</sub> and EP<sub>4</sub> receptors. Both EP receptor subtypes activate the small GTPase Rap 1 (Kojima et al., 2009). Rap was also involved in EP<sub>4</sub>-mediated brain natriuretic peptide expression (Qian et al., 2006). EP<sub>4</sub> receptors have also been shown to be involved in COX-2 mRNA induction and stabilization and stimulation of translation (Faour et al., 2001; Martineau et al., 2004), p38 MAP kinase playing a key role. EP<sub>4</sub> receptor activation of p38 MAP kinase has been found in certain cells: astrocytes, podocytes (Fiebich et al., 2001; Martineau et al., 2004), RAW 264.7 cells (Chen et al., 2006), Caco-2 cells (Leone et al., 2007), and colonic myofibroblasts (Hoang et al., 2007). Such an EP<sub>4</sub> effect on p38 MAP kinase is not always observed, for example in tracheobronchial epithelial cells (Gray et al., 2004) and neonatal ventricular myocytes (Qian et al., 2006). In cardiac myocytes, EP<sub>4</sub> receptor-mediated hypertrophy (Mendez and LaPointe, 2005) and BNP promoter regulation (Qian et al., 2006) both involve p42/44 MAP kinase. The potential complexities of EP<sub>4</sub> activating signaling cascades is illustrated by the concomitant activation of PKC in some instances (Fiebich et al., 2001; Chen et al., 2006).

TABLE 7  
*Potential therapeutic application of EP<sub>3</sub> antagonists*

Antagonist	Route	Dose	Species	Experimental Model	Indication	Reference
DG-041	Intravenous	10 mg/kg	Rat	Urinary bladder distension	Overactive bladder	Su et al., 2008a
DG-041	Oral	0.1–100 mg/kg	Rat	Ex vivo platelet aggregation; bleeding time	Atherothrombosis	Singh et al., 2009
L-826266	Intraperiaqueductal grey	0.125 nM–0.5 nM	Mouse	Formalin induced hyperalgesia	Pain	Oliva et al., 2006
L-826266	Intraplantar	3–30 nmol	Mouse	Paw-licking	Rheumatoid arthritis	Kassuya et al., 2007
ONO-AE3-240	Subcutaneous	3.30 mg/kg	Mouse	HSV-1 inoculation	Post-herpetic pain	Takasaki et al., 2005
ONO-AE3-240	Subcutaneous	50 nmol/tumor	Mouse	Sarcoma 180 cell tumor	Cancer	Amano et al., 2003

HSV-1, Herpes simplex virus 1.



*b. Distribution and biological functions.* EP<sub>4</sub> receptors were originally pharmacologically characterized predominantly from studies on smooth muscle (Coleman et al., 1994a). Thereafter, EP<sub>4</sub> receptor-mediated effects on smooth muscle tone have received little attention. EP<sub>4</sub> receptors mediate vasorelaxation of pulmonary arterial veins but produce no effect on pulmonary arteries (Foudi et al., 2008). EP<sub>4</sub> receptors, however, do not play an entirely passive role in the human pulmonary artery: COX-2 induction by bradykinin in human pulmonary arterial smooth muscle cells involves cAMP response element activation by EP<sub>4</sub> and EP<sub>2</sub> agonists (Bradbury et al., 2003). In the rat aorta, EP<sub>4</sub> receptor-mediated vasorelaxation was endothelium-dependent and involved endothelial nitric-oxide synthase and cGMP (Hristovska et al., 2007). The most therapeutically significant study demonstrated EP<sub>4</sub> receptor-mediated vasodilation of the human middle cerebral and meningeal arteries and obviated the potential clinical utility of EP<sub>4</sub> antagonists for treating migraine (Davis et al., 2004; Maubach et al., 2009). EP<sub>4</sub> overexpression in atherosclerotic plaque results in an unstable phenotype prone to inflammation and instability (Cipollone et al., 2005). Endothelial cell migration and angiogenesis in vivo are also produced by EP<sub>4</sub> receptor activation (Rao et al., 2007; Jain et al., 2008).

An interesting aspect of EP<sub>4</sub> cardiovascular biology is its potential involvement in cardiac hypertrophy. EP<sub>4</sub> receptor activation produces cardiomyocyte hypertrophy, as measured by increased protein synthesis (Mendez and LaPointe, 2005; Miyatake et al., 2007; He et al., 2010), cell size and surface area (Frias et al., 2007; Miyatake et al., 2007; He et al., 2010), re-expression of fetal genes, and activation of hypertrophic marker genes such as *BNP* (Qian et al., 2006; Miyatake et al., 2007; He et al., 2010). This was confirmed in living animals, where cardiospecific EP<sub>4</sub> receptor deletion resulted in decreased hypertrophy and fibrosis after experimental myocardial infarction (Qian et al., 2008). The transcription factor STAT3 was activated and correlated with hypertrophy and fibrosis (Frias et al., 2007; Qian et al., 2008). Cardiac function, paradoxically, was worsened in hearts lacking EP<sub>4</sub> receptor expression (Qian et al., 2008), which actually correlates with EP<sub>4</sub> protection from ischemia-reperfusion injury (Xiao et al., 2004).

The ductus arteriosus is a shunt in the fetus between the pulmonary artery and the aorta. Closure of the ductus arteriosus in the newborn is vital to prevent pulmonary hypertension-induced lung edema and congestive heart failure. The EP<sub>4</sub> receptor is critical for remodeling of the ductus arteriosus at birth (Nguyen et al., 1997). Likewise, EP<sub>4</sub> agonists reopen the ductus arteriosus in neonates dependent on placental oxygenation (Momma et al., 2005). Analogous to some extent, the uterine cervix also plays a crucial role in pregnancy; closed during gestation, soft and dilated during labor. PGE<sub>2</sub> has been used for inducing cervical ripening for many years

(Woodward and Chen, 2004), and the changes that occur resemble those that are observed in physiological ripening. EP<sub>4</sub> receptor expression has been shown to be maximum at parturition in rats (Chien and Macgregor, 2003) and has been implicated in LPS-induced cervical ripening (Fukuda et al., 2007). Glycosaminoglycan biosynthesis is an important part of cervical ripening, and EP<sub>4</sub> receptor stimulation produces this effect in human cervical fibroblasts via a PKA-independent pathway (Schmitz et al., 2001).

Results on EP<sub>4</sub> receptor involvement in mucus secretion are mixed. EP<sub>4</sub> and EP<sub>1</sub> receptors evoke mucin exocytosis from central mucous cells (Ohnishi et al., 2001), whereas an EP<sub>4</sub> agonist inhibited LPS-induced mucus secretion from airway epithelial cells (Hattori et al., 2008). PGE<sub>2</sub> was claimed to protect guinea pig gastric mucosal cells from ethanol-induced apoptosis (Hoshino et al., 2003): set against this finding, the EP<sub>4</sub> antagonist (S)-4-(1-(5-chloro-2-(4-fluorophenoxy)benz-amido)ethyl)benzoic acid (CJ-42794) did not damage the rat gastric mucosa or worsen the response to aspirin or stress (Takeuchi et al., 2007). Cytoprotective duodenal HCO<sub>3</sub><sup>-</sup> secretion is also EP<sub>4</sub> receptor-mediated in rats (Aoi et al., 2004) and humans (Larsen et al., 2005).

There is an extensive volume of literature on PGs and bone formation, and EP<sub>4</sub> agonists occupy a prominent place. EP<sub>4</sub> receptors mediate not only bone formation but also bone resorption, as indicated in EP<sub>4</sub>(-/-) mice (Miyaura et al., 2000). Nevertheless, a litany of reports shows EP<sub>4</sub>-mediated bone formation, augmentation of bone morphogenetic protein-induced bone mass, and beneficial effects on fracture healing. These findings suggest that, physiologically, EP<sub>4</sub> receptors favor resorption, but exogenously administered EP<sub>4</sub> agonists have an anabolic effect on bone formation. It may be speculated that exogenous EP<sub>4</sub> agonists locate receptors not under PGE<sub>2</sub> regulation under normal physiological circumstances. The potential therapeutics associated with EP<sub>4</sub> and EP<sub>2</sub> agonists for treating bone loss and accelerating bone repair have been reviewed (Li et al., 2007; Graham et al., 2009). It should also be noted that findings in vitro may not transition into in vivo studies; thus, although EP<sub>4</sub> receptors are essential for anabolic responses to PGE<sub>2</sub>, in osteoblasts, they are not essential for bone remodeling in living animals (Gao et al., 2009). In quite marked contrast, the EP<sub>4</sub> antagonist *N*-(((2-(4-(2-ethyl-4,6-dimethyl-1*H*-imidazo(4,5-*c*)pyridin-1-yl)phenyl)ethyl)amino)carbonyl)-4-methylbenzenesulfonamide (CJ-023,423) reduced bone destruction in the rat adjuvant-induced arthritis model (Okumura et al., 2008). In mouse collagen-induced arthritis, the EP<sub>4</sub> antagonist ONO-AE3-208 (Fig. 5) did not alter the arthritis in wild-type mice (Honda et al., 2006). These results indicate that EP<sub>4</sub> receptor participation in bone remodeling is disease-model specific.

Endochondral bone formation involves chondrocytes as well as coordinated bone formation and mineralized

matrix resorption by osteoblasts and osteoclasts. In the growth plate, chondrocytes undergo a maturation process. Resting chondrocytes transition into proliferating chondrocytes, which express type II collagen mRNA and synthesize proteoglycan. Chondrocytes then mature into hypertrophic cells characterized by a 5- to 10-fold increase in cell volume. Terminal differentiation is associated with expression of genes associated with calcification of bone matrix, such as osteocalcin. Terminally differentiated chondrocytes undergo apoptosis, and the calcified cartilage left behind provides a template for primary bone formation (Cheung et al., 2003). An understanding of PG involvement in each phase of chondrocyte maturation is of significance for osteoarthritis and rheumatoid arthritis. EP<sub>4</sub> receptor stimulation alone did not up-regulate type II collagen expression or increase proteoglycan in rat growth plate chondrocytes; concomitant EP<sub>2</sub> receptor activation was required (Miyamoto et al., 2003). Other murine chondrocyte studies are not entirely in agreement (Clark et al., 2005; Brochhausen et al., 2006), these studies using more physiological concentrations. The concentrations of PGE<sub>2</sub> in studies on human chondrocytes are also sufficiently high to be in the nonselective range.

Fibroblast-like synoviocytes are important in the pathogenesis of rheumatoid arthritis, where they proliferate and secrete enzymes involved in joint degradation and cytokine/chemokine production. In human synovial fibroblasts, IL-1 $\alpha$  (Yoshida et al., 2001) and IL-1 $\beta$  (Faour et al., 2001) effects are mediated by PGE<sub>2</sub> release. EP<sub>2</sub> and EP<sub>4</sub> receptors are consistently expressed in human synovial fibroblasts (Yoshida et al., 2001; Mathieu et al., 2008; Kojima et al., 2009). IL-1 $\beta$ -induced up-regulation of COX-2 was ascribed to interaction of released PGE<sub>2</sub> with EP<sub>4</sub> receptors (Faour et al., 2001). IL-1 $\beta$  also increases PGE<sub>2</sub> production in human tendon fibroblasts, which mediates down-regulation of type 1 collagen via EP<sub>4</sub> receptors, which in turn may disturb tendon homeostasis (Thampatty et al., 2007).

Studies on leukocyte EP<sub>4</sub> receptor function have been largely restricted to mononuclear cells. In mouse peritoneal neutrophils, EP<sub>4</sub> and EP<sub>2</sub> receptors suppress TNF $\alpha$  production, and EP<sub>2</sub> receptors augment IL-6 production (Yamane et al., 2000). In human eosinophils, EP<sub>4</sub> mRNA was found to be significantly higher than EP<sub>2</sub> (Mita et al., 2002). In human blood monocytes, EP<sub>2</sub> and EP<sub>4</sub> receptors up-regulated C-C chemokine receptor 7 mRNA, which is essential for migration to secondary lymphoid tissues (Côté et al., 2009). In a further study on human macrophages, only EP<sub>4</sub> receptors were expressed and therefore considered to mediate PGE<sub>2</sub> inhibition of macrophage inhibitory proteins 1 $\alpha$  and 1 $\beta$ , IL-8, monocyte chemotactic protein-1, and IL-10 release (Takayama et al., 2002).

PGE<sub>2</sub> was also found to be a key factor for increased C-C chemokine receptor 7 mRNA expression in monocyte-derived dendritic cells (Scandella et al., 2002), al-

though the relative importance of EP<sub>4</sub> and EP<sub>2</sub> receptors is uncertain because of indeterminate pharmacological definition. PGE<sub>2</sub> is regarded as essential for the development of a migratory phenotype of human dendritic cells, and this is ascribed to EP<sub>4</sub> and EP<sub>2</sub> receptor mediation (Luft et al., 2002; Harizi et al., 2003; Legler et al., 2006; McIlroy et al., 2006). Polarization into Th-2 helper cells seems to involve both EP<sub>4</sub> and EP<sub>2</sub> receptors (Kubo et al., 2004; McIlroy et al., 2006; Krause et al., 2007). The choice and concentrations of the pharmacological "tools" in these studies was not always ideal, and further investigation would be worthwhile. EP<sub>4</sub> receptors were shown to initiate cutaneous immune responses by promoting Langerhans cell maturation and migration, in a comprehensive study employing EP<sub>4</sub>(-/-) mice and selective EP<sub>4</sub> agonist and antagonist compounds (Kabashima et al., 2003b).

T-cell functions are also regulated by PGE<sub>2</sub> and EP<sub>4</sub> receptor activation. PGE<sub>2</sub> was shown to suppress Th1 and Th2 T-helper cell activity. Both EP<sub>4</sub> and EP<sub>2</sub> receptors acted additively to suppress Th1 cell proliferation and IFN- $\gamma$  release. In Th2 T-helper cells, independent EP<sub>2</sub> and EP<sub>4</sub> receptor activation virtually abolished proliferation, but combined EP<sub>2</sub>/EP<sub>4</sub> agonism was most effective in inhibiting IL-4 production (Okano et al., 2006). Some studies have implicated EP<sub>4</sub> and EP<sub>2</sub> receptors in development of the Th17 phenotype from naive T cells. Th17 T cells are distinct from Th1 and Th2 subsets. Human Th17 T-helper cell differentiation is controlled by the retinoic acid receptor-related orphan receptor- $\gamma$ t, and PGE<sub>2</sub> seems to synergize with IL-1 $\beta$  and IL-23 to up-regulate retinoic acid receptor-related orphan receptor- $\gamma$ t and down-regulate T cell-specific T box transcription factor, IFN- $\gamma$  and the anti-inflammatory cytokine IL-10 (Boniface et al., 2009; Napolitani et al., 2009). No role for EP<sub>1</sub> or EP<sub>3</sub> receptors was apparent. A clear role for EP<sub>2</sub> receptors was established, but the choice of EP<sub>4</sub> agonists and the high concentrations used cannot rule out EP<sub>2</sub>/EP<sub>4</sub> mutual inhibitory activity, EP<sub>2</sub>/EP<sub>4</sub> synergism, or other nonspecific activity. In an in vivo transplantation model, a combination of EP<sub>2</sub>, EP<sub>3</sub>, and EP<sub>4</sub> agonists was needed to match the immunosuppressive effect of PGE<sub>2</sub> (Fujimoto et al., 2005). A singular EP<sub>4</sub> receptor-mediated event was found in HIV-1-infected T cells with respect to pro-viral DNA activation (Dumais et al., 1998). Based on *Pt ger 4* (EP<sub>4</sub>) behaving as a strongly expressed, delayed early gene that inhibits B cell proliferation (Murn et al., 2008), EP<sub>4</sub> receptors could represent a novel target for treatment of B cell malignancies.

Numerous reports have linked EP<sub>4</sub> receptor activation to cancer. EP<sub>4</sub> receptors promote cancer in many dimensions: cell proliferation (Cherukuri et al., 2007; Zheng et al., 2009), cell survival (George et al., 2007), invasiveness (Spinella et al., 2004; Pan et al., 2008), angiogenesis (Jain et al., 2008), migration (Kim et al., 2010), and tumor metastasis (Ma et al., 2006; Yang et al., 2006). PGE<sub>2</sub> produces proliferation of colon cancer cell lines

H-29 (Chell et al., 2006) and HCA7 (Cherukuri et al., 2007). This is consistent with the idea that EP<sub>4</sub> is the most abundant transcript in both H-29 and HCA7 cells, which also biosynthesize PGE<sub>2</sub> via COX-2 (Doherty et al., 2009). EP<sub>4</sub> receptors stably transfected into HT-29 cells promoted anchorage-independent growth and increased resistance to apoptosis and the formation of fluid-filled cysts (Hawcroft et al., 2007). Transfection of EP<sub>4</sub> receptors into a human adenoma cell line (RG/C2) also introduces anchorage-independent growth (Chell et al., 2006). PGE<sub>2</sub> also caused EP<sub>4</sub>-mediated cell growth in untransfected RG/C2 adenoma cells (Chell et al., 2006). COX-2 (Gustafsson et al., 2007; Yuan et al., 2008; Doherty et al., 2009) and EP<sub>4</sub> receptor expression are increased in colorectal cancer (Chell et al., 2006), although the latter result was not confirmed (Gustafsson et al., 2007). Animal models provide evidence for a major role for EP<sub>4</sub> receptors in the development of colon cancer (Mutoh et al., 2002; Kitamura et al., 2003b; Yang et al., 2006). EP<sub>4</sub> receptors have been implicated in many forms of cancer: these include lung (Han et al., 2007; Zheng et al., 2009), upper urinary tract (Miyata et al., 2005), stomach (Okuyama et al., 2002), prostate (Jain et al., 2008), breast (Timoshenko et al., 2003; Ma et al., 2006; Pan et al., 2008; Robertson et al., 2008; Subbaramiah et al., 2008), cervix (Sales et al., 2001; Muller et al., 2006; Oh et al., 2009), ovary (Spinella et al., 2004), and nonmelanoma skin cancer (Lee et al., 2005).

Prostanoid EP<sub>4</sub> receptors have been extensively implicated in mediating hyperalgesia and allodynia (Jones et al., 2009). All EP subtypes are expressed in sensory neurons, but EP<sub>4</sub> may be regarded as the most important because it causes sensitization (Southall and Vasko, 2001) and is exclusively expressed in a subset of primary sensory dorsal root ganglia, which increases in subchronic inflammation (Lin et al., 2006). EP<sub>4</sub> receptors protect against NMDA-induced acute excitotoxicity (Ahmad et al., 2005). A dichotomy of effects was found with respect to EP<sub>4</sub> receptors in Alzheimer's disease. PGE<sub>2</sub> was found to stimulate amyloid- $\beta$  peptide production via EP<sub>4</sub> receptor internalization (Hoshino et al., 2009). However, cell death produced by  $\beta$ -amyloid was attenuated by EP<sub>4</sub> and EP<sub>2</sub> receptor stimulation (Echeverria et al., 2005).

*c. Gene deletion studies.* Two lines of systemic-null EP<sub>4</sub>-deficient mice were generated independently (Nguyen et al., 1997; Segi et al., 1998). Most EP<sub>4</sub>-deficient mice on a C57BL/6 background die within 3 days after birth because of marked pulmonary congestion and heart failure due to a patent ductus arteriosus. It is well known that administration of indomethacin to maternal mice during late pregnancy induces closure of the ductus in wild-type mice, which led to the proposal that endogenous PGs maintains the ductus open. However, indomethacin treatment did not induce closure of the ductus in EP<sub>4</sub>-deficient fetuses, indicating that a PG-independent dilatory mechanism comes into play in the absence of EP<sub>4</sub>. These findings led

the authors to suggest that opening of this vessel during the embryonic period is mediated by the PGE<sub>2</sub>-EP<sub>4</sub> signaling and that in its absence, the compensatory dilatory mechanism is mobilized, and this mechanism continues to maintain the vessel open after the birth, resulting in a paradoxical patent ductus arteriosus in the EP<sub>4</sub>-deficient mice. However, in contrast to this general idea on the patent ductus arteriosus phenotype of EP<sub>4</sub>(-/-) mice, Trivedi et al. (2006) suggested an alternative possibility that the PGE<sub>2</sub>-EP<sub>4</sub> signaling functions close, not open, the ductus. Their suggestion was based on their findings that COX-2 is induced in the ductus around term; that the loss of COX-2 or long-term treatment with COX inhibitors leads to its opening, not closure; and that the COX-2 expression in the ductus around term was attenuated in EP<sub>4</sub>-deficient mice.

Survival of EP<sub>4</sub>-deficient mice can be improved using the mixed genetic background C57BL/6 and 129s/v. Therefore, colonies of EP<sub>4</sub>(-/-) mice are maintained by intercrossing surviving EP<sub>4</sub>(-/-) mice, and progenies of these colonies are used with progenies of the littermate wild-type mice as a control. Using these mice, functions of EP<sub>4</sub> in several physiological processes have been examined, one being that in bone metabolism. Osteoclasts develop from precursor cells of the macrophage lineage in the bone microenvironment. Factors such as PTH, vitamin D, IL-1, and IL-6 act on osteoblasts to induce the synthesis of RANKL, which in turn stimulates the formation of mature osteoclasts from hematopoietic precursors through cell-cell interaction. These factors induce COX-2 expression in osteoblasts, and their induction of osteoclast differentiation is inhibited, at least in part, by aspirin-like drugs; such inhibition is reversed by the addition of PGE<sub>2</sub>, implicating PGE<sub>2</sub> in this process (Tai et al., 1997). Sakuma et al. (2000) and Miyaura et al. (2000) examined the identity of the EP subtype responsible for mediating this action of PGE<sub>2</sub>. Sakuma et al. (2000) found that PGE<sub>2</sub>-induced osteoclast formation was impaired in cocultures of osteoblasts from EP<sub>4</sub>-deficient mice and osteoclast precursors from the spleen of wild-type mice. IL-1 $\beta$ , TNF- $\alpha$ , and basic fibroblast growth factor also failed to induce osteoclast formation in these cultures. Miyaura et al. (2000) added PGE<sub>2</sub> to cultures of parietal bone from mice deficient in each of the EP subtypes, as well as from wild-type mice, and examined bone resorption by measuring the release of Ca<sup>2+</sup> into the medium. They found that the induction of bone resorption by PGE<sub>2</sub> was greatly impaired, whereas bone resorption in response to dibutyl cAMP was unaffected, in bone from EP<sub>4</sub>-deficient mice. These studies unequivocally established a role for EP<sub>4</sub> receptors in the induction of osteoclast differentiation factor and in PGE<sub>2</sub>-dependent bone resorption. On the other hand, Li et al. (2000) showed that the osteoclastogenic response to PGE<sub>2</sub>, PTH, or 1,25-dihydroxyvitamin D *in vitro* was impaired in cultures of cells derived from EP<sub>2</sub>-deficient mice. These findings probably reflect redundant roles of



the two relaxant PGE receptor subtypes. In addition to bone resorption, systemic administration of PGE<sub>2</sub> has long been known to induce bone formation *in vivo*. To examine the identity of the EP receptor in this process, Yoshida et al. (2002) infused PGE<sub>2</sub> into the periosteal region of the femur of wild-type mice or mice deficient in each EP subtype with a miniosmotic pump and found that, after 6 weeks, PGE<sub>2</sub> induced extensive callus formation on the femur at the site of infusion in wild-type as well as EP<sub>1</sub>-, EP<sub>2</sub>-, and EP<sub>3</sub>-deficient mice but not in EP<sub>4</sub>-deficient mice. Infusion of an EP<sub>4</sub>-selective agonist, but not those specific for other EP subtypes, consistently induced bone formation in wild-type mice with increased density of both osteoblasts and osteoclasts. These findings suggest that EP<sub>4</sub> is responsible for both bone resorption and bone formation induced by PGE<sub>2</sub> and that activation of EP<sub>4</sub> *in situ* integrates these two actions for bone remodeling.

Given the marked induction of COX-2 and high production of PGE<sub>2</sub> in the heart during myocardial infarction and preferential expression of EP<sub>4</sub> among EPs in this organ, Xiao et al. (2004) subjected EP<sub>4</sub>-deficient mice to an ischemia-reperfusion model by ligation of the left anterior descending coronary artery for 1 h, followed by 24-h reperfusion. They found that EP<sub>4</sub>-deficient mice developed larger infarct sizes than wild-type mice and that, conversely, administration of ONO-4819CD (Xiao et al., 2004), an EP<sub>4</sub> agonist, administered 1 h before or 50 min after occlusion reduced the infarct size in wild-type mice. Based on these findings, the authors suggested that EP<sub>4</sub> exerts cardioprotective actions under ischemia-reperfusion conditions in the heart. Another study using EP<sub>4</sub>-deficient mice in cardiovascular diseases concerned its role in atherosclerosis. Because macrophages that accumulate in early lesions of atherosclerosis are capable of producing a large amount of PGE<sub>2</sub>, and cells in atherosclerotic plaques express EP<sub>2</sub> and EP<sub>4</sub> among EP subtypes, Babaev et al. (2008) generated chimera mice in which fetal liver cells from either EP<sub>2</sub>(-/-) or EP<sub>4</sub>(-/-) mice were transplanted into lethally irradiated LDLR(-/-) mice. They then fed these mice with western chow for 8 weeks to develop atherosclerosis. They found that atherosclerotic lesions were significantly smaller in mice transplanted with cells of EP<sub>4</sub>-deficient mice [EP<sub>4</sub>(-/-) → LDLR(-/-)] than either WT → LDLR(-/-) or EP<sub>2</sub>(-/-) → LDLR(-/-) mice with significantly more apoptotic cells in the lesions. Further analysis revealed that macrophages from EP<sub>4</sub>(-/-) chimeras are more sensitive to apoptotic stimuli, which may derive from attenuated PI3K and nuclear factor-κB signaling in this line. These findings have led the authors to suggest that the macrophage EP<sub>4</sub> signaling may be a target for suppressing development of atherosclerosis.

Studies on EP<sub>4</sub>-deficient mice also revealed a variety of actions of this EP subtype in immune inflammation. One action is on dendritic cells. Kabashima et al. (2003b) found that migration of Langerhans cells in the skin to

draining lymph nodes on hapten application is impaired in EP<sub>4</sub>(-/-) mice, and this impairment can be mimicked by treatment of animals with an EP<sub>4</sub>-selective antagonist. Consequently, contact hypersensitivity to the hapten was suppressed in EP<sub>4</sub>-deficient mice. Further analysis revealed that the PGE<sub>2</sub>-EP<sub>4</sub> signaling facilitates mobilization, migration, and maturation of Langerhans cells after initial antigen application. Honda et al. (2006) found that EP<sub>4</sub> also plays a role in collagen-induced arthritis. In this model, the PGI<sub>2</sub>-IP signaling and the PGE<sub>2</sub> signaling through EP<sub>2</sub> and EP<sub>4</sub> additively mediate joint inflammation through regulation of expression of arthritis-related genes, including those for IL-6, vascular endothelial growth factor-A, and RANKL, in synovial fibroblasts. More recently, Yao et al. (2009) used T cells from mice deficient in EP<sub>2</sub> or EP<sub>4</sub> and found that EP<sub>2</sub> and EP<sub>4</sub> receptors redundantly facilitate IL12-mediated Th1 differentiation and IL-23-mediated Th17 expansion. EP<sub>4</sub> also functions in production of IL-23 from activated dendritic cells. Development of immune inflammation in experimental allergic encephalomyelitis is consistently and significantly suppressed by treatment of mice with EP<sub>4</sub>-selective antagonist or in EP<sub>4</sub>-deficient mice. On the contrary to these proinflammatory actions in immune inflammation, EP<sub>4</sub> can exert anti-inflammatory action. Kabashima et al. (2002) found that EP<sub>4</sub>-deficient mice developed severe colitis in response to treatment with 3% dextran sodium sulfate, a dose that can be tolerated in wild-type animals. Again, this phenotype was mimicked by administration of an EP<sub>4</sub>-selective antagonist to wild-type mice. EP<sub>4</sub> deficiency was shown to result in impairment of mucosal barrier function that was associated with epithelial loss, crypt damage, and accumulation of neutrophils and CD4<sup>+</sup> T cells in the colon. DNA microarray analysis revealed increased expression of genes associated with immune responses and reduced expression of genes associated with mucosal repair and remodeling in the colon of EP<sub>4</sub>-deficient mice. Given the elevated level of PGE<sub>2</sub> and high expression of COX-2 in the brains of patients with Alzheimer's disease, Hoshino et al. (2007) examined the role of PGE<sub>2</sub> in processing β-amyloid precursor protein in cultured cells and found that PGE<sub>2</sub> stimulated production of amyloid-β peptide by activating γ-secretase and that this action occurs via EP<sub>2</sub> and EP<sub>4</sub>. They then cross-mated an Alzheimer's disease model of APP-23 transgenic mice with either EP<sub>2</sub>(-/-) mice and EP<sub>4</sub>(-/-) mice and found that the levels of Aβ peptides were significantly lower in the presence of either deletion. Besides these actions in complex inflammatory diseases, EP<sub>4</sub> is also implicated in a simple form of acute inflammation. Kabashima et al. (2007) subjected the ear of mice deficient in each of the EP subtypes to ultraviolet B irradiation and examined the extent of skin inflammation. They found that ear swelling caused by UV irradiation was significantly suppressed with reduced inflammatory cell infiltration and blood flow in EP<sub>2</sub>(-/-) and EP<sub>4</sub>(-/-)

mice compared with wild-type mice and that the effect of EP<sub>4</sub> deficiency was mimicked by administration of an EP<sub>4</sub> antagonist to wild-type mice. They further found that blockade of EP<sub>2</sub> and EP<sub>4</sub> is additive, suggesting that they function redundantly.

Epidemiological as well as experimental studies have implicated COX isoforms and PGs in familial adenomatous polyposis and development of colon cancer. To gain an insight into the receptor involved in this process, Watanabe et al., (1999) and Mutoh et al. (2002) examined azoxymethane-induced formation of aberrant cryptic foci in mice deficient in each prostanoid receptor. They found that the formation of such foci was suppressed in both EP1(−/−) and EP4(−/−) mice but not in those deficient in other receptor types or subtypes. In both instances, the number of foci was reduced to 50 to 60% of that apparent in wild-type mice.

Although systemically null EP<sub>4</sub>-deficient mice have contributed to our understanding of physiological functions of this receptor, their mixed genetic background has often limited their use. To conquer this weakness, Schneider et al. (2004) generated conditional EP<sub>4</sub><sup>fllox/fllox</sup> mice in which deletion of the EP<sub>4</sub> gene can be achieved by expression of Cre recombinase. Using endothelial cells derived from these mice and rendered EP<sub>4</sub>-null by transfection with adenovirus harboring Cre, Rao et al. (2007) found that EP<sub>4</sub> is required for PGE<sub>2</sub>-mediated migration, in vitro formation of capillary-like structure, cAMP production, and ERK activation. Combining these in vitro findings with in vivo findings that EP<sub>4</sub> agonists can induce angiogenesis in sponge implanted into mice, they argued proangiogenic potential for PGE<sub>2</sub>-EP<sub>4</sub> signaling. Gao et al. (2009) used this EP<sub>4</sub><sup>fllox/fllox</sup> mouse line and generated mice in which one allele of EP<sub>4</sub> was globally deleted and the other was targeted in osteoblasts. They found that this line of KO mice developed normal bone and exhibited no change in bone volumes or bone formation, whereas osteoblasts of these mice lost their responsiveness to PGE<sub>2</sub> in vitro. They argued that either EP<sub>4</sub> signaling may not be required for physiological regulation of bone development and maintenance or the loss of EP<sub>4</sub> may be compensated for by other mechanisms. Their findings are not compatible with the previous report by Li et al. (2005), who found osteopenia and impaired fracture healing in aged globally KO EP<sub>4</sub>-deficient mice and that by Akhter et al. (2001), who reported that EP<sub>4</sub>-deficient mice have small distal femur and vertebral bone volume and exhibit reduced structural and apparent material strength in the femoral shaft and vertebral body. Another study focusing on EP<sub>4</sub> in bone using conditional knockout mice examined the action of EP<sub>4</sub> in periaplastic osteolysis and the identity of the cell type mediating this action. Tsutsumi et al. (2009) implanted polyethylene beads to the periosteal surface of calvaria in EP1(−/−) mice, EP2(−/−) mice, and mice with conditionally deleted EP<sub>4</sub> in FSP1+ fibroblasts and examined osteolysis. They also prepared fi-

broblasts and osteoblasts from these mice, stimulated them with titanium beads or PGE<sub>2</sub>, and assessed production of RANKL. They found that polyethylene-bead-induced osteolysis is impaired only in conditional EP<sub>4</sub> KO mice. It is noteworthy that this conditional knockout exhibited reduced RANKL production only in fibroblasts and not in osteoblasts. On the basis of these findings, the authors suggested that osteolysis associated with total joint replacement is induced by RANKL produced by fibroblasts at the tissue-implant interface. Another example of the use of EP<sub>4</sub><sup>fllox/fllox</sup> mice is generation of mice with loss of EP<sub>4</sub> selectively in cardiomyocytes. Qian et al. (2008) generated this line of mice by crossing EP<sub>4</sub><sup>fllox/fllox</sup> with mice carrying Cre recombinase driven by  $\alpha$ -myosin heavy chain promoter. This line of cardiac-specific EP<sub>4</sub> KO mice does not show any abnormality but exhibits less hypertrophy and less fibrosis with attenuated STS3 activation in a model of myocardial infarction induced by left anterior descending coronary artery ligation. They do, however, show reduced ejection function. A later study by the same group (Harding et al., 2010) revealed that male mice of this line of conditional EP<sub>4</sub> knockouts spontaneously develop dilated cardiomyopathy at 23 to 33 weeks of age. Therefore, the cardiac-specific EP<sub>4</sub>-null mice exhibit a cardiac phenotype different from that of systemic EP<sub>4</sub>-null mice.

*d. Agonists and antagonists.* Selective EP<sub>4</sub> agonists typically contain a 16-phenyl group, the importance of which to vasodilator activity was recognized early (Johnson et al., 1980). Within the ONO series, a 3,7-dithia substitution pattern in the  $\alpha$ -chain was found to dramatically favor EP<sub>4</sub>/EP<sub>3</sub> selectivity (Maruyama et al., 2002a). Addition of large groups (e.g., *m*-phenyl) to the 16-phenyl ring resulted in retention of EP<sub>4</sub> binding affinity but reduction of functional potency, perhaps because of loss of efficacy (intrinsic activity) (Maruyama et al., 2002b). ONO-AE1-329, with a *m*-methoxymethyl substituent, emerged as a highly selective EP<sub>4</sub> full agonist (Cao et al., 2002). Prostanoids with a 8-aza-9-oxo functionality were originally identified as potent ligands for the rat kidney prostaglandin E receptor (Smith et al., 1977). In the Merck series of selective EP<sub>4</sub> agonists, this ring system is combined with a 16-phenyl group, and an acidic 5-tetrazole ring replaces the carboxylate, thereby preventing  $\beta$ -oxidation (Billot et al., 2003; Young et al., 2004); L-902688 (Fig. 4) seems to be the preferred molecule. CP-734432 (Fig. 4), the active metabolite of 5-(3-(2-(3-hydroxy-4-(3-(trifluoromethyl)phenyl)butyl)-5-oxopyrrolidin-1-yl)propyl)thiophene-2-carboxylate (PF-04475270), has an EC<sub>50</sub> value of 1 nM in a human rc-EP<sub>4</sub> assay (Prasanna et al., 2009). However, 8-aza-9-oxo prostanoids, with a 15,16-didehydro-15-methyl structure, switch from EP<sub>4</sub> to EP<sub>2</sub> agonist selectivity (Brugger et al., 2008). Finally, compound 12 (Fig. 4) in Blouin et al. (2010), which moves away from the classic prostanoid template, is an EP<sub>4</sub> full agonist.

The first EP<sub>4</sub> antagonist reported was 7-(5-(((1,1-biphenyl)-4-yl)methoxy)-2-(4-morpholinyl)-3-oxocyclo-

pentyl)-4-heptanoic acid (AH 23848) (Coleman et al., 1994a), and it played a pivotal role in the early pharmacological definition of the EP<sub>4</sub> receptor. AH 23848 was the prototype and has been overtaken by more selective and much more potent compounds, some of which contain a diaryl-acylsulfonamide as a key component of the scaffold: L-161982 (Fig. 5) bears structural resemblance to GW 627368 and BGC-20-1531 (Fig. 5) (Jones et al., 2009). Other EP<sub>4</sub> antagonists are carboxylic acids, such as MK-2894 (Blouin et al., 2010) (Fig. 5). Finally, ER-819762 (Fig. 5) is an EP<sub>4</sub> antagonist containing a novel *spiro* ring system (Chen et al., 2010).

*e. Therapeutics.* The most prominent utility for EP<sub>4</sub> antagonists is for treating inflammatory diseases, related hyperalgesia, and allodynia (Jones et al., 2009). In common with most prostanoid receptors, EP<sub>4</sub> receptors have been implicated in carcinogenesis, and EP<sub>4</sub> antagonists are shown to be effective in animal models (Table 8). It is noteworthy that Yanni et al., (2009) have presented data showing that EP<sub>4</sub> receptor antagonists may be useful for treating neovascular eye disease.

The potential uses of EP<sub>4</sub> agonist are many and diverse (Table 9). There is enormous diversity: hearing loss (Hori et al., 2009), nephritis (Nagamatsu et al., 2006) glaucoma (Woodward et al., 2009), and myocardial infarction (Xiao et al., 2004). This wide array of EP<sub>4</sub> agonist and antagonist activities portends a potential catalog of unwanted side effects, especially associated with oral systemic administration. Beneficial and deleterious side effects may occur in the same target tissue in response to EP<sub>4</sub> receptor agonist administration. Although EP<sub>4</sub> receptors are cardioprotective in ischemia-reperfusion injury (Xiao et al., 2004), they are likely to exacerbate hypertrophy that occurs as an adaptive response to cardiovascular disease (Mendez and LaPointe, 2005; Frias et al., 2007). EP<sub>4</sub> receptors stimulate the BNP promoter and could result in an antifibrotic action in the heart (Qian et al., 2006), which may be compensatory. It seems difficult to assess the pros and cons of treating a patient with heart failure with an EP<sub>4</sub> agonist. Added to all this, EP<sub>4</sub> agonists are likely to lower blood pressure in humans, because they are well known as potent vasodilators and may also contribute to atherosclerotic plaque destabilization (Cipollone et al., 2005).

Potential cardiovascular and other safety risks are likely to be avoided where local drug delivery is feasible. An EP<sub>4</sub> agonist may be useful in treating colitis (Nitta et

al., 2002; Jiang et al., 2007), and a compound could perhaps be designed so that systemic absorption is limited. Set against this, EP<sub>4</sub> receptors are implicated in colon carcinogenesis (Mutoh et al., 2002; Chell et al., 2006; Doherty et al., 2009), a highly undesirable side effect. In glaucoma therapy, where local topical drug administration is routinely employed, the highly efficacious ocular hypotensive effects produced by EP<sub>4</sub> agonists are accompanied by ocular surface hyperemia and corneal neovascularization (Aguirre et al., 2009; Prasanna et al., 2009; Woodward et al., 2009). Although some redness of the eyes is not harmful, the patients do not necessarily want it. Perhaps the most therapeutically and commercially successful utility may be for the recently reported utility of EP<sub>4</sub> agonists for sensorineural hearing loss (Hori et al., 2009). This would involve local therapy, probably an implant.

### C. FP Receptors

*1. Second Messenger Signaling.* The prostanoid FP receptor is predominantly G<sub>q</sub>-coupled, with activation of the classic pathway. Thus, after PLC $\beta$  activation, there is PI turnover, with resultant diacylglycerol-mediated PKC activation and a Ca<sup>2+</sup> transient signal in response to inositol trisphosphate formation (Nakao et al., 1993; Abramovitz et al., 1994; Ito et al., 1994; Sugimoto et al., 1994a; Woodward and Lawrence, 1994; Carrasco et al., 1996). Downstream of G<sub>q</sub>, other protein kinases are activated and receptors transregulated. Elevated [Ca<sup>2+</sup>]<sub>i</sub> results in calmodulin-mediated myosin light-chain kinase activation (Ansari et al., 2003). PKC activates the Raf/MEK/MAP kinase signaling pathway (Chen et al., 1998; Bos et al., 2004; Husain et al., 2005; Xu et al., 2008). In addition to activating MAP kinases, a PKC/Ca<sup>2+</sup>-calcineurin-nuclear factor of activated T cells pathway has been implicated in PGF<sub>2 $\alpha$</sub> -mediated cell growth (Horsley and Pavlath, 2003; Sales et al., 2009). FP receptors may also activate MAP kinases via PLC-mediated phosphorylation of the epidermal growth factor (EGF) receptor (Sales et al., 2004).

The repertoire of FP receptor G protein coupling extends beyond G<sub>q</sub>. It also includes activation of Rho via G<sub>12</sub>/G<sub>13</sub> (Pierce et al., 1999). The FP receptor is also reported to couple to G<sub>i</sub> (Melien et al., 1998; Hébert et al., 2005), providing an alternative route to the Raf/MEK/MAP kinase pathway (Bos et al., 2004). The early response gene *Cyr61* is also up-regulated by FP receptor stimulation (Liang et al., 2003) by a pathway sequen-

TABLE 8  
Potential therapeutic application of EP<sub>4</sub> antagonists (inflammation/pain is reviewed in Jones et al., 2009)

Agonist	Route	Dose	Species	Experimental Model	Indication	Reference
L 161982	Intravitreal injection	0.01–1 $\mu$ M	Rat	O <sub>2</sub> -retinopathy laser induced choroidal neovascularization	Age-related macular degeneration	Yanni et al., 2009
ONO-AE2-227	In diet	100–400 ppm	Mouse	Azoxymethane induced aberrant crypt foci; min mouse	Colon cancer	Mutoh et al., 2002
ONO-AE3-208	Oral	10 mg/kg	Mouse	Injection of M26 cells	Colon cancer	Yang et al., 2006



TABLE 9  
Potential therapeutic application of  $EP_4$  agonists

Agonist	Route	Dose	Species	Experimental Model	Indication	Reference
3,7-Dithia PGE <sub>1</sub> and its isopropyl ester 4819-CD	Topical	0.01–0.1%	Monkey	“Glaucomatous” monkey intraocular pressure	Glaucoma	Woodward et al., 2009
	Subcutaneous	0.3 mg/kg	Mouse	Ischemia-reperfusion injury	Myocardial infarction	Xiao et al., 2004
AEI-734	Subcutaneous	3 mg/kg	Mouse	Dextran sodium sulfate colitis	Colitis	Kabashima et al., 2002
AGN 205203 and its methyl ester					Colitis	Jiang et al., 2007
CP-734432	Subcutaneous	0.3–3 mg/kg	Rat	Ovariectomized rats	Osteoporosis	Ke et al., 2006
ONO-4819	Subcutaneous	10, 30 $\mu$ g/kg	Rat	Femoral drill-hole injury	Bone fracture healing, osteoporosis	Tanaka et al., 2004
	Subcutaneous	3, 30 $\mu$ g/kg	Rat	Mechanical loading, to tibia	Bone fracture, osteoporosis	Hagino et al., 2005
ONO-4819	Intravenous	100 ng/kg	Rat	Ovariectomized rat	Osteoporosis	Yoshida et al., 2002
ONO-AE1-329	Intracolonic	25, 100 $\mu$ g/kg	Rat	Dextran sodium sulfate colitis	Colitis	Nitta et al., 2002
	Intravenous	1–10 $\mu$ g/kg	Rat	Indomethacin-induced lesions	NSAID-induced gastrointestinal lesions	Kunikata et al., 2002
ONO-AE1-329	Intraperitoneal	0.1 mg/kg	Rat	Indomethacin-induced lesions	NSAID-induced gastrointestinal lesions	Hatazawa et al., 2006
ONO-AE1-329	Subcutaneous	0.1–1000 ng/g	Rat	Neonatal ductus arteriosus dilatation	Premature birth	Momma et al., 2005
ONO-AE1-329	Local gelatin plug	1 mg/ml	Guinea pig	Auditory brain stem response	Sensorineural hearing loss	Hori et al., 2009
ONO-AE1-329	Subcutaneous	0.05, 0.5 mg/kg 55 $\mu$ g daily infusion per animal	Mouse	N2 receptor knockout mice	Nephrogenic diabetes insipidus	Li et al., 2009
ONO-AE1-329	Subcutaneous	100 $\mu$ g/kg	Mouse	Anti-glomerular basement membrane antibody nephritis	Glomerulonephritis	Nagamatsu et al., 2006
ONO-AE1-329	Intravenous	1–10 $\mu$ g/kg	Rat	Endotoxin	Septicemia	Sakamoto et al., 2004
PF-04475270	Topical	0.002–0.1 mg/ml	Dog	Normal dog intraocular pressure	Glaucoma	Prasanna et al., 2009

tially involving Ras/Raf signaling and Tcf transcription independent of MEK/ERK (Xu et al., 2009).

**2. Distribution and Biological Functions.** FP receptors have a wide distribution and subserve many important functions. The importance of these functions is reflected by the fact that, among the prostanoid receptors, the FP receptor has been the most successful therapeutic target. It has prominent functions in reproduction. The FP receptor is highly expressed in ovarian tissue, with mRNA expressed exclusively in the corpus luteum (Sugimoto et al., 1994a). Although the FP receptor is central to corporal luteal regression and regulating the estrous cycle in farm animals (Coleman et al., 1994b), this does not seem to be the case in mice, because mice lacking FP receptors exhibited an unchanged estrous cycle and were fertile (Sugimoto et al., 1997). FP receptors are also present in the human corpus luteum (Narko et al., 1997; Väänänen et al., 1998), but their function is uncertain and certainly not dramatic. In mice lacking the FP receptor, parturition was abolished (Sugimoto et al., 1997). This has stimulated interest in the use of the FP antagonists for preventing preterm labor.

Functional expression of FP receptor in the myometrium mediating contraction has long been known (Senior et al., 1992; Carrasco et al., 1996). FP receptors are also expressed in the human endometrium, where PGF<sub>2 $\alpha$</sub>  is biosynthesized in endometrial epithelial cells and causes cell proliferation (Asselin et al., 1997; Milne and Jabbour, 2003). FP receptor expression has been

reported in human deciduae, and this has been suggested to contribute to parturition (Makino et al., 2007). By far the most prominent implication of FP receptors in disease is in uterine cancer, specifically endometrial adenocarcinomas. The role of the FP receptors in the progression of endometrial adenocarcinoma includes potentiation of angiogenesis by EGF receptor transactivation and induction of VEGF mRNA expression (Sales et al., 2005), alteration of adhesion, morphology, and migration (Sales et al., 2008, 2009).

In the CNS, PGF<sub>2 $\alpha$</sub>  given intracisternally alleviates kainic acid-induced seizures potentiated by COX-2 inhibitors, suggesting that PGF<sub>2 $\alpha$</sub>  behaves as an endogenous anticonvulsant (Kim et al., 2008). In contrast, FP receptors are claimed to significantly contribute to brain damage associated with focal brain ischemia (Saleem et al., 2009a). The colocalization of the FP receptor and PGF synthase 1 in the spinal cord suggest a role in pain, notably because of intense FP immunostaining in spinal laminae I and II of the dorsal horn (Suzuki-Yamamoto et al., 2009). Indeed, FP receptors mediate  $\alpha$ B-methylene ATP-evoked allodynia (Kunori et al., 2009). These data are supported by the finding that spinal intrathecal administration of PGF<sub>2 $\alpha$</sub>  produces allodynia by activating FP receptors (Muratani et al., 2003).

Renal expression of FP receptors is high (Sugimoto et al., 1994a; Saito et al., 2003). It is most abundant in the distal convoluted tubule and aquaporin-2-positive cortical collecting ducts (Saito et al., 2003; Hébert et al.,

2005). This distribution is consistent with known FP-mediated effects on water and solute transport in these segments of the kidney. Renal FP receptors seem to signal via a pertussin toxin-sensitive mechanism (Hébert et al., 2005).

The FP receptor is widely distributed in the human eye (Schlötzer-Schrehardt et al., 2002). Despite high FP receptor expression in the corneal epithelium, ciliary epithelium, and iridial stroma and smooth muscle (Schlötzer-Schrehardt et al., 2002), FP agonists are functionally important only in those cells involved in aqueous humor outflow. Thus, ciliary smooth muscle and trabecular meshwork cells are targeted by FP-based therapeutics, without a litany of unwanted side effects. The success of FP agonists, in the form of ester prodrugs, as first-line antiglaucoma agents originated from the vision and determination of Bito (2001). The mechanism by which FP agonists lower intraocular pressure is now entirely understood. FP agonists increase aqueous humor outflow from the anterior segment of the eye, predominantly through the uveoscleral pathway (Stjernschantz et al., 2001; Toris et al., 2005). Effects on trabecular outflow have also been noted in human subjects (Ziai et al., 1993; Toris et al., 2007) and anterior segment preparations (Bahler et al., 2008). Uveoscleral outflow occurs via a widening of the interstitial spacing between ciliary muscle fiber bundles and a controlled remodeling that creates organized drainage channels (Richter et al., 2003). Consistent with a controlled remodeling process, tissue inhibitor of metalloproteinases regulation (Anthony et al., 2002), altered matrix metalloproteinase transcription, and translation products occur (Ocklind, 1998; Sagara et al., 1999; Gaton et al., 2001; Weinreb and Lindsey, 2002). Morphological changes are not restricted to the anterior portion of the ciliary body; they also occur in the trabecular meshwork (Richter et al., 2003). It is not surprising, therefore, that tissue inhibitor of metalloproteinases and matrix metalloproteinase regulation occurs in trabecular meshwork cells (Oh et al., 2006). Gene regulation associated with FP receptor stimulation, however, is not identical in ciliary muscle and trabecular meshwork with respect to genes that would influence aqueous humor dynamics (Zhao et al., 2003). More specifically related to tissue remodeling, Cyr61, connective tissue growth factor, epidermal growth factor receptor-1, and HIF-1 $\alpha$  are up-regulated by FP receptor stimulation in ciliary muscle cells (Liang et al., 2003; Hutchinson et al., 2010). The full repertoire of genes that regulate ciliary body remodeling and increased uveoscleral outflow remains to be elucidated.

The side effects associated with FP receptor antiglaucoma therapy are also well understood. The major side effect of latanoprost is iridial hyperpigmentation. This results from a benign eumelanogenic effect, with larger and more mature melanosomes present in iridial melanocytes (Stjernschantz, 2001). Mechanistic investigation

demonstrated that latanoprost does not directly stimulate melanocytes but rather activates FP receptors on neighboring fibroblasts (Smith-Thomas et al., 2004). This is quite different from dermal melanocytes, where FP receptor activation directly produces dendricity and increases tyrosinase activity (Scott et al., 2005). FP receptor-induced ocular surface hyperemia involves NO-mediated, endothelium-dependent vasodilation (Chen et al., 1995). Latanoprost also produces hypertrichosis of the eyelashes (Johnstone and Albert, 2002).

FP receptor stimulation has long been known to stimulate fibroblast proliferation. FP receptor activation in normal rat kidney fibroblasts adds a new dimension. Here FP receptor stimulation caused membrane depolarization of transformed normal rat kidney cells (Almirza et al., 2008). A positive feedback loop involving FP receptors and COX-2 up-regulation was proposed (Almirza et al., 2008). A most interesting recent report shows that FP receptors facilitate bleomycin-induced pulmonary fibrosis (Oga et al., 2009). FP, via a Rho kinase signaling pathway, produces fibrosis independent of TGF $\beta$ , which, until now, has been considered the dominant profibrotic mediator.

Rat ventricular cardiomyocytes undergo hypertrophic growth in response to FP receptor stimulation (Adams et al., 1996; Lai et al., 1996; Pönicke et al., 2000). PGF<sub>2 $\alpha$</sub>  has also been claimed to mediate inflammatory tachycardia (Takayama et al., 2005). Studies on the contractility of the intact heart have revealed that FP receptors produce a negative inotropic effect, which could contribute to cardiac dysfunction (Jovanović et al., 2006). PGF<sub>2 $\alpha$</sub>  seems to have pathophysiological consequences for the cardiovascular system, because it also elevates blood pressure and promotes atherosclerosis (Yu et al., 2009b). PGF<sub>2 $\alpha$</sub>  also increases skeletal muscle cell growth (Horsley and Pavlath, 2003).

Most prostanoid receptors have been implicated in cancer, and FP is no exception. PGF<sub>2 $\alpha$</sub>  stimulates the motility and invasiveness of colorectal tumor cells, with potency equal to that of PGE<sub>2</sub> (Qualtrough et al., 2007). PGF<sub>2 $\alpha$</sub> -mediated COX-2 up-regulation has been suggested to potentiate tumorigenesis with respect to endometrial adenocarcinoma (Jabbour et al., 2005). Numerous early studies on PGF<sub>2 $\alpha$</sub>  have shown a mitogenic effect on fibroblast cell lines, and this was often suggested as contributory to tumor growth (De Asua et al., 1975).

Studies on murine osteoclast development suggest that PGF<sub>2 $\alpha$</sub>  would inhibit bone resorption (Kamon et al., 2008). FP receptor expression has been detected in primary human osteoblasts (Sarrazin et al., 2001), and in UMR-106 cells, functional FP receptors were identified (Yamaguchi et al., 1988). Finally, FP receptor activation potently inhibits adipose cell differentiation according to studies on adipocyte precursors obtained from murine inguinal fat pads (Serrero and Lepak, 1997).

**3. Gene Deletion Studies.** Sugimoto et al. (1997) generated a line of FP-deficient mice by replacing the second exon of the FP gene with  $\beta$ -galactosidase- and neomycin-resistance genes, and this line has been used in all studies below.  $\text{PGF}_{2\alpha}$  is known as a luteolytic substance in animals. Initial analysis of this line of mice revealed that FP(–/–) female mice did not show any abnormality in the luteal cycle but exhibited parturition failure (Sugimoto et al., 1997). The failure was apparently due to the lack of labor, and these mice did not exhibit prepartum decline in the plasma progesterone level. Ovariectomy 24 h before the expected term decreased the progesterone level and induced normal parturition in FP(–/–) dams. Given the high expression of FP in the corpora lutea that produces progesterone and the luteolytic action of  $\text{PGF}_{2\alpha}$ , these findings suggest that the  $\text{PGF}_{2\alpha}$ -FP pathway triggers parturition by inducing luteolysis in the ovary.

Several PGF analogs are used as antiglaucoma drugs. FP(–/–) mice were used to examine the identity of the receptor mediating IOP-lowering activity of these PG analogs. These mice were resistant not only to FP agonists such as latanoprost and travoprost but also to bimatoprost and unoprostone, PG analogs suggested not to bind to FP with high affinity, although these compounds lowered IOP to a comparable extent in wild-type C57BL/6 mice (Crowston et al., 2004, 2005; Ota et al., 2005). These results indicate that FP mediates IOP-lowering action of all of these PG analogs at least in mice. Whereas FP mediates potent reduction of IOP in response to exogenously applied FP agonists, FP(–/–) mice showed no abnormality in diurnal variation of IOP compared with wild-type mice, indicating that the  $\text{PGF}_{2\alpha}$ -FP pathway is not involved in physiological regulation of IOP (Crowston et al., 2007). Furthermore, administration of ONO-AE1-259 (Fig. 4), an  $\text{EP}_2$  agonist, and ONO-AE1-329 (Fig. 4), an  $\text{EP}_4$  agonist, induced reduction of IOP equally well in wild-type and FP(–/–) mice, indicating the presence of a PG-dependent pathway different from that of FP in lowering IOP (Saeki et al., 2009).

Systemic inflammation induces many adaptive symptoms, one being tachycardia. Takayama et al. (2005) used mice deficient in individual prostanoid receptors and examined their involvement in this inflammatory tachycardia. They first found that  $\text{PGF}_{2\alpha}$  and a TP agonist, I-BOP, could elevate the heart rate of mice through FP and TP, respectively, and that this action was exerted locally in the atria. They then showed that tachycardia induced by systemic administration of lipopolysaccharide was abrogated partially in either TP(–/–) or FP(–/–) mice and completely in mice deficient in both FP and TP receptors. Inflammatory tachycardia was believed to result from increased sympathetic discharge, and the results by Takayama et al. (2005) have changed this traditional view. The function of  $\text{PGF}_{2\alpha}$  in the cardiovascular system was further elu-

cidated by the use of FP(–/–) mice (Yu et al., 2009b). Systolic blood pressure in FP(–/–) mice was significantly lower than that of wild-type mice, with significantly lower plasma concentrations of renin and angiotensin-1 in FP(–/–) mice. To explore the basis of this phenotype, they examined FP receptor localization in the kidney and found that FP is expressed in the preglomerular artery (Yu et al., 2009b), in addition to previously identified localization in distal tubules and collecting ducts (Saito et al., 2003); stimulation of FP increased renin mRNA expression in isolated JG cells, though it did not stimulate renin release. In addition, FP is expressed in the medial smooth muscle layer of resistant arterioles, and infusion of  $\text{PGF}_{2\alpha}$  elevated blood pressure in wild-type but not in FP(–/–) mice. Yu et al. (2009b) further cross-mated FP(–/–) with mice deficient in the LDL receptor, and reported that atherosclerosis was significantly attenuated on the FP(–/–) background. Although macrophage infiltration and inflammatory cytokine expression in atherosclerotic plaques were diminished in FP(–/–) mice, it remained unclear how FP is involved in atherogenesis. Saleem et al. (2009a) subjected FP(–/–) mice to transient brain ischemia-reperfusion by reversible ligation of the middle cerebral artery. They found that the loss of FP does not alter physiological parameters such as cerebral blood flow,  $\text{PaO}_2$ ,  $\text{PaCO}_2$ , mean arterial blood pressure, and body temperature before, during, or after brain ischemia. Nonetheless, FP(–/–) mice exhibited significantly less neurological deficit and smaller infarct volume than wild-type mice. To examine whether this difference is due to excitoneurotoxicity associated with transient ischemia-reperfusion, they then injected NMDA into the striatum unilaterally and found that FP(–/–) mice exhibited significantly smaller infarct volumes than wild-type mice again under these conditions, indicating that FP somehow regulates the excitation of glutamatergic neurons after ischemic injury. Consistent with these findings, they showed that administration of latanoprost widened the infarction and worsened the deficit. Finally, Oga et al. (2009) subjected mice deficient in individual prostanoid receptor to bleomycin-induced pulmonary fibrosis and found significant suppression of fibrosis in FP(–/–) mice. They found that pulmonary inflammation comparable with that in wild-type mice was induced by bleomycin, but subsequent fibrosis was suppressed in FP(–/–) mice. Microarray analysis revealed significant attenuation of the induction of genes associated with fibrosis, such as various isoforms of collagen and other matrix proteins in FP(–/–) mouse lung, and FP stimulation in cultured lung fibroblasts facilitated cell proliferation and collagen production in vitro. It is noteworthy that the loss of FP receptors did not affect expression and activation of TGF- $\beta$ , a critical cytokine in fibrosis, and  $\text{PGF}_{2\alpha}$  and TGF- $\beta$  contributed additively to fibrosis. On the basis of these results, the authors suggested that  $\text{PGF}_{2\alpha}$  functions as a mediator of fibrosis independent of



TGF- $\beta$ , and FP may be a drug target for idiopathic pulmonary fibrosis in humans.

**4. Agonists and Antagonists.** Early studies established the high FP selectivity of the (racemic) 16-*m*-trifluoromethyl-phenoxy analog of PGF<sub>2 $\alpha$</sub>  (ICI-81008, fluprostenol; Dukes et al., 1974), particularly its minimal TP agonism (Welburn and Jones, 1978). Fluprostenol continues to be the FP agonist of choice and is commercially available as the more active (+)-enantiomer. The 16-*m*-chlorophenoxy analog of PGF<sub>2 $\alpha$</sub>  (cloprostenol) is the most potent FP agonist reported but retains considerable EP<sub>3</sub> agonism; change to a 3-hydroxy-tetrahydrofuran ring with shift of the 5,6-*cis* bond to the 4 position as in AL-12180 (Fig. 6) improved the selectivity ratio (Sharif et al., 2006). 13,14-Dihydro-17-phenyl PGF<sub>2 $\alpha$</sub>  (latanoprost-free acid) also has high FP selectivity, showing 22-fold less EP<sub>1</sub> agonism than 17-phenyl PGF<sub>2 $\alpha$</sub>  (Ungrin et al., 2001). Introduction of a 2-indanyl group into the  $\omega$ -chain has resulted in claims for FP antagonism (Griffin et al., 1999; Sharif et al., 2000; AL-3138, AL-8810), although partial agonism seems to be a better description (Woodward et al., 2007). Structures are given in Fig. 6. Woodward et al. (2000) reported that although the C1-alcohol and C1-methyl ether analogs of PGF<sub>2 $\alpha$</sub>  were very weak agonists at FP-receptors of cat and human, they showed high potency on the cat lung parenchyma preparation, an action that could not be ascribed to other known prostanoid receptors. Subsequent studies have suggested the existence of a new receptor type, the prostamide F receptor, which recognizes carboxyl, alcohol, ethanolamide, and alkylamide moieties at C1, PGF<sub>2 $\alpha$</sub> -ethanolamide (Fig. 6) being a potential natural ligand (Woodward et al., 2008). In cat iris sphincter digests, 17-phenyl PGF<sub>2 $\alpha$</sub> -ethylamide (bimatoprost) activated one set of cells and 17-phenyl PGF<sub>2 $\alpha$</sub>  and PGF<sub>2 $\alpha$</sub> , another (Spada et al., 2005).

C-1 amine and amide derivatives of PGF<sub>2 $\alpha$</sub>  show weak FP antagonism in some systems, but their utility is

severely limited (see Jones et al., 2009). 15-Indanyl- $\omega$ -pentanor PGF analogs, such as AL-3138 and AL-8810 (Fig. 7), represent improvements (Griffin et al., 1999; Sharif et al., 2000). However, AL-8810 has modest affinity for FP receptors (IC<sub>50</sub>, 8.7  $\mu$ M in human ciliary muscle cells; Sharif et al., 2006) and blocks TP receptors in mouse uterus (Hutchinson et al., 2003), pig ciliary artery (Vysniauskiene et al., 2006), and human recombinant receptor assays (Krauss and Woodward, unpublished). Partial block of epidermal growth factor-induced contraction of guinea pig trachea by AL-8810 (Schaafsma et al., 2005) may be explained by TP antagonism rather than the proposed FP antagonism. Moreover, both PGF analogs often exhibit FP partial agonism (Griffin et al., 1999; Hutchinson et al., 2003), and AL-8810 was even a full agonist in the cat iris preparation, an action not blocked by a prostamide F antagonist (Woodward et al., 2007). More potent, selective prostamide F antagonists, such as AGN 211334 (Fig. 7) have more recently been reported (Wan et al., 2007; Woodward et al., 2008).

Peptides of the THG series (Chemtob and Peri, 2006; Peri et al., 2009) block PGF<sub>2 $\alpha$</sub> -induced responses, but this may not necessarily involve competition for the FP receptor. The octapeptide THG-113.31 [Ile-Leu-Gly-His-(D-Cit)-Asp-Tyr-Lys] insurmountably blocked PGF<sub>2 $\alpha$</sub> -induced contraction of pig retinal blood vessels, while having minimal effect on contraction to 17-phenyl PGE<sub>2</sub>, U-46619, phenylephrine, Angiotensin II and endothelin-1. However, the affinity of TGH-131.31 for the human recombinant FP receptor was poor ( $\sim$ 13% at 10  $\mu$ M). Moreover, TGH-131.31 (10  $\mu$ M) weakly antagonized contraction of sheep myometrium induced by PGF<sub>2 $\alpha$</sub>  and had no effect on PGF<sub>2 $\alpha$</sub> -induced contraction of human pregnant myometrium but inhibited spontaneous and oxytocin-induced contractions at much lower concentrations (Friel et al., 2005). At 10 to 50  $\mu$ M, TGH-113.31 enhanced BK<sub>Ca</sub> channel opening in human uter-

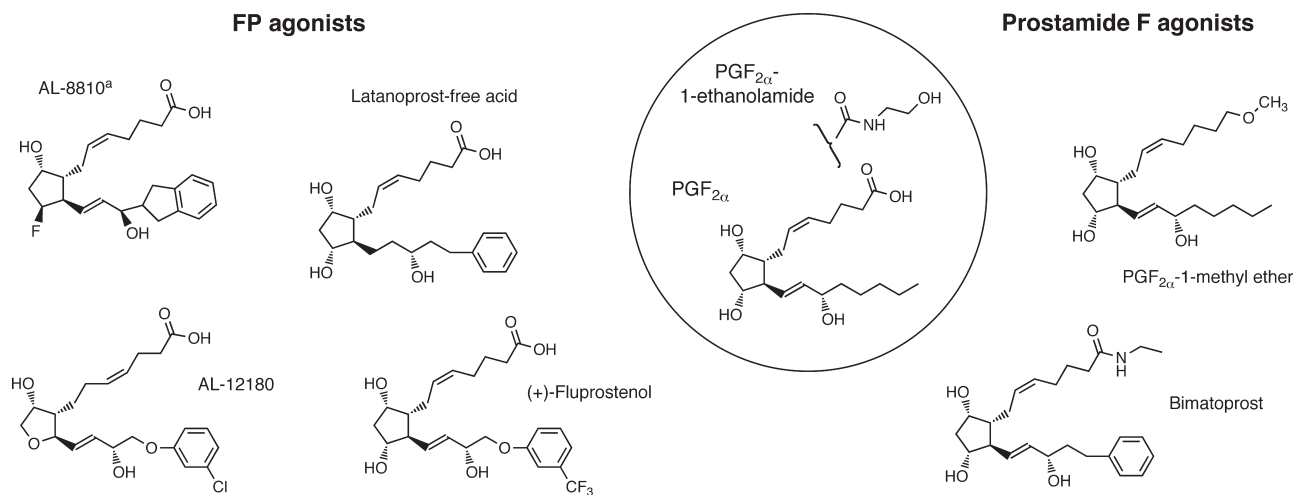


FIG. 6. Structures of agonists for the prostanoid FP receptor and the corresponding prostamide F receptor. PGF<sub>2 $\alpha$</sub>  potently activates both receptors, whereas its C1-ethanolamide activates only the prostamide F receptor. <sup>a</sup>, partial agonist.

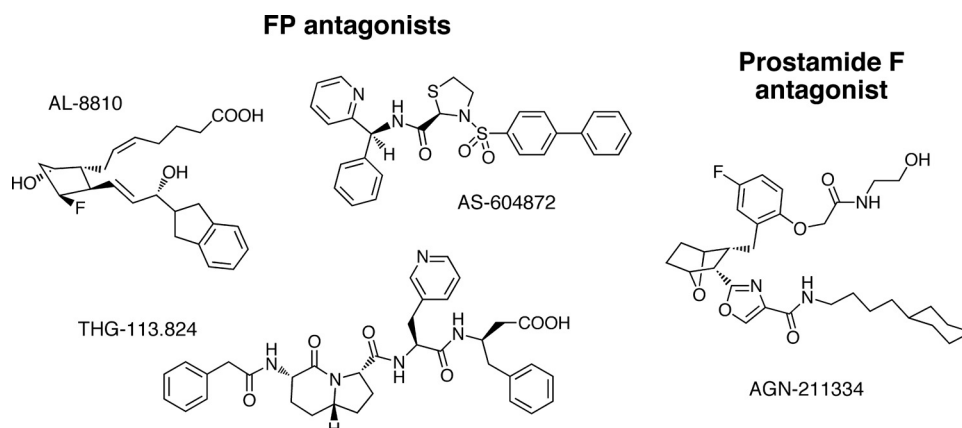


FIG. 7. Structures of FP antagonists and the prostamide antagonist (AGN 211334).

ine myocytes, which was reversed by iberiotoxin (Doheny et al., 2007). Simpler peptides (e.g., TGH-113.824; Fig. 7) also block  $\text{PGF}_{2\alpha}$ -induced contraction (Peri et al., 2009).

**5. Therapeutics.** By far the greatest therapeutic success of prostanoid pharmacology is the use of FP receptor agonist prodrugs for the treatment of glaucoma. Latanoprost was the first in its therapeutic class (Bito, 2001; Stjernschantz, 2001). This was followed by the isopropyl ester of fluprostenol, named travoprost (Hellberg et al., 2001) and tafluprost (Takagi et al., 2004; Cirillo et al., 2007). Comparisons between these drugs are widely available and are not reviewed here.

An alternative therapeutic use for FP agonists emerged from studies on glaucoma: hair growth. Latanoprost and related compounds cause eyelash growth (Johnstone and Albert, 2002). Studies in the stump-tailed macaque model of androgen-induced scalp alopecia showed latanoprost to be effective (Uno et al., 2002). This opportunity does not seem to have been commercially realized. The reasons for this are obscure. For this use, the domain of pharmacocosmetics (Woodward et al., 2008) is accessed. In the domain of cosmetics, unlike treating pain, inflammation, cancer, etc., the options are not restricted to drug intervention and surgery. Cosmetic aspirations can be instantly gratified by colorants, surgery, or implantation/replacement. This may have discouraged development of latanoprost for male pattern baldness. However, acceptance of cosmetic intervention and instant results is widely accepted by women but not by men. A gradual improvement in natural scalp hair is likely to be more socially acceptable to men because it would avoid derision from friends and colleagues. Regrowth of natural scalp hair would take a long time, because anaphase extends over years. In addition, like any other disease, early intervention is essential. These may have been major discouraging factors. On the other hand, the psychological impact of male pattern baldness cannot be underestimated, and balding men are typically highly motivated and inclined toward subtle and natural intervention. FP receptor expression has been

detected in the human dermal papillae (Colombe et al., 2008), which supports a favorable outcome for human clinical investigation. Fat reduction/antiobesity is a further potential pharmacocosmetic/medical use of FP agonists, but this is still at the adipocyte precursor stage essentially (Serrero and Lepak, 1997).

With the exception of osteoporosis, the effects of FP receptor activation are pathophysiological, and antagonist therapy would be the requirement. The major focus has been the prevention of preterm labor. These antagonists include the peptide THG-113.31 and derivatives (Olson, 2005; Chemtob and Peri, 2006; Jones et al., 2009). The non-PG structure FP antagonist AS-604872 (Fig. 7) (Cirillo et al., 2007) seems to have a better commercial prospect as an orally bioavailable small molecule. It showed no agonism in a human FP receptor-inositol phosphate assay ( $\text{IC}_{50}$ , 47 nM for  $\text{PGF}_{2\alpha}$ ). In vivo, AS-604872 (1–30 mg/kg i.v.) inhibited  $\text{PGF}_{2\alpha}$ -induced uterine contraction in the nonpregnant rat: inhibition of oxytocin-induced contractions was slight (Chollet et al., 2007). These apart, no serious attention seems to have been given to FP antagonist design. This may change in light of reports implicating FP receptors in cardiovascular disease (Yu et al., 2009b) and fibrosis (Oga et al., 2009).

#### D. IP Receptors

**1. Second Messenger Signaling.** The IP receptor has long been known to be both  $\text{G}_s$ - and  $\text{G}_q$ -coupled, resulting in increased cAMP formation, PI turnover, and  $\text{Ca}^{2+}$  signaling (Coleman et al., 1994; Namba et al., 1994; Narumiya et al., 1999). IP receptor activation of the cAMP-PKA pathway seems to be exclusively responsible for some prostacyclin-mediated events (Nasrallah et al., 2001; Ritchie et al., 2004; Kamio et al., 2007; Muja et al., 2007). Solution structure studies indicate that the first and third intracellular loops of the IP receptor are in contact with C-terminal residues of  $\text{G}\alpha_s$  to initiate cAMP signaling (Zhang et al., 2006a; Zhang et al., 2006b). N-glycosylation is involved in both adenylyl cyclase and inositol phosphate formation (Zhang et al., 2001).

IP receptors are not unique in activating dual or multiple signaling but seem to be among the best studied.  $G_i$  and  $G_q$  coupling to IP receptors has been reported to be dependent on  $G_s$  coupling as a prerequisite, the key event being phosphorylation of Ser<sup>357</sup> of the IP receptor by PKA (Lawler et al., 2001). This mechanism is consistent with investigations invoking both cAMP and cAMP-independent mechanisms in many instances. These include MaxiK channel activation (Yamaki et al., 2001) and STAT3 phosphorylation (Lo et al., 2008), in addition to straightforward analysis of dose-dependent effects on cAMP formation (Accomazzo et al., 2002). PKA-mediated switching from  $G_s$  to  $G_q$  and/or  $G_i$  coupling, however, does not seem to be a universal phenomenon (Chow et al., 2003). G-protein coupling seems to be cell-specific. No evidence for  $G_i$  coupling could be found in NG108-15 and SK-N-SH cells and in Chinese hamster ovary or human embryonic kidney 293 cells expressing recombinant IP receptors (Chow et al., 2003). In marked contrast,  $G_i$ -coupled IP receptors were found exclusively in human erythroleukemia HEL cells with respect to STAT1 and STAT3 phosphorylation (Lo et al., 2006) and rat medullary thick ascending limb cells (Hébert et al., 1998). Finally, but beyond the intended scope of this review, there are reports claiming that prostacyclin interacts with peroxisome proliferator-activated receptors to exert a portion of its activities (Lim and Dey, 2002; Ali et al., 2006).

**2. Distribution and Biological Functions.** The role of prostacyclin in cardiovascular homeostasis is well publicized. Thus, COX-2 inhibitors may present a cardiovascular hazard by depressing prostacyclin production by vascular endothelial cells without a concomitant reduction in platelet COX-1-derived TxA<sub>2</sub>. This results in a potentially deleterious effect on thrombosis and blood pressure and accelerated atherogenesis (Cheng et al., 2002; Dogné et al., 2005; Wang et al., 2005). Of particular relevance is that COX-2 expression and PGI<sub>2</sub> production in human coronary arterial endothelial cells may be increased by proinflammatory stimuli (Tan et al., 2007). These findings imply that local PGI<sub>2</sub> produced in the coronary vasculature may directly counteract the vasoconstriction and platelet aggregation production by TxA<sub>2</sub> during episodic vascular insult. The cardioprotective action of prostacyclin has been proposed to extend to preventing atherosclerosis, intimal hyperplasia, and restenosis (Fetalvero et al., 2007).

Vascular smooth muscle cells are not terminally differentiated and can proliferate and further differentiate. IP receptors inhibit vascular smooth muscle cell proliferation (Lin et al., 2008) by inhibiting  $G_i$ -to-S-phase progression (Fetalvero et al., 2007) and inducing apoptosis (Li et al., 2004). Prostacyclin also induces genes characteristic of a contractile phenotype and inhibits vascular smooth muscle cell migration (Blindt et al., 2002; Fetalvero et al., 2007). Vascular endothelial cells are, in addition to vascular smooth muscle cells, an

important source of prostacyclin, which is induced by laminar shear stress (Di Francesco et al., 2009). Beyond prostacyclin effects on vascular smooth muscle and endothelial cells, the inhibitory effects on platelet aggregation and its vasodilator properties should not be understated. IP receptor stimulation increases retinal, choroidal (Mori et al., 2007), and coronary blood flow (Gwózdź et al., 2007). In ApoE-deficient mice, IP receptors conferred protection against the initiation and progression of atherogenesis by limiting platelet activation and leukocyte-vascular endothelial cell interaction (Kobayashi et al., 2004). Mice lacking IP receptors showed augmented cardiac hypertrophy in response to pressure overload (Hara et al., 2005) and increased myocardial infarct size after ischemia-reperfusion injury (Xiao et al., 2001), suggesting a cytoprotective effect on cardiomyocytes independent of platelet and neutrophil inhibition. IP (−/−) mice also develop salt-sensitive hypertension and cardiac fibrosis, in addition to cardiac hypertrophy (Francois et al., 2005).

Continuing the subject of systemic hypertension, decreased susceptibility to renovascular hypertension was reported in mice lacking IP receptors (Fujino et al., 2004). Prostacyclin, therefore, indirectly produces hypertension in the two-kidney, one-clip hypertension model: the mechanism seems to involve increased renin mRNA expression and production according to studies on cicaprost in juxtaglomerular cells (Fujino et al., 2004). In addition to juxtaglomerular cells, IP message is also present in the renal cortex, outer and inner medulla, and inner medullary collecting duct, according to reverse transcription-polymerase chain reaction (Nasrallah et al., 2001). Localization studies using in situ hybridization detected IP receptor mRNA in the tubules of the inner and outer medulla, the vasculature, and in the arteries, glomeruli, and tubules of the cortex (Nasrallah et al., 2001). Renal IP receptor distribution in murine species does not necessarily correlate with that in humans (Nasrallah and Hébert, 2005). No obvious pathological renal condition is apparent in IP (−/−) mice. Prostacyclin, however, may also be involved in renal hypertrophy, fibrosis, and apoptosis (Nasrallah and Hébert, 2005).

The prostanoid IP receptor plays a prominent role in edema formation, hyperalgesia, and pain (Bley et al., 1998; Hata and Breyer, 2004). Nociceptive responses in the acetic acid writhing model are virtually abolished in mice lacking IP receptors (Murata et al., 1997). In models of rheumatoid arthritis, such as carrageenin-induced paw edema and collagen antibody-induced arthritis (Murata et al., 1997; Ueno et al., 2000; Honda et al., 2006; Pulichino et al., 2006), a pronounced reduction is observed in IP (−/−) mice. Likewise, pleurisy induced by carrageenin (Yuhki et al., 2004) and zymosan (Yuhki et al., 2008) is attenuated in mice lacking IP receptors. Results obtained with IP antagonists in rat models of pain and inflammation (Bley et al., 2006; Pulichino et



al., 2006) are consistent with results obtained in the IP(−/−) mice studies. The involvement of IP receptors in pain and hyperalgesia are supported by IP expression studies (Bley et al., 1998; Doi et al., 2002) and functional studies on neurons conducting at C velocity (Smith et al., 1998). Studies on dorsal root ganglia support an IP receptor role in sensitization of sensory neurons (Smith et al., 1998; Rowlands et al., 2001; Nakae et al., 2005).

IP receptor expression in the thymus and spleen was reported more than a decade ago (Narumiya et al., 1999). Since then, some insight into IP receptor involvement in lymphocyte function has emerged. IP receptors tend to suppress Th2-mediated responses (Hata and Breyer, 2004). Thus, IP receptor stimulation inhibits Th2 cytokine (IL-4, IL-10, IL-13) and Th1 cytokine (IFN $\gamma$ ) production (Zhou et al., 2007). Plasmacytoid dendritic cells, which are believed critical for controlling adaptive immunity, also express functional IP receptors, which suppressed Toll-like receptor-mediated TNF $\alpha$  and INF $\gamma$  production and enhanced IL-10 production (Hung et al., 2009). IP signaling has been claimed to prevent recruitment of Th2 cells into airways in an asthma model (Jaffar et al., 2007), but the use of highly unstable PGI $_2$  confounds interpretation. Nevertheless, because allergic airway and cutaneous inflammation involving Th2 cells in the same species is augmented in IP receptor-deficient mice (Takahashi et al., 2002), it seems correct that IP receptors inhibit Th2 cell function. Further support is provided by a study that implicates prostacyclin in the anti-inflammatory activity of 1-methylnicotinamide in experimental contact hypersensitivity (Brynarski et al., 2008). Finally, in the macrophage cell line Raw 264.7, IP activation via LPS-induced prostacyclin production generates VEGF (Park et al., 2007). Because this pathway involved Akt signaling, it is relevant to angiogenesis and cancer. IP receptors up-regulate angiogenic genes in the human endometrium (Smith et al., 2006).

IP receptors have been functionally characterized in the human myometrium (Senior et al., 1992). Prostacyclin synthase and IP receptor expression are increased during the menstrual phase (Battersby et al., 2004), indicating a possible role in normal and/or dysfunctional menstruation. Perhaps a better case could be made for PGI $_2$  participation in pregnancy and labor by up-regulating contractile proteins and connexin 43 that could prime the myometrium for parturition (Fetalvero et al., 2008; Taggart et al., 2008). IP receptors have also been implicated in preimplantation embryo development (Huang et al., 2007a) and may contribute to embryo transport by decreasing the amplitude of oviductal contractility (Arbab et al., 2002).

Prostacyclin receptor deletion has been shown to aggravate hippocampal neuronal loss caused by ischemia (Wei et al., 2008) and cortical cell loss after traumatic brain injury (Lundblad et al., 2008). These experiments indirectly suggest that IP receptors could promote neu-

ronal survival. The ulcerogenic response to ischemia-reperfusion was increased in severity in IP(−/−) mice, suggesting that IP receptors play a crucial role in gastric mucosal defense in vascular injury (Kotani et al., 2006). IP involvement in adaptive cytoprotection produced by mild irritants seems controversial (Boku et al., 2001; Takeuchi et al., 2001b).

**3. Gene Deletion Studies.** Two lines of IP-deficient mice were generated independently (Murata et al., 1997; Cheng et al., 2002). A study on IP(−/−) mice (Kotani et al., 2006) showed that, although these animals develop and age normally, they manifest an increased thrombotic tendency in the presence of endothelial damage. These findings confirmed the long-held view of PGI $_2$  as an endogenous antithrombotic agent and suggest that this antithrombotic system is activated in response to vascular injury. Indeed, Cheng et al. (2002) examined the interplay between IP and TP signaling in response to vascular injury by subjecting IP(−/−) mice and TP(−/−) mice to vascular injury by a balloon catheter. They found that IP deficiency increased, whereas TP deficiency decreased, injury-induced vascular proliferation and platelet activation. They further showed that the augmented response apparent in IP(−/−) mice was abolished by ablation of TP. Such augmented intimal hyperplasia was also seen in IP(−/−) mice subjected to common carotid artery ligation and transplant arteriosclerosis (Rudic et al., 2005). Furthermore, Xiao et al. (2001) subjected IP-deficient mice to cardiac ischemia-reperfusion injury and found that IP(−/−) mice exhibited a significantly larger size of myocardial infarct than wild-type mice, whereas TP(−/−) showed no difference, suggesting that receptor PGI $_2$ -IP signaling exerts a protective action during these conditions. As for the role of IP in chronic vascular disease, Kobayashi et al. (2004) examined progression of atherosclerosis in ApoE(−/−)/IP(−/−) double-knockout mice and found that atherosclerosis was accelerated in these mutant animals, despite the fact that they manifested similar plasma cholesterol and triglyceride concentrations. The lumen of the innominate artery was almost completely occluded in 45-week-old ApoE(−/−)/IP(−/−) mice. Mice deficient in IP do not show abnormalities of blood pressure under basal conditions. There are two reports suggesting the involvement of IP in development of hypertension under some conditions. Fujino et al. (2004) examined the contribution of prostanoids to the development of this condition by subjecting mice deficient in either IP or each of the four EP subtypes to a two-kidney, one-clip model of renovascular hypertension. They found that hypertension in this model was ameliorated in IP-deficient mice but not in any of the EP-deficient animals. Consistent with these observations, plasma renin activity, the abundance of renin mRNA in the kidney, and the plasma concentration of aldosterone were all substantially reduced in the IP knockout animals compared with those in the wild type. Given that

IP is expressed in the afferent glomerular arterioles, that expression of the renin gene is expanded to these arterioles in response to reduced perfusion of the kidney, and that PGI<sub>2</sub> induces renin release from cultured juxtaglomerular cells in vitro, these researchers suggested that PGI<sub>2</sub>-IP signaling directly stimulates renin release. Alternatively, such signaling may regulate the perfusion pressure of the juxtaglomerular apparatus locally and induce renin release indirectly. Francois et al. (2005) found that IP-deficient mice fed on a high-salt (6% NaCl) diet developed a significantly larger increase in blood pressure than wild-type mice without alteration in renin and aldosterone levels, and concomitantly developed cardiac hypertrophy and fibrosis. Their finding on salt-induced hypertension in IP(−/−) is consistent with that of Watanabe et al. (2005), who found that with a high-salt diet, systolic blood pressure in female IP(−/−) mice gradually increased, whereas that in the IP(+/+), TP(−/−), or TP(+/+) mice remained unchanged. On the other hand, Hara et al. (2005) examined cardiac hypertrophy and fibrosis in response to pressure overload. They subjected mice deficient in each prostanoid receptor to banding of the transverse aorta and found augmented hypertrophy and fibrosis only in IP(−/−) mice. Thus, a protective role of IP signaling in cardiac hypertrophy and fibrosis is found in mice on two different backgrounds and different models.

There are several studies using IP(−/−) mice to determine the role of IP in pain sensation and inflammatory swelling. Murata et al. (1997) found that in IP-deficient mice, the acetic acid writhing response was reduced to a level similar to that observed in wild-type mice treated with the COX inhibitor indomethacin. The capsaicin receptor TRPV1 is a nociceptor for heat and pH. Moriyama et al. (2005) found that thermal hyperalgesia in response to PGI<sub>2</sub> is absent not only in IP(−/−) mice but also in TRV1(−/−) mice. They further found that capsaicin-activated current in dorsal root ganglia neurons was augmented by PGI<sub>2</sub> as well as an IP agonist, and this augmentation was absent in neurons from IP(−/−) mice. It is noteworthy that the TRV1 augmentation is seen at a rather high concentration of PGI<sub>2</sub>, 1000 nM. As for inflammatory swelling, Murata et al. (1997) subjected IP-deficient mice to the carrageenin-induced paw swelling model and found that inflammatory swelling was reduced by ~50% in these animals, an effect similar in magnitude to that achieved by treatment of wild-type mice with nonsteroidal anti-inflammatory drugs. Yuhki et al. (2004) showed that IP, as well as EP<sub>2</sub> and EP<sub>3</sub> receptors, mediate exudate formation in carrageenin-induced mouse pleurisy, suggesting that PGE<sub>2</sub> and PGI<sub>2</sub> elicit inflammatory responses in a context-dependent manner (i.e., one dependent on the type of stimulus and site in the body) and that their contribution may also change during the course of inflammation. The inflammatory action of IP is not limited to inflammatory swelling exerted through regulation of the

peripheral circulation in acute inflammation. Honda et al. (2006) examined the role of prostanoids by backcrossing mice deficient in each prostanoid receptor on a DBA background and subjecting them to collagen-induced arthritis. They found that, whereas the incidence of arthritis was unaffected, the extent and progression of this condition were markedly suppressed in IP-deficient mice as well as in EP<sub>2</sub>-deficient mice treated with an EP<sub>4</sub> antagonist. These findings thus indicated that the PGI<sub>2</sub>-IP signaling and PGE<sub>2</sub> signaling at EP<sub>2</sub> and EP<sub>4</sub> receptors mediate joint inflammation in this model. Further analysis revealed that both PGI<sub>2</sub> and PGE<sub>2</sub> pathways regulate expression of arthritis-related genes, including those for IL-6, vascular endothelial growth factor-A, and RANKL, in synovial fibroblasts and thereby contribute to arthritic inflammation, bone destruction, and pannus formation. On the other hand, Pulichino et al. (2006) evoked arthritis by administering collagen antibodies to their IP(−/−) mice on a C57BL/6 background and found that arthritis was almost completely suppressed in this line of mice. They further used a newly developed IP antagonist [N-[4-(imidazolidin-2-ylideneamino)-benzyl]-4-methoxy-benzamide], administered it into heterozygous IP(+/−) mice before the antibody injection or after the onset of arthritis, and examined its prophylactic or therapeutic effects. They found that although the prophylactic administration of the IP antagonist exerted suppression of arthritis in IP(+/−) mice comparable with that in IP(−/−) mice, it showed no effects when administered after the disease onset. These results suggest that the PGI<sub>2</sub>-IP signaling functions as part of the process where self-reactive antibodies trigger the disease. However, whether this is the same mechanism as that identified by (Honda et al., 2006) remains unknown. In addition, starting with the finding that COX-2-deficient mice show exaggerated fibrotic response in bleomycin-induced pulmonary fibrosis, Lovgren et al. (2006) subjected mice deficient in either EP<sub>2</sub>, EP<sub>4</sub>, or IP to this model. They found that only IP-deficient mice exhibited significantly enhanced fibrosis and suggested that COX-2-derived PGI<sub>2</sub> exerts a protective action against fibrosis in this model.

PGI<sub>2</sub> has been shown to exert a protective effect on the gastric mucosa in response to injurious stimuli. IP-deficient mice have also been used to examine this phenomenon. Boku et al. (2001) and Arai et al. (2003) examined the role of IP in releasing CGRP in the stomach in response to a mild stimulant, 1 M NaCl, or capsaicin, which exerts protection to ethanol. They found that release of CGRP to both stimuli is impaired in IP(−/−) mice, and coadministration of capsaicin and beraprost enhanced the release, suggesting that endogenous PGI<sub>2</sub>-IP signaling functions to release CGRP for gastric protection. Furthermore, Terashima et al. (2009) used IP(−/−) mice and showed that iloprost can decrease histamine-stimulated acid secretion in the stomach in an IP-dependent manner. They further found that this

IP-mediated decrease is dependent on the somatostatin SST2 receptor. On the basis of these findings, they suggested that PGI<sub>2</sub> receptors may mediate somatostatin release in the gastric mucosa, which in turn suppresses acid secretion. Given expression of IP in various types of immune cells, including macrophages and T cells, Takahashi et al. (2002) subjected IP-deficient mice to the OVA-induced asthma model and examined its role. They found that IP(−/−) mice exhibited substantially higher plasma concentrations of antigen-specific and total IgE, indicating that PGI<sub>2</sub>-IP signaling is somehow involved in sensitization to IgE production. Finally, IP(−/−) mice were used to distinguish some actions of PGI analogs on IP and PPARβ. Ali et al. (2006) used lung fibroblasts from IP(−/−) mice and those from PPARβ(−/−) mice, examined inhibitory effects of treprostinil on their proliferation, and found that this activity was lost in PPARβ(−/−) cells but not in IP(−/−) cells.

**4. Agonists and Antagonists.** The upper row in Fig. 8 shows stabilization of the vinyl ether of PGI<sub>2</sub> by carbon replacement (iloprost), electron withdrawal with fluorine (AFP-07), and conjugation with or within aromatic moieties (beraprost, taprostene). In each case, orientation of the α-carboxyl terminus away from the ω-chain, which is crucial to potent IP agonism, is maintained; a more detailed treatment may be found in Wise and Jones (1996). A 16-methyl/18.19-triple bond structure has often been favored for the ω terminus. Iloprost and AFP-07 also show potent EP<sub>1</sub> agonism and to a lesser extent EP<sub>3</sub> agonism, whereas cicaprost is more selective (Dong et al., 1986; Lawrence et al., 1992; Abramovitz et al., 2000; McCormick et al., 2010) and is the preferred choice for an IP standard agonist. Taprostene is a IP partial agonist (Jones and Chan, 2001, 2005).

EP-157 (Fig. 8) is a PGH<sub>2</sub> analog exhibiting both TP antagonism and IP agonism (Armstrong et al., 1986); the

diphenylmethyl-heteroatomic group is critical to the latter activity (Jones et al., 1993). The related compounds in Fig. 4 also possess a similar or 1,2-diphenylethyl group and have been referred to as “nonprostanoid prostacyclin mimetics” (Meanwell et al., 1994). BMY-45778 (Fig. 8) is the most potent agent within this subclass (Jones et al., 1997; Seiler et al., 1997). Caution is necessary in using these agonists to characterize IP receptors because of their high lipophilicity, partial agonism (ocimibate; Merritt et al., 1991a) and ability to inhibit G<sub>q</sub>/PLC-driven effects (Chow et al., 2003). ONO-1301 (Kondo et al., 1995 also inhibits TxA<sub>2</sub> synthase because of the presence of a 3-pyridyl group). MRE-269 is obtained by hydrolysis of the methylsulfonamide moiety in the prodrug NS-304 (Kuwano et al., 2007, 2008).

Chemical library screening was the starting point for two classes of selective IP antagonist (Bley et al., 2006): RO-1138452 is a 2-(phenylamino)-imidazoline and RO-3244019 is an N-substituted phenylalanine (Fig. 9). In addition to its high affinity for human native (platelet) and recombinant IP receptors (pK<sub>i</sub>, 9.3 and 8.7, respectively), RO-1138452 also binds to platelet-activating factor (pK<sub>i</sub>, 7.9) and imidazoline (pK<sub>i</sub>, 8.3) receptors (Bley et al., 2006). RO-1138452 exhibited surmountable antagonism in functional studies on blood vessel preparations from human, rabbit, and guinea pig, with pA<sub>2</sub> values in the range 8.1 to 8.4 (Jones et al., 2006). Higher concentrations of RO-1138452 slightly suppressed the cicaprost maximum response, probably because of the EP<sub>3</sub> agonist action of cicaprost. In contrast, insurmountable antagonism was found for RO-1138452 in a chemokine release assay involving human airway epithelial cells (Ayer et al., 2008). RO-1138452 inhibition of IP agonist-induced cAMP response element-dependent transcription was not reversed after a 20-h “washout” period. It was pro-

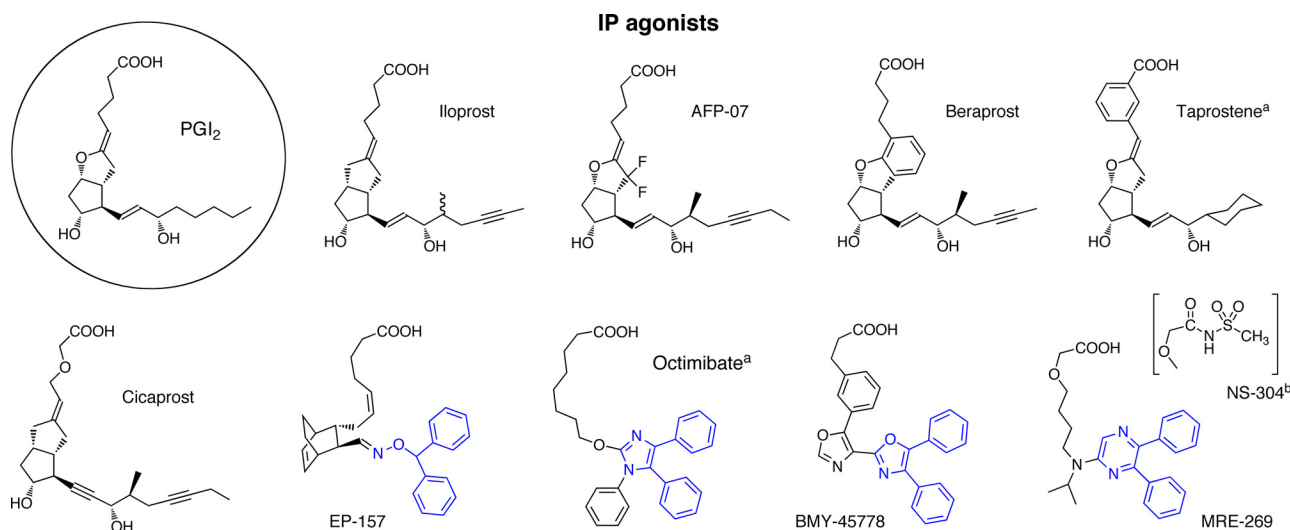


FIG. 8. Structures of agonists for the prostanoid IP receptor. PGI<sub>2</sub> (prostacyclin), the most active natural agonist, is shown in the circle. The diphenyl-heteroatomic unit critical to IP agonism in nonprostacyclin mimetics of prostacyclin is shown in blue. <sup>a</sup>, partial agonist. <sup>b</sup>, prodrug for MRE-269.



## IP antagonists

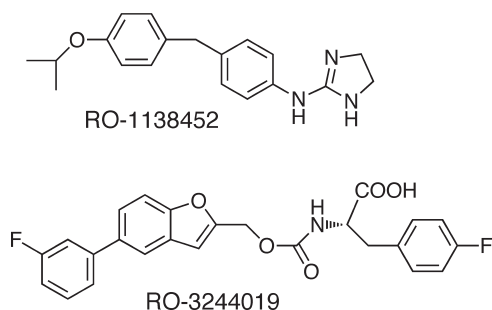


FIG. 9. Structures of IP receptor antagonists.

posed that allosterism or a state of antagonist hemiequilibrium and may underlie this profile (Ayer et al., 2008).

**5. Therapeutics.** Prostacyclin infusion has been in medical use for some time for treating pulmonary hypertension (Wise and Jones, 1996; Olschewski et al., 2004; Gryglewski 2008; Mubarak, 2010). The stable prostacyclin analog iloprost provides several practical advantages for intravenous therapy and may be given by aerosol (Olschewski et al., 2004; Mubarak, 2010). Treprostinil was developed for subcutaneous administration, and beraprost is an orally active prostacyclin mimetic (Olschewski et al., 2004; Mubarak, 2010). Prostacyclin and its analogs are also useful for treating advanced critical limb ischemia that may occur in Buerger's disease, Raynaud's syndrome, and atherosclerosis (Zardi et al., 2005; Gryglewski, 2008). New IP agonists continue to be developed, the latest being NS-304 (Fig. 8). This is both long-acting and has better selectivity than beraprost or iloprost and represents better future therapy for pulmonary arterial hypertension (Kuwano et al., 2008). Beyond critical care, IP agonists have not enjoyed wide use as medical therapy. The potential for profound decreases in blood pressure and for IP receptor-induced pain and inflammation provide a significant barrier to widespread use. Other limited and certain local therapeutic applications may be possible.

There is also an IP receptor therapeutic focus on designing antagonists. The impetus for this is to provide analgesic/anti-inflammatory agents, with no effect on bleeding time (Bley et al., 2006; Pulichino et al., 2006; Brescia et al., 2007; Zhao et al., 2008; Jones et al., 2009). A potential use for IP antagonists for treating overactive bladder disorders has been advanced. Thus, RO-3244019 decreased bladder contraction frequency and increased the micturition threshold in isovolumetric bladder contraction and refill models, respectively (Cefalu et al., 2007), and improved intercontractile interval and voiding volumes as assessed by cystometry (Khara et al., 2007).

## E. TP Receptors

**1. Second Messenger Signaling.** TP receptors have been shown to couple to  $G_q$ , thereby initiating the PLC $\beta$

→ inositol trisphosphate/diacylglycerol →  $[Ca^{2+}]_i$ /PKC signaling cascade (Hirata et al., 1991; Dorn and Becker, 1993; Kinsella et al., 1997; Offermanns, 2006; Nakahata 2008). Subsequent studies have strongly implicated  $G_{12/13}$ -Rho as a major signaling pathway for TP receptors (Huang et al., 2004; Honma et al., 2006; Mir and Le Breton, 2008; Nakahata, 2008; Song et al., 2009; Zhang et al., 2009; Saito et al., 2010). More than any other prostanoid receptor, TP receptors have been subjected to extensive second messenger pathway analyses. Thus, at the molecular level, there is detailed understanding of TP receptor-G protein interactions obtained by using G protein fusion constructs and guanosine 5'-O-(3-thio) triphosphate binding studies (Hildebrandt, 2006; Zhang et al., 2006, 2009). The G protein coupling repertoire for TP receptors is beyond extensive; it seems all-encompassing. Other members of the  $G_q$  family are TP receptor-linked:  $G_{11}$  (Kinsella et al., 1997),  $G_{15}$ , and  $G_{16}$  (Offermanns and Simon, 1995), activating the  $G_q$ -mediated signaling cascade. TP receptors may couple to  $G_s$  (Hirata et al., 1996; Walsh et al., 1998; Mir and Le Breton, 2008) and  $G_i$  (Ushikubi et al., 1994; Song et al., 2009). Finally, TP receptors are reported to couple to  $G_h$  (Vezza et al., 1999).

TP receptor signaling may result in transactivation, for example of ERK 1/2 (Nakahata, 2008) and EGF receptors (Uchiyama et al., 2009). Likewise, TP receptors possess a PKA/protein kinase G phosphorylation site and four PKC phosphorylation sites, providing mechanisms for modulation/desensitization (Huang et al., 2004). The type of G protein coupling may also alter TP $\alpha$  conformation, which may influence ligand binding and activation of the receptor (Zhang et al., 2006).

Human TP receptors exist in two isoforms, TP $\alpha$  and TP $\beta$  (Hirata et al., 1991; Raychowdhury et al., 1994). TP $\beta$  is an alternative mRNA splicing variant with an extended carboxyl terminus. The differences in intracellular termini may influence desensitization, internalization, and G protein coupling (Parent et al., 1999; Reid and Kinsella, 2007; Nakahata, 2008). Thus, the TP $\beta$  isoform may couple to  $G_i$  (Hirata et al., 1996). However, only the TP $\alpha$  isoform is translated in platelets (Habib et al., 1999). It is noteworthy that because TP receptors play a prominent role in mediating the activity of isoprostanes, heterodimerization of TP $\alpha$  and TP $\beta$  have been reported to enhance isoprostane signaling (Wilson et al., 2007).

**2. Distribution and Biological Functions.** The platelet has always been a major focus of TP receptor research. The TP receptor was originally purified from human platelets (Ushikubi et al., 1989). TP receptor activation produces shape change and platelet aggregation, providing a positive feedback event for causing thrombus formation and thrombosis (Offermanns, 2006; Nakahata, 2008). TP receptor-deficient mice exhibit prolonged bleeding times and are unable to form stable thrombi (Thomas et al., 2001). It has been reported that

TxA<sub>2</sub> promotes soluble CD40 ligand release from platelets (Enomoto et al., 2010). The platelet TxA<sub>2</sub>-vascular endothelial PGI<sub>2</sub> balance is of central importance to the rationale for low-dose aspirin therapy and is the basis for COX-2 inhibitor cardiovascular side effects, underscoring the pathophysiological importance of TP receptors.

Beyond platelets, TP receptors are widely expressed in the cardiovascular system and represent a liability for cardiovascular disease at all levels. In endothelial cells, TxA<sub>2</sub> accelerates the expression of adhesion proteins (Ishizuka et al., 1998) and impairs insulin signaling (Song et al., 2009). Increased TxA<sub>2</sub> production may contribute to the development of endothelial dysfunction, with resultant vasoconstriction (Gendron and Thorin, 2007; Francois et al., 2008; Denniss and Rush, 2009; Félétou et al., 2009; Graham and Rush, 2009). TxA<sub>2</sub> has also been implicated in atherogenesis (Dogné et al., 2005), and mice that are TP receptor-deficient develop fewer atherosclerotic lesions (Kobayashi et al., 2004). The presence of TP receptors on peripheral blood monocytes (Allan and Halushka, 1994) would contribute to the formation of atherosclerotic plaque. Cardiovascular

TP receptor expression even extends to cardiomyocytes, and the TxA<sub>2</sub> mimetic U-46619 (Fig. 10) causes arrhythmia (Wacker et al., 2009). TP receptors are also located on afferent sympathetic nerve endings in the heart and may participate in the sympathoexcitatory reflex that occurs during myocardial ischemia (Fu et al., 2008).

Indirect evidence for functional TP receptor expression on peripheral sensory neurons is provided by the pruritogenic activity of U-46619 (Andoh et al., 2007). Introduction of U-46619 into the fourth ventricle rapidly produces emesis (Kan et al., 2008). In relation to the CNS, TP receptors are reported to be expressed on oligodendrocytes (Blackman et al., 1998; Mir and Le Breton, 2008) and human astrocytoma cells (Honma et al., 2006).

In addition to blood vessels, TP receptors are expressed in several smooth muscle types (Coleman et al., 1994b; Nakahata, 2008). Among these, the lung has always featured prominently. The presence of TP receptors on human bronchial smooth muscles was pharmacologically defined in 1989 (Coleman and Sheldrick, 1989), to provide early impetus to TP receptor studies in

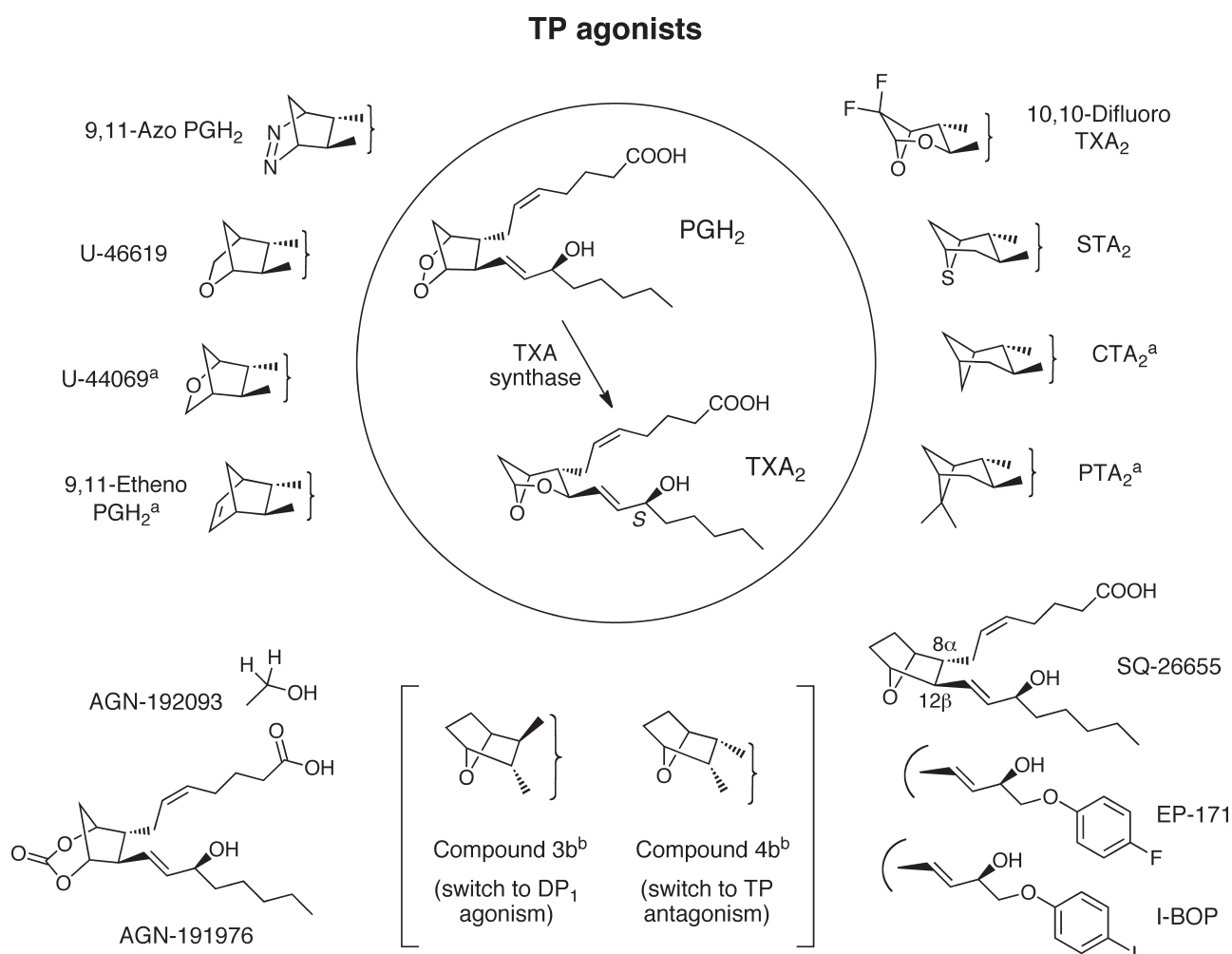


FIG. 10. Structures of agonists for the prostanoid TP receptor. TxA<sub>2</sub> and PGH<sub>2</sub>, the most active natural agonists, are shown in the circle. Only structural alterations relative to PGH<sub>2</sub> and TxA<sub>2</sub> are illustrated in the figure. The vertical bracket indicates the presence of 2-series α- and ω-chains. <sup>a</sup>, partial agonist.

the bronchopulmonary system. TP receptors played a substantial role in mediating leukotriene-mediated bronchoconstriction (Piper and Samhoun, 1981; Weichman et al., 1982).  $\text{TxA}_2$  is a potent constrictor of bronchial smooth muscle and has been implicated in both asthma and chronic obstructive pulmonary disease (Rolin et al., 2006). TP receptors are also uterotonic (Senior et al., 1991) and seem to be particularly important in parturition, because  $\text{PGF}_{2\alpha}$  responsiveness, but not that of  $\text{TxA}_2$ , is lost in the human myometrium during labor (Fischer et al., 2008). Finally, on the smooth muscle cell theme, TP receptors seem capable of differentiating human stem cells to smooth muscle-like cells (Kim et al., 2009).

In common with all prostanoid receptors, TP receptors have been implicated in cancer. U-46619 has been reported to stimulate endothelial cell migration in vivo and angiogenesis and lung metastasis in vivo (Nie et al., 2000). TP receptors also cause proliferation and growth of human lung cancer cells (Li and Tai, 2009; Wei et al., 2010). TP receptors are elevated in prostate cancer and stimulate motility of human prostate cancer cell lines (Nie et al., 2008). It is noteworthy that the  $\text{TP}\beta$  isoform, but not  $\text{TP}\alpha$ , promoted cell proliferation and migration and invasion by bladder cancer cells; moreover,  $\text{TP}\beta$  receptor expression was increased in tissue obtained from bladder cancer patients (Moussa et al., 2008).

Among the additional functions attributed to TP receptors is their role in immune regulation. Naive T cells obtained from mice expressing TP receptors suppress interaction between dendritic cells and inhibit dendritic cell-dependent T cell proliferation (Kabashima et al., 2003a). In TP-deficient mice, immune responses to foreign antigenic stimuli were enhanced, a phenomenon reproduced in wild-type mice by TP receptor blockade during the sensitization period (Kabashima et al., 2003a). In contrast, splenocytes obtained from IP-deficient mice exhibited a reduced proliferative response to phytohemagglutinin or anti-CD3 antibody (Thomas et al., 2003). Although survival of transplanted hearts from wild-type mice into TP-deficient mice was not prolonged, there was reduced pathological severity in the allografts (Thomas et al., 2003). A role for  $\text{TxA}_2$  in inflammatory bowel disease is also a possibility (Rampton and Collins, 1993). The vasoconstrictor activity of TP receptors may also be a factor in renal disease (Michel et al., 2008; Araujo and Welch, 2009).  $\text{TxA}_2$  has also been advanced as a key regulator during *Trypanosoma cruzi* infection and may contribute to mortality and parasitism (Ashton et al., 2007). Finally, thromboxane synthase and TP receptors are expressed in the murine retina and vasoconstriction in the streptozotocin diabetes model was attenuated by TP receptor blockade (Wright et al., 2009a,b).

**3. Gene Deletion Studies.** Two lines of  $\text{TP}(-/-)$  mice were generated independently (Thomas et al., 1998; Kabashima et al., 2003a). Both lines of mice show no ab-

normalities of blood pressure under the basal conditions but do show increased bleeding tendencies and are resistant to cardiovascular shock induced by intravenous infusion of arachidonic acid or the TP agonist U-46619. Given the antagonistic actions of  $\text{TxA}_2$  and  $\text{PGI}_2$  on platelets and blood vessels, TP signaling in cardiovascular homeostasis was studied using these lines of mice by comparing their phenotypes with those of  $\text{IP}(-/-)$  mice. Cheng et al. (2002) subjected  $\text{IP}(-/-)$  mice and  $\text{TP}(-/-)$  mice to vascular injury by a balloon catheter and found that IP deficiency increased injury-induced vascular proliferation and platelet activation, whereas TP deficiency decreased it. They further showed that the augmented response apparent in  $\text{IP}(-/-)$  mice was abolished by ablation of TP. These results showed that, once endothelial integrity is disrupted, IP functions protectively, whereas TP aggravates the remodeling. The same group also examined the involvement of TP signaling in vascular remodeling using the external carotid artery ligation model that retains endothelial integrity (Rudic et al., 2005). In this model, treatment of animals with nimesulide augmented the neointimal hyperplastic response of the artery, which was reduced by the loss of TP receptors, suggesting again that  $\text{TxA}_2$ -TP signaling facilitates vascular remodeling after injury or stress. During the above ligation model, production of not only  $\text{TxA}_2$  but 8,12-iso-isoprostane  $\text{F}_{2\alpha}$  was increased. Isoprostanes (iPs) are free radical-catalyzed products of arachidonic acid that reflect lipid peroxidation in vivo. Among them,  $\text{iPF}_{2\alpha}\text{-III}$  and  $\text{iPE}_2\text{-II}$  can activate platelets and increase vascular tone. The pressor response in vivo and platelet aggregation in vitro induced by these substances were abolished in  $\text{TP}(-/-)$  mice, suggesting that actions of these iPs are mediated by TP (Audoly et al., 2000). To test the involvement in chronic vascular diseases, Kobayashi et al. (2004) generated the  $\text{ApoE}(-/-)/\text{TP}(-/-)$  double-knockout mice, and examined the effects of TP deficiency on the progression of atherosclerosis. In contrast to acceleration of atherosclerosis in the  $\text{ApoE}(-/-)/\text{IP}(-/-)$  double-knockout mice, atherosclerosis was delayed in the  $\text{ApoE}(-/-)/\text{TP}(-/-)$  mice, despite the fact that they manifested similar plasma cholesterol and triglyceride concentrations. A recent bone marrow transfer experiment (Zhuge et al., 2006) indicated that the effects of TP deficiency attenuating atherogenesis cannot be attributed simply to bone marrow-derived cells such as macrophages. Finally, Francois et al. (2005) found that  $\text{IP}(-/-)$  mice developed salt-sensitive hypertension, which led to cardiac hypertrophy and severe cardiac fibrosis, and coincidental deletion of TP did not suppress hypertension but ameliorated the hypertrophy and abolished the fibrosis, suggesting that  $\text{TxA}_2$ -TP signaling is responsible for hypertension-induced cardiac complications. These findings have verified that the presumed antagonistic roles of the  $\text{PGI}_2$ -IP signaling and  $\text{TxA}_2$ -TP signaling exists in not only subacute vascular remodeling but also chronic vascular lesions such as atherosclerosis and cardiac fibrosis and



may explain the increased incidence of cardiovascular events that has been observed in clinical trials with COX-2 inhibitors. Thus, the loss of TP usually suppresses the progression of pathological vascular conditions. However, a paradoxical example was reported in kidney lesions induced by  $N^G$ -nitro-L-arginine methyl ester. Francois et al. (2008) subjected TP(−/−) mice to the  $N^G$ -nitro-L-arginine methyl ester-induced hypertension model and found that the extent of hypertension and resultant cardiac hypertrophy were significantly suppressed in TP(−/−) mice compared with wild-type mice but that the kidney lesion, including glomerulosclerosis, tubule vacuolization, and chronic intestinal inflammation, was worsened with the loss of TP, suggesting that TxA<sub>2</sub>-TP signaling exerts protective actions against kidney injury, at least in this model. In addition, as described in the FP section, the inflammatory tachycardia induced by either cytokines or LPS was suppressed in mice deficient in TP or FP and was almost completely abolished in mice deficient in both TP and FP. Because both PGF<sub>2α</sub> and I-BOP, a TP-selective agonist, induced potent positive chronotropic effects on direct application to the nodal area, these results suggest that both TxA<sub>2</sub> and PGF<sub>2α</sub> are produced in response to inflammatory stimuli in situ in the heart and act on TP and FP receptors expressed on pacemaker cells to induce tachycardia. In addition, TxA<sub>2</sub>-TP signaling also regulates the microcirculation under pathological conditions. During endotoxemia, TNF-α induces dysfunction of the hepatic microcirculation. Katagiri et al. (2008) found that this TNF-α-induced dysfunction, examined as leukocyte adhesion to microvessel endothelial cells, increased the number of nonperfused sinusoids, which was significantly lessened in TP(−/−) mice, indicating that TNF-α mobilizes TxA<sub>2</sub>-TP signaling and impairs the microcirculation.

In addition to the cardiovascular actions of TP described above, studies on TP(−/−) mice revealed thus-far-unappreciated actions of TP in immunity and inflammation. Kabashima et al. (2003a) noticed lymphadenopathy and splenomegaly in aged TP(−/−) mice and found that immune response to foreign antigens was enhanced in TP(−/−) mice. They further found that stimulation of TP receptors enhanced chemokinesis of T lymphocytes and down-regulated dendritic cell-dependent T-cell proliferation in vitro. These findings led them to suggest that TxA<sub>2</sub>-TP signaling in T cells regulates the interaction between T cells and dendritic cells and thereby the extent of the immune response. On the other hand, Thomas et al. (2003) found that T cells deficient in TP receptor exhibited less proliferation in response not only to phytohemagglutinin or anti-CD3 antibody but also in the mixed lymphocyte response (MLR). Reduced proliferation of TP(−/−) lymphocytes in MLR was mimicked by the MLR of wild-type T cells treated with a TP antagonist, 7-(3-((2-((phenylamino)carbonyl)hydrazino)methyl)-7-oxabicyclo (2.2.1)hept-2-yl)-5-heptenoic acid (SQ-29548) or a thromboxane synthase inhibitor, carboxyhexyl imidazole. These findings are opposite those of Kabashima et

al. (2003a), who found that T-cell proliferation in MLR was suppressed by treatment with a TP agonist, I-BOP. Thomas et al. (2003) reported that their TP(−/−) mice exhibited prolonged cardiac allograft survival compared with wild-type mice. We do not know why such a discrepancy arose using different lines of TP(−/−) mice. In addition to these functions of TP in the peripheral immune system, TxA<sub>2</sub>-TP signaling also seems to regulate the immune system centrally. Ushikubi et al. (1993) found that TP mRNA was markedly expressed in the thymus, particularly in CD4<sup>−</sup>/CD8<sup>−</sup> and CD4<sup>+</sup>/CD8<sup>+</sup> immature thymocytes, and that the addition of a TP agonist, stable TxA<sub>2</sub> analog, induced apoptosis of CD4<sup>+</sup>/CD8<sup>+</sup> cells. Rocha et al. (2005) found that administration of LPS into mice markedly increased production of TxA<sub>2</sub> and PGE<sub>2</sub> in the thymus, and caused apoptotic deletion of CD4<sup>+</sup>/CD8<sup>+</sup> cells there. They then demonstrated that thymocyte apoptosis in response to LPS was significantly attenuated in TP(−/−) mice, suggesting that thymocyte apoptosis mediated by TxA<sub>2</sub>-TP signaling functions physiologically.

In addition to these actions in the immune system, TxA<sub>2</sub>-TP signaling exerts its action in a different system to combat against infection. Ashton et al. (2007) found that a parasite, *T. cruzi*, is itself capable of producing TxA<sub>2</sub> and that TP(−/−) mice exhibited higher mortality and more severe cardiac pathology and parasitism than wild-type mice after infection. Bone marrow transfer experiments showed that TP receptor in somatic cells, and not bone marrow-derived cells, is important for protection against parasites; how it functions remains to be determined.

**4. Agonists and Antagonists.** Thromboxane A<sub>2</sub> and its precursor endoperoxide PGH<sub>2</sub> both activate TP receptors. Both are very chemically unstable and therefore of limited practical use. For this reason, stable analogs were developed. The methanoepoxy analog 9-11-dideoxy-11α,9α-epoxymethano-prostaglandin F<sub>2α</sub> (U-46619; Fig. 10) has been widely used. More potent compounds, such as I-BOP, are also available for study but may have an inconveniently slow onset and offset in isolated tissue preparations.

Both PGH<sub>2</sub> and TxA<sub>2</sub> activate TP receptors, the latter showing higher potency (Needleman et al., 1976; Svensson and Hamberg, 1976; Salzman et al., 1980; Armstrong et al., 1985; Vezza et al., 2002), although their chemical lability and the enzymatic conversion of PGH<sub>2</sub> to PGD<sub>2</sub>, PGE<sub>2</sub>, PGF<sub>2α</sub>, and PGI<sub>2</sub> complicate these measurements (Hornberger and Patscheke, 1989). Stabilization of the TxA<sub>2</sub> ring structure by difluoro substitution at C10 yields a potent TP agonist (Morinelli et al., 1989). However, the majority of stable TP agonists derive from substitution of one or both ring oxygens in PGH<sub>2</sub> or TxA<sub>2</sub> with carbon (Fig. 10). U-46619 (the most commonly used standard agonist), 9,11-azo PGH<sub>2</sub> and stable TxA<sub>2</sub> analogs behave as full agonists on isolated vascular and respiratory smooth muscle and human platelet prepara-

tions (Tymkewycz et al., 1991), as does the cyclic carbonate AGN-191976 (Fig. 10) (Krauss et al., 1996). U-44069, 9,11-etheno PGH<sub>2</sub>, pinane TxA<sub>2</sub>, and carbocyclic TxA<sub>2</sub> often show partial agonism on smooth muscle preparations; that is, they show a reduced maximum response and block the action of U-46619 without affecting the action of other contractile agonists (e.g., phenylephrine or histamine) (Jones et al., 1982; Krauss et al., 1996). They may also show partial agonism on human platelets, but results from this assay system must be interpreted cautiously. Thus, there are three distinct, concentration-dependent, functionally related components of "platelet activation," namely shape change, primary (reversible) aggregation, and secondary aggregation as a result of release of mediators, including PGH<sub>2</sub>/TxA<sub>2</sub> from de novo synthesis. The generated PGH<sub>2</sub> could spontaneously decay to PGE<sub>2</sub> and/or PGD<sub>2</sub>, with corresponding activation of EP<sub>3</sub> receptors enhancing platelet activation (Oelz et al., 1977; Matthews and Jones, 1993) and DP<sub>1</sub> receptors inhibiting platelet activation (Keery and Lumley, 1988) through changes in cAMP levels (see Gresele et al., 1988). True TP partial agonists could elicit either submaximal shape change or complete shape change and reversible aggregation, as is the case for 9,11-etheno PGH<sub>2</sub> (Jones and Wilson, 1980). However, functional antagonism may also modulate the response as in the case of carbocyclic TxA<sub>2</sub>, which weakly raises the cyclic AMP level (possibly through activation of sensitive DP<sub>1</sub> and/or IP systems) sufficient to suppress the maximum aggregation response (Armstrong et al., 1985). Finally, COX/thromboxane synthase involved in the release reaction may be inhibited by PGH/TxA analogs (Wlodawer et al., 1971). Taking these issues into account, Krauss et al. (1996) identified the C1-alcohol derivative AGN-192903 (Fig. 10) as an agent that behaved as a potent full agonist on rat aorta but only caused shape change in human platelets at high concentration. The authors discuss their findings in relation to

the identification of two TP receptor subtypes (Takahara et al., 1990; Furci et al., 1991), one of which elicits smooth muscle contraction and platelet shape change, whereas the other elicits platelet aggregation (see also section on identification of TP $\alpha$  and TP $\beta$  subtypes).

Analogues with a 7-oxabicyclo[2.2.1]heptane ring system (Fig. 10, bottom right) were prepared as TP agonists on the basis of a favorably directed ring oxygen (Sprague et al., 1985). Of the eight possible  $\alpha$ -oxy isomers, the "natural" 8 $\alpha$ ,12 $\beta$ ,15S analog (SQ-26655; Fig. 10) is a potent TP agonist. Introduction of a 16-*p*-halophenoxy moiety into the  $\omega$  terminus, as in EP-171 (Fig. 10) (Wilson et al., 1988) and I-BOP (Sessa et al., 1990), markedly enhances TP agonism. Near-maximal shape change of the human platelet elicited by EP-171 at 0.1 nM is only slowly reversed by a high concentration of TP antagonist, reflecting the slow dissociation of this agonist from the TP receptor (Jones et al., 1989). [<sup>125</sup>I]BOP is a useful radioligand for the TP receptor (Morinelli et al., 1989). The 8 $\alpha$ ,12 $\alpha$ ,15R analog retains TP agonism, whereas the 8 $\alpha$ ,12 $\alpha$ ,15S analog shows a switch to TP antagonism (Sprague et al., 1985) and was the starting point for potent and selective TP antagonists such as SQ-29548 and BMS-180291 (ifetroban; Fig. 11). The 8 $\beta$ ,12 $\alpha$ ,15S analog shows broad inhibitory activity on human platelets, which was subsequently attributed to DP<sub>1</sub> agonism (see Fig. 11).

TP antagonist design has been ongoing for more than 2 decades. Many potent and selective TP antagonists of diverse structure have been synthesized (Jones et al., 2009). Partial agonism is prevalent among carba-ring analogs of PGH<sub>2</sub> or TxA<sub>2</sub> (see Wilson and Jones, 1985), and further modification of the  $\omega$ -chain often leads to pure antagonism. A prime example is I-SAP (Fig. 11), which contains a pinane ring akin to the dioxabicyclo [3.1.1]heptane ring of TxA<sub>2</sub> and *trans* orientation of the

### TP antagonists

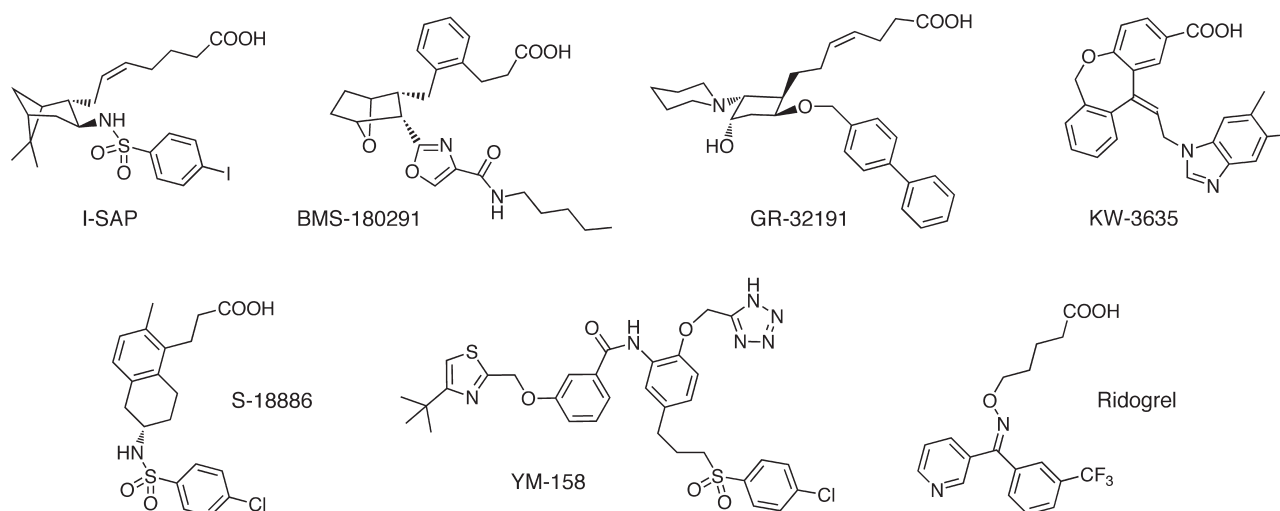


FIG. 11. Structures of representative antagonists for the prostanoid TP receptor.

side chains (Naka et al., 1992). In other prostanoid-like antagonists, a *cis* orientation of the side chains affords high affinity as in GR-32191 (Lumley et al., 1989) and BMS-180291 (Ogletree et al., 1993) (Fig. 11). A benzenesulfonamido moiety present in I-SAP features in simpler TP antagonist molecules, such as BM-13505 (daltroban; Yanagisawa et al., 1987), Z-335 (Tanaka et al., 1998) and S-18886 (terutroban; Fig. 11) (Cimetière et al., 1998); the distance from the C1-carboxylate group seems to be critical. 14,15-Epoxy-eicosatrienoic acid has been claimed to be an endogenous TP antagonist; it shows modest affinity for human DP<sub>1</sub>, FP, and TP receptors ( $K_i$  = 6.1, 5.3, and 3.2  $\mu$ M) with lower affinity for other prostanoid receptors ( $\geq 13$   $\mu$ M) (Behm et al., 2009).

For many of these compounds, surmountable antagonism on isolated smooth muscle preparations points toward reversible-competitive mechanism, and conventional Schild analysis confirms this classification. However, a slow approach to steady-state block by high-affinity antagonists can confound  $pA_2$  estimation. Under the Cheng-Prusoff inhibition-curve protocol, 6-(2-(2-chlorophenyl-4-hydroxyphenyl)-1,3-dioxan-5-yl)hexanoic acid (ICI-192605) (Brewster et al., 1988) had not reached steady-state block on guinea pig aorta after 90-min incubation ( $pA_2$  = 10.25) (Jones et al., 2008). With higher affinity TP antagonists in human and rat platelet assays, surmountability is seen for the shape-change response, whereas insurmountability occurs for aggregation. This profile is still compatible with a reversible-competitive mechanism, because slow dissociation of the high-affinity antagonist from the TP receptor retards agonist (U-46619) occupancy in the early stage of the aggregation response, thereby favoring the disaggregation process and insurmountability; in contrast, the nonfading shape-change response affords a truer measure of dose ratios at steady state (Armstrong et al., 1985; Jones et al., 1989; Lumley et al., 1989; Tymkewycz et al., 1991; Ogletree et al., 1993).

There has been much debate about the existence of different TP receptor subtypes in platelet and vascular smooth systems (Mais et al., 1985, 1988; Swayne et al., 1988; Morinelli et al., 1989; Masuda et al., 1991; Tymkewycz et al., 1991; Folger et al., 1992). It is certainly clear that species heterology exists; for example, higher affinities are often found for human and rat platelet TP receptors compared with rabbit platelet TP receptors (Tymkewycz et al., 1991). However, the situation may be more complex. There is evidence for two saturable binding sites for TP agonists on human platelets using several radioligands (Armstrong et al., 1983; Pollock et al., 1984; Ahn et al., 1988; Hedberg et al., 1988). The high-affinity site was associated with the platelet shape change (and increase in cytosolic  $Ca^{2+}$ ), whereas the lower-affinity site was associated with aggregation (and activation of PLC) (Dorn, 1989; Takahara et al., 1990). <sup>3</sup>H-labeled GR-32191 (vapiprost) was bound reversibly to the “shape change site” and irreversibly to the “aggre-

gation site” (Takahara et al., 1990). Given the structure of GR-32191 (Fig. 11), it is unlikely that covalent bonding is involved. Exposure of human platelets to GR-32191 for 30 min resulted in approximately 50% loss of binding sites for either [<sup>3</sup>H]GR-32191 or [<sup>3</sup>H]SQ-29548, whereas neither SQ-29548 nor BM-13177 (sulotroban) affected  $B_{max}$ . It was speculated that GR-32191 binds to internalized TP receptors (Armstrong et al., 1983). 4-[2-(4-azido-benzenesulfonylamino)-ethyl]phenoxyacetic acid, a light-activated, covalent-binding TP antagonist, also discriminated these platelet sites by blocking aggregation but not shape change induced by U-46619 (Zehender et al., 1988). Two TP receptor isoforms (TP $\alpha$  and TP $\beta$ ) have been identified using a human umbilical vein endothelial cDNA library (Raychowdhury et al., 1994), and mRNAs for the  $\alpha$  and  $\beta$  isoforms have been detected in human platelets (Hirata et al., 1996). However, these isoforms, which arise by alternative gene splicing and differ only in their cytoplasmic tails, do not show the ligand discrimination typical of the high- and low-affinity binding sites. Finally, only the TP $\alpha$  isoform was found in human platelets (Habib et al., 1999).

TP antagonism associated with IP agonism, TX synthase inhibition and LT receptor antagonism in the same molecule are all known. The bicyclo[2.2.2]octene PGH<sub>2</sub> analog EP-157 (Figs. 8 and 11) activates IP receptors in both platelet and vascular systems (Armstrong et al., 1986, 1989; Jones et al., 1993). The presence of a diarylhetero(cyclic) moiety in the  $\omega$ -chain is crucial (Jones et al., 1993). Similar properties were found for octimibate, which lacks a prostanoid ring system (Merritt et al., 1991a,b) and is a member of a series of nonprostanoid prostacyclin mimetics (Meanwell et al., 1994; Seiler et al., 1997). Some of these agents also inhibit (nonprostanoid) Gq-PLC-driven responses (Chow et al., 2001).

Combined TP antagonism/thromboxane synthase inhibition usually requires the presence of a (*N*)-imidazole (dazoxiben; Randall et al., 1981) or a pyridin-3-yl group (isbogrel; Imura et al., 1990; ridogrel, Hoet et al., 1990; Fig. 11) to combine with the heme site of the synthase (see Hsu et al., 1999). Existing TP antagonists have also been modified to include similar reactive moieties: ZD-1542 (Brownlie et al., 1993), a relative of ICI-192605, contains a pyridin-3-yl group and GR-83783 (Campbell et al., 1991a), a relative of GR-32191, has a 4-(pyridin-3-yl)-phenyl moiety. In addition, sulotroban/daltroban moieties have been combined with ridogrel/isbogrel moieties (Campbell et al., 1991b; CGS-22652 (Bhagwat et al., 1993; Soyka et al., 1994) and the whole or part of the ICI-192605 nucleus has been tethered to either a dazoxiben or an isbogrel nucleus (Ackerley et al., 1995). More recently, compounds with dual antagonist properties have been designed. For example, (2-(*N*-(4-(4-chlorobenzenesulfonylamino)butyl)-*N*-(3-(4-isopropylthiazol-2-yl)methoxy)benzyl)sulfamoyl)benzoic acid (KP-496) (Mizutani et al., 2008; Ishimura et al., 2009) and YM-



158 (Fig. 11) (Arakida et al., 1998) are dual TP/cysteinyl leukotriene antagonists.

**5. Therapeutics.** The original purpose for designing TP antagonists was as cardiovascular therapy but low-dose aspirin has proven a more economically viable proposition (Jones et al., 2009). Nevertheless, even recently the TP antagonist terutroban was shown to exhibit superior antithrombotic activity compared with aspirin in humans (Bal Dit Sollier et al., 2009). Another early indication for TP antagonists was asthma, which met with some clinical success, albeit limited (Rolin et al., 2006; Jones et al., 2009). Other early indications do not seem to have met with any clinical success. These include cancer, glomerulonephritis, allergic rhinitis, inflammatory bowel disease, septic shock, and diabetes (Jones et al., 2009). The unique role of the TP $\beta$  receptor isoform in bladder cancer and the delayed onset and prolonged survival afforded by TP antagonist treatment in mice transfected with bladder cancer cells (Moussa et al., 2008) holds promise for therapeutic utility in at least one form of cancer.

There are other potential uses for TP antagonists that have emerged more recently. The TP antagonist seratro-dast seems to possess antitussive properties (Xiang et al., 2002; Ishiura et al., 2003). Ramatroban has been reported to attenuate cough in subjects with cough variant asthma (Kitamura et al., 2003a). The potential use of TP antagonists for treating preterm labor is not new but deserves new impetus based on recent findings on the human myometrium that PGF<sub>2 $\alpha$</sub>  responsiveness, but not TxA<sub>2</sub> responsiveness, of the uterus is lost during labor (Fischer et al., 2008). The most recent potential uses for TP antagonists are listed in Table 10.

## F. DP<sub>2</sub> Receptors (CRTR2)

**1. Second Messenger Signaling.** Three independent but convergent research pathways led to the discovery of the DP<sub>2</sub> (CRTH2) receptor subtype. A new surface marker on human Th2 cells in vivo (Nagata et al., 1999) was termed the “chemoattractant receptor-homologous molecule expressed on Th2 cells” (CRTH2). This proved to be identical to the orphan G protein-coupled receptor GPR44 (Marchese et al., 1999), which had sequence

homology similar to that of typical chemoattractant receptors. Nevertheless, the naturally occurring ligand for the CRTH2 receptor was subsequently found to be PGD<sub>2</sub> (Hirai et al., 2001; Monneret et al., 2001), and the term DP<sub>2</sub> was introduced. There are now three descriptors: CRTH2, DP<sub>2</sub>, GPR44, with license for upper/lower case and subscripted/nonsubscripted variations. It has been suggested that the widespread distribution of this receptor, which extends far beyond the immune system, makes placement in the prostanoid receptor classification as DP<sub>2</sub> more appropriate (Jones et al., 2009). The DP<sub>2</sub> designation is used herein, given the topic of this review.

DP<sub>2</sub> receptors are G<sub>i</sub>-coupled, but signal transduction pathways have been subject to only limited investigation. DP<sub>2</sub> receptor activation may result in pertussis toxin-sensitive decreases in cAMP levels (Sawyer et al., 2002; Gallant et al., 2007) and Ca<sup>2+</sup> mobilization (Hirai et al., 2001; Sawyer et al., 2002). PI3K signaling has also been implicated in mediating DP<sub>2</sub> effects (Hata et al., 2003; Xue et al., 2007). DP<sub>2</sub> receptor trafficking has been studied and, interestingly, PGD<sub>2</sub> induced DP<sub>2</sub> but not DP<sub>1</sub> receptor internalization (Gallant et al., 2007), which was decreased by inhibition of PKA and PKC. DP<sub>2</sub> receptor internalization may be regulated by PKC, GRK2, GRK3, GRK6 and arrestin-3 (Gallant et al., 2007) and the determinants located in the carboxyl terminus have been studied (Roy et al., 2010).

**2. Distribution and Biological Functions.** By combining functional data and transcription profiles together, it is clear that DP<sub>2</sub> receptor expression is widespread. Northern blotting revealed high expression in the human stomach, small intestine, heart, and thymus; intermediate expression in the colon, spinal cord, and blood; and lower expression in the brain, skeletal muscle, and spleen (Sawyer et al., 2002). Functional studies demonstrate DP<sub>2</sub> receptors present in smooth muscle, the cardiovascular system, the gastrointestinal tract, and the eye (Jones et al., 2009). The principal research focus during the past decade, after the cloning of DP<sub>2</sub>, however, has been inflammation.

PGD<sub>2</sub> has long been known to cause eosinophil infiltration but with a pharmacological profile inconsistent

TABLE 10  
Recently identified potential therapeutic application of TP antagonists

Antagonist	Route	Dose	Species	Experimental Model	Indication	Reference
GR-32191 (vapiprost)	Oral (drinking water)	1 mg · kg <sup>-1</sup> · day <sup>-1</sup>	Mouse	STZ diabetes	Diabetic retinopathy	Wright et al., 2009
KP-496	Intratracheal	30 and 100 $\mu$ m/ animal	Rat	Sephadex induced airway inflammation	Asthma	Ishimura et al., 2009
KP-496	Intranasal	0.003–0.05%	Guinea pig	Ovalbumin, Japanese cedar pollen allergy	Allergic rhinitis	Mizutani et al., 2008
ONO-3708	Intradermal	0.1 and 1 nmol/site	Mouse	TxA <sub>2</sub> -induced scratching	Pruritus	Andoh et al., 2007
S18886 (terutroban)	Oral	10 and 30 mg/kg	Rat	Uninephrectomized obese Zucker rats	Type 2 diabetes	Sebeková et al., 2007

STZ, streptozotocin; ONO-3708, (1S-(1 $\alpha$ ,2 $\beta$ (Z),3 $\alpha$ (S\*),5 $\alpha$ ))-7-(3-(cyclopentylhydroxyacetyl)amino)-6,6-dimethylbicyclo(3.1.1)hept-2-yl)-5-heptenoic acid.

with DP<sub>1</sub> receptor mediation (Woodward et al., 1990, 1993b). The subsequent cloning of the receptor (Marchese et al., 1999; Nagata et al., 1999) and discovery of DP<sub>2</sub> receptor as a Th2 cell chemoattractant (Hirai et al., 2001) provided important new dimensions. The role of DP<sub>1</sub> and DP<sub>2</sub> receptors in immunology and inflammation has been the subject of more than one review (Herlong and Scott, 2006; Kostenis and Ulven, 2006; Pettipher, 2008) and this topic is therefore discussed only briefly.

The discovery of PGD<sub>2</sub> as an activator of Th2 lymphocytes via DP<sub>2</sub> (CRTH2) receptors implicated them in immune regulation. The polarization of lymphocytes to the Th-2 phenotype is typically implicated in the development of allergic responses such as asthma and atopic dermatitis. DP<sub>2</sub> receptor expression in CD4<sup>+</sup> T cells is low in healthy humans but enhanced in atopic subjects and correlates with the severity of the disease (Kostenis and Ulven, 2006). In addition to recruitment of Th2 cells, DP<sub>2</sub> receptor activation results in the production of cytokines such as IL-4, IL-5, IL-9, and IL-13 (Honda et al., 2003; Gallant et al., 2005; Xue et al., 2005; Kostenis and Ulven, 2006; Herlong and Scott, 2006). Recruitment and activation of eosinophils is a central feature of allergic responses, and they also express DP<sub>2</sub> receptors. Activation of DP<sub>2</sub> receptors leads to chemotaxis and degranulation of eosinophils (Gervais et al., 2001; Hirai et al., 2001; Monneret et al., 2001; Sugimoto et al., 2003; Böhm et al., 2004). DP<sub>2</sub> receptor stimulation produces eosinophil infiltrates in living animals, and these are located at sites typically associated with allergy, such as the conjunctiva (Woodward et al., 1990, 1993b), cornea (Fujishima et al., 2005), lung (Almishri et al., 2005; Shiraishi et al., 2005; Spik et al., 2005), and skin (Spik et al., 2005). An important role for DP<sub>2</sub> receptors in chronic cutaneous inflammation was revealed in DP<sub>2</sub>-deficient mice (Sato et al., 2006). Prostaglandin D<sub>2</sub> also activates basophils via DP<sub>2</sub> receptors (Hirai et al., 2001; Cossette et al., 2007). A final comment on the roles of DP<sub>1</sub> and DP<sub>2</sub> in regulating allergy/inflammation (Kostenis and Ulven, 2006; Pettipher, 2008): the employment of a DP<sub>2</sub> antagonist may switch PGD<sub>2</sub> from a pro- to an anti-inflammatory mediator in many instances.

Although widely distributed, notably in the gastrointestinal tract (Sawyer et al., 2002), the focus has been almost exclusively on leukocytes. Studies in the eye demonstrate that even within the context of allergic inflammation, DP<sub>2</sub> receptor stimulation produces more than leukocyte activation. Thus, DP<sub>2</sub> receptors are also associated with goblet cell depletion and increased microvascular permeability in the conjunctiva (Woodward et al., 1990, 1993b). In human retinal pigmented epithelial cells, DP<sub>2</sub> receptors induce heme oxygenase-1 expression (Satarug et al., 2008): these cells are essential for photoreceptor survival. DP<sub>2</sub> has been suggested as a therapeutic target for delaying the onset of age-related macular degeneration and cerebral malaria (Satarug et

al., 2008). Both DP receptor subtypes have been implicated in astrogliosis and demyelination and involved in the neuroinflammatory effects of PGD<sub>2</sub> (Mohri et al., 2006).

**3. Gene Deletion Studies.** DP<sub>2</sub> knockout mice were generated independently by two groups (Chevalier et al., 2005; Sato et al., 2006), and apparently different phenotypes were reported for these two lines. Chevalier et al. (2005) subjected their DP<sub>2</sub>-deficient mice to the OVA-induced allergic asthma model and found markedly increased eosinophil recruitment into the bronchoalveolar lavage (BAL) fluid of KO mice compared with WT mice. This was opposite to the known function of DP<sub>2</sub> (CRTH2), in that it mediates the chemotactic action of PGD<sub>2</sub> on eosinophils. Indeed, DP<sub>2</sub> receptor stimulation has been shown to increase the degree of inflammation in mice (Spik et al., 2005). To examine this discrepancy, they found that IL-5 production by activated T cells from DP<sub>2</sub>-deficient mice in vitro was increased compared with that observed with wild-type cells. They suggested that DP<sub>2</sub> indeed functions to facilitate allergy in situ at the site of inflammation, but that this receptor also regulates IL-5 production in the early phase of allergy development. On the other hand, Sato et al. (2006) injected anti-dinitrophenyl-specific IgE into the skin of the ear lobe of their DP<sub>2</sub>-deficient mice, challenged them with DNFB, and found that this type of IgE-induced dermatitis was significantly suppressed in DP<sub>2</sub>-deficient mice. They further showed that acute as well as chronic contact hypersensitivity was partially suppressed in DP<sub>2</sub>-deficient mice. In the Discussion section of their article, Sato et al. (2006) commented that their DP<sub>2</sub>-deficient mice did not exhibit enhanced eosinophilia when subjected to the asthma model and that splenocytes from their KO mice did not show exaggerated IL-5 production on activation. The DP<sub>2</sub>-deficient mice that Sato et al. (2006) generated consistently showed less inflammatory response in allergic dermatitis induced by cutaneous application of Japanese cedar pollen (Oiwa et al., 2008) or allergic rhinitis induced by repeated intranasal sensitization with Cry j 1, Japanese cedar pollen antigen (Nomiya et al., 2008). In the latter study, suppression of the increase in serum Cry j 1-specific IgE, a significant reduction of IL-4, and a slight reduction of IL-5 produced by draining lymph node cells in DP<sub>2</sub>-deficient mice were reported. Furthermore, Shiraishi et al. (2008) examined the role of DP<sub>2</sub> in the poly I:C-induced enhancement of allergic inflammation in the OVA asthma model and found that although DP<sub>2</sub>-deficient mice showed a comparable inflammatory eosinophil infiltration in BAL to WT mice after OVA challenge alone, they showed complete loss of the poly I:C-induced enhancement. No difference was found in the number of neutrophils and lymphocytes or the amounts of IL-5 and IL-13 in the BAL between WT and DP<sub>2</sub>-deficient mice. In addition to these studies focusing on the role of DP<sub>2</sub> in eosinophil recruitment in allergic reactions, Tajima et al. (2008)

found enhanced production of PGE<sub>2</sub> and PGD<sub>2</sub> during macrophage activation with LPS and examined the role of DP<sub>2</sub> in LPS-induced migration of peritoneal macrophages. They reported that LPS-mediated migration was significantly suppressed in macrophages from DP<sub>2</sub>-deficient mice.

**4. Agonists and Antagonists.** Potent and selective agonists for the DP<sub>2</sub> receptor have been discovered or devised. Although no therapeutic utility for DP<sub>2</sub> agonists has been established, they have provided invaluable pharmacological tools. Indeed, 13,14-dihydro,15-keto PGD<sub>2</sub> was the first DP<sub>2</sub> agonist to be discovered (Jones, 1976a,b), and over the past 3 decades, numerous unique natural ligands have been discovered (Pettipher, 2008). Synthetic selective DP<sub>2</sub> agonists include 15*R*-methyl PGD<sub>2</sub> (Monneret et al., 2003) and (9-((4-chlorophenyl)thio)-6-fluoro-2,3-dihydro-1*H*-pyrrolo(1,2-*a*)indol-1-yl)acetic acid (L-888,607) (Gervais et al., 2005). It is also interesting that indomethacin is a DP<sub>2</sub> agonist (Hirai et al., 2002).

Attaining selective DP<sub>2</sub> agonism has mainly involved alterations to the 15-hydroxyl group. The 15*S* stereochemistry is atypically not crucial to DP<sub>2</sub> agonism, with 15*R*- and 15*R*-15-methyl PGD<sub>2</sub> showing high DP<sub>2</sub>/DP<sub>1</sub> selectivity (Jones, 1976a,b, 1978; Monneret et al., 2001, 2003; Kim et al., 2005; Cossette et al., 2007). Moreover, 15-oxo structures (usually considered to be bioinactive products of 15-hydroxy PG dehydrogenase) retain high DP<sub>2</sub> agonism. The biologically active form of 15-oxo PGD<sub>2</sub> is not clear, however, because of its ready interconversion between 11,5-dioxo-13*E*-ene (as drawn in Fig. 2) and 11,15-dioxo-12-ene tautomers, of which there are two *E* and *Z* geometric isomers (Jones and Wilson, 1978); the corresponding conjugated enols (e.g., 11-oxo-12,14-diene-15-hydroxy) may even be active given the high potency of Δ<sup>12</sup>-PGJ<sub>2</sub> (Monneret et al., 2002). Saturation of the 13,14-double bond obviates this chemical lability (Jones, 1976a,b, 1978; Rangachari et al., 1995) as does conversion of the 15-oxo group to an ethylene ketal (Jones, 1978). Loss of the 15-oxygen function also results in retention of DP<sub>2</sub> agonism, as in 15-deoxy-Δ<sup>12,14</sup>-PGJ<sub>2</sub> (Monneret et al., 2002). The variance allowed at C15 is not unlimited, however; 15*S*-15-methyl PGD<sub>2</sub> shows only weak DP<sub>2</sub> agonism (Monneret et al., 2003). Finally, replacement of the 11-oxo group in PGD<sub>2</sub> by methylene essentially abolishes DP<sub>1</sub> agonism and results in weak DP<sub>2</sub> antagonism (Cossette et al., 2007).

DP<sub>2</sub> antagonists have been a major focus for drug discovery in recent years. This has been reviewed (Jones et al., 2009) and therefore is not discussed in depth herein. Several distinct structural classes have been used as design templates. Indomethacin being a DP<sub>2</sub> agonist (Hirai et al., 2002; Stubbs et al., 2002), indole acetic acids have been designed as antagonists. Likewise, the nonsteroidal anti-inflammatory drug fenclofenac was also used as a starting point for DP<sub>2</sub> antagonist design (Jones et al., 2009). Additional scaffolds include tetrahydroquinolines (Jones et al., 2009) and thia-

zoleacetic acids (Grimstrup et al., 2010). Ramatroban (Bay U 3405) has played a central role in DP<sub>2</sub> antagonist design, and elements of this core structure are incorporated into many structures (Jones et al., 2009), including recently described indole-based DP<sub>2</sub> antagonists (Stearns et al., 2009). Ramatroban is also a TP antagonist and is marketed for the treatment of allergic rhinitis (Sugimoto et al., 2003). Development of ramatroban was first directed toward TP antagonism (McKenniff et al., 1988). Later studies revealed low-affinity DP<sub>2</sub> antagonism (pA<sub>2</sub>, 7.44) (Sugimoto et al., 2003; Mathiesen et al., 2006). Modification of ramatroban led to TM-30089 (CAY-10471; Fig. 3), which has much higher DP<sub>2</sub>/TP-selectivity (Ulven and Kostenis, 2005). In assays of guanosine 5'-O-(3-[<sup>35</sup>S]thio)triphosphate binding/inositol phosphate accumulation and PGD<sub>2</sub>-induced eosinophil shape change, ramatroban and the related TM-30642 were surmountable competitive antagonists, whereas TM-30643 and TM-30089 (Ulven and Kostenis, 2005) show insurmountability. Indomethacin has also provided a template for DP<sub>2</sub> blockers, based to some extent on its weak agonism for the DP<sub>2</sub> receptor (Hirai et al., 2002; Stubbs et al., 2002). Many of these analogs are designed around an inverted indole template (Birkinshaw et al., 2006). This latter profile may be due to slow dissociation from the DP<sub>2</sub> receptor (Mathiesen et al., 2006). The embodiment of both DP<sub>1</sub> and DP<sub>2</sub> receptor antagonism in a single molecule seems to offer a potentially more effective therapeutic approach (Pettipher, 2008). This approach for antiallergic drug design is in progress, with at least one positive outcome in the form of AMG 009 (Liu et al., 2009).

**5. Therapeutics.** A variety of DP<sub>2</sub> antagonists have been claimed active in models of allergic rhinitis, asthma, and atopic dermatitis (Table 11). Lung inflammation resulting from smoke inhalation has recently reported to be reduced by a DP<sub>2</sub> antagonist (Stebbins et al., 2010).

### G. Receptor Heterodimerization

It is now established that G protein-coupled receptors exist as dimers (Prinster et al., 2005; Lohse, 2010) that are precoupled to heterotrimeric proteins in living cell membranes (Nobles et al., 2005). IP receptor precoupling to G<sub>s</sub> has been shown in the cell membrane (Nobles et al., 2005). Heterodimerization allows the receptor to extend its repertoire of G protein coupling and ligand binding. IP/TP heterodimerization provides an example of both phenomena. In terms of second-messenger signaling, PGI<sub>2</sub>-like characteristics are conferred on TxA<sub>2</sub> mimetics, which occurs as a robust cAMP response (Wilson et al., 2004). IP/TP heterodimerization creates a new binding site, which recognizes isoprostane E<sub>2</sub> (Wilson et al., 2004). Heterodimerization of a prostanoid receptor with a member outside of the prostanoid family has been observed in the form of EP<sub>1</sub>/β<sub>2</sub>-adrenoceptor dimerization (McGraw et al., 2006).



TABLE 11  
Therapeutic applications of DP<sub>2</sub> antagonists

DP <sub>2</sub> antagonist	Route	Dose mg/kg	Species	Experimental Model	Indication	Reference
AM 156	Oral	10	Mouse	Smoke exposure	COPD	Stebbins et al., 2010
Compound 23	Oral	10	Mouse	Ovalbumin allergy	Allergic rhinitis	Stearns et al., 2009
Compound A	Oral	10	Mouse	FITC-cutaneous allergy	Atopic dermatitis	Boehme et al., 2009
Ramatroban	Oral	5	Mouse	Allergic (ovalbumin) eosinophilic airway inflammation	Asthma	Uller et al., 2007
Ramatroban	Oral	30	Mouse	Cry $\gamma$ 1-induced pollinosis	Allergic rhinitis	Nomiya et al., 2008
TM-30089	Oral	5	Mouse	Allergic (ovalbumin) eosinophilic airway inflammation	Asthma	Uller et al., 2007

AM 156, (2'-((cyclopropanecarbonylethylamino)methyl)-6-methoxy-4'-trifluoromethyl-biphenyl-3-yl)acetic acid; COPD, chronic obstructive pulmonary disease; FITC, fluorescein isothiocyanate.

Heterodimerization of alternative mRNA splicing variants may also occur. Isoprostane responses were enhanced when TP $\alpha$  and TP $\beta$  were coexpressed (Wilson et al., 2007). Coexpression of the wild-type FP receptor and an alternative mRNA splicing variant resulted in ligand recognition that was quite different from wild-type FP receptors (Liang et al., 2008). Unlike FP receptors, wild-type/alternative FP/FP heterodimers were able to respond to the prostamide F<sub>2 $\alpha$</sub>  mimetic bimatoprost, and the antagonist AGN 211335 (Liang et al., 2008) blocked responses to bimatoprost but not PGF<sub>2 $\alpha$</sub> . This pharmacology was entirely consistent with that observed for the prostamide receptor in ocular cells and isolated tissue preparations (Liang et al., 2003; Woodward et al., 2003, 2007; Matias et al., 2004; Spada et al., 2005; Wan et al., 2007; Stamer et al., 2010). Therapeutic uses for the prostamide F<sub>2 $\alpha$</sub>  mimetic bimatoprost have extended beyond glaucoma to include trichomegaly (Tauchi et al., 2010). No potential medical uses for prostamide antagonists (Woodward et al., 2009) have been reported.

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Wrote or contributed to the writing of the manuscript: Woodward, Jones, and Narumiya.

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