

International Union of Pharmacology. XX. Current Status of the Nomenclature for Nicotinic Acetylcholine Receptors and Their Subunits

RONALD J. LUKAS,¹ JEAN-PIERRE CHANGEUX, NICOLAS LE NOVÈRE, EDSON X. ALBUQUERQUE, DAVID J. K. BALFOUR, DARWIN K. BERG, DANIEL BERTRAND, VINCENT A. CHIAPPINELLI, PAUL B. S. CLARKE, ALLAN C. COLLINS, JOHN A. DANI, SHARON R. GRADY, KENNETH J. KELLAR, JON M. LINDSTROM, MICHAEL J. MARKS, MARYKA QUIK, PALMER W. TAYLOR, AND SUSAN WONNACOTT

Division of Neurobiology, Barrow Neurological Institute, Phoenix, Arizona (R.J.L.); Neurobiologie Moleculaire, Institut Pasteur, Paris, France (J.-P.C., N.L.N.); Department of Pharmacology and Experimental Therapeutics, University of Maryland School of Medicine, Baltimore, Maryland (E.X.A.); Department of Pharmacology, University Medical School, University of Dundee, Dundee, United Kingdom (D.J.K.B.); Department of Biology, University of California at San Diego, La Jolla, California (D.K.B.); Department of Physiology, University of Geneva/Central Medical University, Geneva, Switzerland (D.B.); Department of Pharmacology, George Washington University School of Medicine, Washington, D.C. (V.A.C.); Department of Pharmacology and Therapeutics, McGill University, Montreal, Quebec, Canada (P.B.S.C.); Institute for Behavioral Genetics, University of Colorado, Boulder, Colorado (A.C.C., S.R.G., M.J.M.); Division of Neuroscience, Baylor College of Medicine, Houston, Texas (J.A.D.); Department of Pharmacology, Georgetown University School of Medicine, Washington, D.C. (K.J.K.); Institute for Neurological Sciences, University of Pennsylvania, Philadelphia, Pennsylvania (J.M.L.); Parkinson's Institute, Sunnyvale, California (M.Q.); Department of Pharmacology, School of Medicine, University of California at San Diego, La Jolla, California (P.W.T.); Department of Biology and Biochemistry, University of Bath, Bath, United Kingdom (S.W.)

This paper is available online at <http://www.pharmrev.org>

I. Introduction	397
II. Current status of nACh receptor subunit nomenclature and receptor type nomenclature and classification	398
III. Present definitions of nACh receptor subunits and receptor types	399
IV. Bibliography	400

I. Introduction

Nicotinic acetylcholine receptors (nACh receptors²) in jawed vertebrates are prototypical members of the multisubunit, neurotransmitter-gated superfamily of ion channels (ionotropic neurotransmitter receptors; see *Section IV* for selected seminal papers, reviews, and collections). nACh receptors mediate some of the effects of the endogenous neurotransmitter, acetylcholine, and they are principal biological targets of the tobacco alkaloid, nicotine, which generally (although exceptions exist) mimics acute actions of acetylcholine at nACh receptors. nACh receptors play critical physiological roles throughout the brain and body, mediating excitatory neurotransmission at the vertebrate neuromuscular junction, across autonomic ganglia, and at selected synapses in the brain and spinal cord. Roles have also been suggested for nACh receptors in the modulation of neurotransmitter release as well as in neurotrophism.

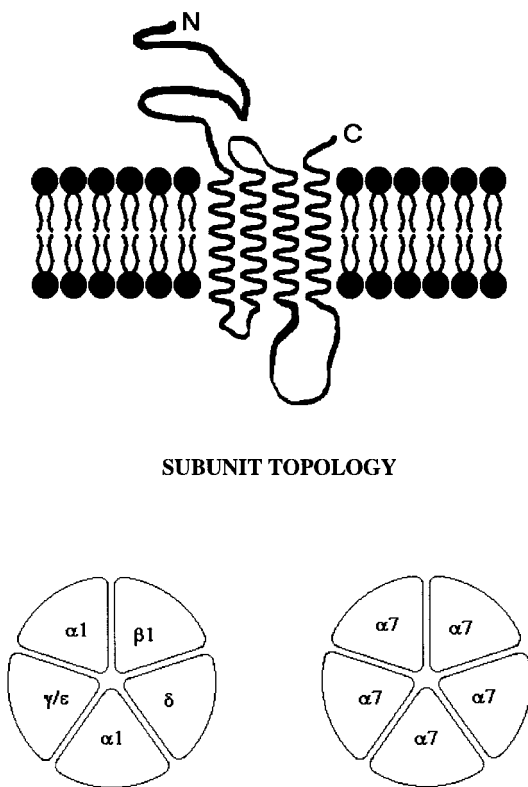
¹ Address for correspondence: Ronald J. Lukas, Division of Neurobiology, Barrow Neurological Institute, 350 West Thomas Rd., Phoenix, AZ 85013. E-mail: rlukas@mha.chw.edu

² Abbreviations: nACh receptor(s), nicotinic acetylcholine receptor(s); NC-IUPHAR, International Union of Pharmacology Committee on Receptor Nomenclature and Drug Classification; 5-HT, 5-hydroxytryptamine (serotonin); CNS, central nervous system.

nACh receptors exist as a variety of types. At least one nACh receptor type is found in developing skeletal muscle, and another is expressed at the mature neuromuscular junction. There is evidence that several nACh receptor types exist in peripheral neurons of autonomic and sensory ganglia and in the brain and spinal cord. Evidence also exists for expression of nACh receptor types in other tissues and cell types, including lymphocytes, fibroblasts, pulmonary neuroendocrine cells, spermatozoa, keratinocytes, granulocytes, chondrocytes, and placenta, as well as in several sensory organs.

Classical pharmacological distinctions between some nACh receptor types endure. For example, decamethonium is a selective inhibitor of nACh receptor types in muscle, and hexamethonium is a selective antagonist of nACh receptor types in autonomic ganglia. More contemporary studies are beginning to yield distinctive pharmacological profiles for some nACh receptor types. However, these findings are not yet clearly integrated with molecular definitions of those receptors. This precludes a pharmacological description of nACh receptor types in this report, which instead focuses on matters of nomenclature and on structural bases of nACh receptor diversity.

Heterogeneity or diversity in nACh receptor types is derived in part from diversity in genes that encode nACh receptor subunits. Diverse nACh receptor types are formed as pentamers from different combinations of genetically distinct subunits, but not from all possible combinations of subunits (Fig. 1 and Table 1). Subunits confer distinctive functional and structural properties to the nACh receptor types that they form. With few exceptions, subunit compositions, stoichiometries, and arrangements of naturally expressed nACh receptor types are not known with absolute certainty.



SUBUNIT TOPOLOGY

RECEPTOR ASSEMBLY

FIG. 1. Schematic diagrams showing (upper) transmembrane topology of nAChR subunits and (lower) features of nAChR subunit assembly. Upper schematic illustrates the existence of an extensive, extracellular N-terminal region, four transmembrane segments, a cytoplasmic loop, and an extracellular C terminus for each nACh receptor subunit (not drawn to scale; surrounding lipid bilayer is also represented schematically). Lower schematic illustrates known arrangements of subunits constituting a nACh receptor type predominant in fetal skeletal muscle (left) and constituting a homooligomeric nACh receptor composed of $\alpha 7$ subunits (right). Assembly of subunits is thought to be regulated and limited by interactions between amino acid residues at subunit interfaces. Ligand binding pockets are thought to be formed at a subset of these assembly interfaces. For example, two distinct classes of ligand binding sites are formed at interfaces between $\alpha 1$ and γ subunits or between $\alpha 1$ and δ subunits of the nACh receptor from fetal muscle. Five largely similar ligand binding domains are thought to be formed at negative and positive faces of $\alpha 7$ subunits in homooligomeric $\alpha 7$ -nACh receptors. All subunits contribute to channel formation, reflected by influences of every nACh receptor subunit identified to date on features of channel activity upon interaction with nicotinic ligand (see Table 1).

TABLE 1
Subunit compositions of well established, naturally expressed and heterologously expressed nACh receptors

Naturally Expressed nACh Receptors		Heterologously Expressed nACh Receptors
$(\alpha 1)_2\beta 1\gamma\delta$ $(\alpha 1)_2\beta 1\epsilon\delta$	Fetal skeletal muscle Adult skeletal muscle	$\alpha 1\beta 1\gamma\delta$ $\alpha 1\beta 1\epsilon\delta$ $\alpha 1\gamma\delta$ plus $\beta 2$ or $\beta 4$ $\alpha 2$ plus $\beta 2$ or $\beta 4$ $\alpha 3$ plus $\beta 2$ or $\beta 4$ $\alpha 3\alpha 5\beta 4$ $\alpha 3\beta 3\beta 4$ $\alpha 3\alpha 5\beta 2$
$\alpha 3\beta 4^*$ $\alpha 3\alpha 5\beta 4$	Autonomic ganglia Autonomic ganglia	$\alpha 4$ plus $\beta 2$ or $\beta 4$ $\alpha 4\beta 2\beta 4$ $\alpha 4\beta 2\beta 3\beta 4$ $\alpha 4\alpha 5\beta 2$ $\alpha 6$ plus $\beta 2$ or $\beta 4$
$\alpha 3\alpha 5\beta 2\beta 4$ $(\alpha 4)_2(\beta 2)_3$	Autonomic ganglia CNS	$\alpha 7$ $\alpha 7\alpha 8^*$ $\alpha 8^*$ $\alpha 9^*$
$\alpha 4\alpha 5\beta 2$	CNS	$\alpha 7$ plus $\beta 2$ or $\beta 4$ $\alpha 7$ $\alpha 7\alpha 8$ (chick) $\alpha 8$ (chick) $\alpha 9$
$\alpha 7^*$ $\alpha 7\alpha 8^*$ $\alpha 8^*$ $\alpha 9^*$	Autonomic ganglia, CNS Chick retina, chick CNS Chick retina Cochlea, pituitary	

Subunit compositions, stoichiometries if known, and principal locations (not exhaustively, but focusing on selected central, sensory, autonomic, and/or somatomotor nervous system locations) are indicated for well-characterized, naturally expressed nACh receptors. Also shown are subunit compositions known to form distinctive, functional receptor-channel complexes in heterologous expression systems based on studies using subunits derived from several species including man, rat, mouse, chick, and/or electric ray. Note that $\alpha 8$ subunits and receptors containing them have only been identified in chick.

To date, 16 nACh receptor subunit genes have been cloned from vertebrates (Table 1). Each of the nACh receptor subunit proteins encoded by these genes is thought to have the same, general structural organization. The features of these integral membrane proteins include an extensive N-terminal region positioned extracellularly, four transmembrane segments (M1–M4), an extended cytoplasmic loop between the third and fourth transmembrane segments containing amino acid sequences absolutely unique to each subunit, and an extracellular C terminus (Fig. 1). All nACh receptor subunits also contain a loop of 13 amino acids between two cysteine residues forming a disulfide bridge in the N-terminal domain. These features are shared with families of subunits that constitute ionotropic glycine receptors, ionotropic γ -amino butyric acid receptors, invertebrate glutamate-gated chloride channels, and ionotropic serotonin receptors (5-HT₃ receptors). Invertebrate nACh receptor subunits can combine with vertebrate nACh receptor subunits to form functional receptors, suggesting that the former also belong to the superfamily. Comparisons between rat and human amino acid sequences typically reveal over 80% identity for a given nACh receptor subunit.

II. Current Status of nACh Receptor Subunit Nomenclature and Receptor Type Nomenclature and Classification

The IUPHAR Subcommittee on Nicotinic Acetylcholine Receptors of the International Union of Pharmacology Committee on Receptor Nomenclature and Drug Classification (NC-IUPHAR) recommends continued use of the present, Greek letter-based nomenclature for

nACh receptor subunits, as is described in *Section III*, for the foreseeable future. The subcommittee also recommends using the suffix “gene” to distinguish the gene encoding a given subunit from the subunit protein itself (e.g., the δ subunit gene encodes the δ subunit). In addition, the subcommittee recommends for the foreseeable future a minimal nACh receptor type nomenclature/classification system based on the subunits that compose those receptor types to the extent that such information is available. Examples of such a nACh receptor type nomenclature and classification are also presented in *Section III*.

The NC-IUPHAR subcommittee consensus is that it is premature to create any alternative to the nomenclature for nACh receptor subunits or receptor types already in place or to generate a classification of nACh receptor types based on any of their properties other than their subunit composition. This consensus is based partly on the subcommittee’s perspectives that there are probably more nACh receptor subunit genes to be cloned, that there likely are more nACh receptor types to be identified, and that the current understanding is deficient about subunit compositions and other properties of many nACh receptor types. Although ligand binding properties, functional characteristics, and schemes for assembly of nACh receptor types differ, the subcommittee’s view is that more information is needed before these features can be exploited as criteria for an enduring system of nACh receptor type classification. The subcommittee’s consensus also recognizes the broad acceptance across the research community of the nACh receptor subunit and receptor type nomenclature/classification system already in place; there are very few deviations from this nomenclature in the literature. Moreover, the subcommittee’s opinion is that the nomenclature/classification system currently in use is reasonably sensible and understandable to the newly initiated, particularly given the inherent complexity of multisubunit, ionotropic neurotransmitter receptors relative to metabotropic neurotransmitter receptors composed of a single polypeptide. The subcommittee acknowledges limitations in the present system and that it is at variance with several of the guidelines established by NC-IUPHAR. For example, NC-IUPHAR prefers to avoid use of Greek letters but bows to historical precedence in this instance. However, the subcommittee does not see any obvious advantages to adoption of a new system at the present time, and the current system is viewed as having flexibility adequate to accommodate at least another series of advances in our understanding of the nACh receptor system. Nevertheless, it is anticipated that research progress over the next few years, including completion of the human genome project and identification of all nACh receptor subunit genes, will ultimately allow rational generation of enduring nACh receptor subunit nomenclature and nACh receptor type nomenclature/classification systems.

The NC-IUPHAR subcommittee recommends that commonly used prefixes such as “neuronal nACh receptor”, “muscle nACh receptor”, or “ganglionic/autonomic nACh receptor” not be adopted into nACh receptor nomenclature or classification. This is because there are many possible nACh receptor subunits or receptor types that can be found in any one of these tissues and because some nACh receptor subunits or receptor types may be found in more than one of these tissues. Such terminology applied to nACh receptor-directed, curaremimetic, snake neurotoxins (“neuronal-bungarotoxin”) instead of a neutral system (such as one based on Greek letters) has been misleading at best.

Similarly, it is recommended that use of the prefix “structural” not be propagated with regard to classification of nACh receptor subunits. This term currently has little use or meaning because evidence exists that each of the nACh receptor subunits identified to date participates in formation of ligand binding pockets and/or influences nACh receptor functional properties. Continued use of the term “non- α subunit” is also discouraged.

III. Present Definitions of nACh Receptor Subunits and Receptor Types

The 16 nACh receptor subunits identified to date are defined using a Greek letter sometimes followed by an Arabic numeral (neither subscripted nor superscripted). For example, there are nine alpha subunits ($\alpha 1$ – $\alpha 9$; $\alpha 8$ subunits have only been derived from chick), four beta subunits ($\beta 1$ – $\beta 4$), and one each gamma (γ), delta (δ), and epsilon (ϵ) subunit. All nACh receptor α subunits share expression of a pair of tandem cysteine residues in the putative, N-terminal, extracellular region. These residues are near to a site on the $\alpha 1$, $\alpha 4$, or $\alpha 7$ subunits known to engage in agonist binding. None of the other subunits (i.e., $\beta 1$ – $\beta 4$, γ , δ , or ϵ) have these tandem cysteines. Several approaches have been taken in attempts to illuminate relationships between nACh receptor subunits and their genes. At a low level of resolution, these analyses make it clear that same nACh receptor subunits and their genes are most closely related (e.g., the $\beta 1$, γ , δ , and ϵ grouping; uniqueness of the $\alpha 7/\alpha 8$ grouping and of the $\alpha 9$ subunit; grouping into an extended family of $\alpha 2/\alpha 4$ and $\alpha 3/\alpha 6$ pairs). However, each of these analyses is unique with respect to the range of amino acid or nucleic acid sequences examined and the analytical/computational tools applied, and each has produced somewhat different results, particularly with regard to placement of the $\alpha 1$ subunit and the $\beta 2/\beta 4$ and $\alpha 5/\beta 3$ subunit pairs. The NC-IUPHAR subcommittee consensus is that these analyses are interesting and help guide those working in the field. However, further evolution in these approaches and identification and sequencing of nACh receptor subunits and their genes are needed to help guide the subcommittee and NC-IUPHAR in future efforts to derive nACh receptor subunit and receptor type classification/nomenclature.

It is useful to point out that the grouping of $\beta 1$, γ , δ , and ε subunits is relevant to their contributions in formation of nACh receptor types in muscle, in combination with the $\alpha 1$ subunit. nACh receptors in vertebrate fetal muscle contain two $\alpha 1$ subunits and one each $\beta 1$, δ , and γ subunit, whereas the ε subunit substitutes for the γ subunit in nACh receptors found in adult muscle. Thus, in vertebrates, one form of nACh receptor found in fetal muscle can be defined as the $\alpha 1\beta 1\gamma\delta$ -nACh receptor, and one form of nACh receptor found in adult muscle can be defined as the $\alpha 1\beta 1\varepsilon\delta$ -nACh receptor. Subscript and parenthetical notation is used to define subunit stoichiometries [e.g., $(\alpha 1)_2\beta 1\gamma\delta$ -nACh receptors for nACh receptors containing two copies of the $\alpha 1$ subunit and single copies of $\beta 1$, γ , and δ subunits]. Note the caveat here with respect to nACh receptor subunits and types found in muscle, as perhaps will be the case with regard to other nACh receptor subunits and types, that there is evidence for alternatively spliced products of $\alpha 1$ and γ subunit genes. Thus, there may be more than one kind of nACh receptor found in fetal or adult muscle, and a formal nACh receptor subunit and receptor type nomenclature will need to accommodate and define features of subunit variants and the nACh receptor types containing them.

The grouping of the $\alpha 7/\alpha 8$ subunit pair and the separation of the $\alpha 9$ subunit from the others is relevant in that nACh receptors containing these subunits happen to be capable of interaction with longer forms of curaremimetic, snake neurotoxins. These subunits also have the capacity to combine as homooligomers to create functional nACh receptor types. nACh receptors known to form only as homooligomers are defined, for example, as $\alpha 7$ -nACh receptors, $\alpha 8$ -nACh receptors, or $\alpha 9$ -nACh receptors, with stoichiometries noted in subscript if known [e.g., $(\alpha 7)_5$ -nACh receptors]. nACh receptors containing both $\alpha 7$ and $\alpha 8$ subunits have been isolated from chick brain and/or retina and can be defined as $\alpha 7\alpha 8$ -nACh receptors. Moreover, there is additional evidence that some nACh receptors might contain $\alpha 7$ plus other kinds of subunits. If, for example, nACh receptors known to contain $\alpha 7$ subunits possibly contain or are known to contain additional kinds of subunits as a pure or mixed population of receptors, the subcommittee recommends a notation employing an asterisk as a "wild card" to define those entities as $\alpha 7^*$ -nACh receptors.

There is strong evidence that $\alpha 3$ and $\beta 4$ subunits coexist in naturally expressed nACh receptors found in autonomic ganglia and derived cell lines. There is additional evidence that these naturally expressed nACh receptors in autonomic neurons also contain $\alpha 5$ subunits, consistent with the fact that genes encoding $\alpha 3$, $\alpha 5$, and $\beta 4$ subunits are clustered. Therefore, a more specific definition of these receptors would be $\alpha 3\alpha 5\beta 4$ -nACh receptors. Moreover, there is evidence that a subset of nACh receptors containing $\alpha 3$, $\alpha 5$, and $\beta 4$ subunits

also contain $\beta 2$ subunits; these receptors can be defined as $\alpha 3\alpha 5\beta 2\beta 4$ -nACh receptors. If a pure or mixed population of nACh receptors is known to contain $\alpha 3$ or $\alpha 3$ plus $\beta 4$ subunits, but the identity of coassembled subunits is not certain, the subcommittee recommends use of $\alpha 3^*$ -nACh receptors or $\alpha 3\beta 4^*$ -nACh receptors, respectively, to describe those entities.

A numerically abundant nACh receptor type in the central nervous system (CNS) is composed of $\alpha 4$ and $\beta 2$ subunits, and data supporting a 2:3 stoichiometry exists. Thus, these receptors can be defined as $\alpha 4\beta 2$ -nACh receptors [or as $(\alpha 4)_2(\beta 2)_3$ -nACh receptors as information warrants]. However, minor populations of nACh receptors containing $\alpha 4$ and $\beta 2$ subunits known to or possibly containing additional subunits can be defined employing the asterisk/wild card notation as $\alpha 4\beta 2^*$ -nACh receptors.

The subunit compositions of nACh receptors naturally expressed at lower abundance in brain are not known, although there must be some roles for $\alpha 2$, $\alpha 3$, $\alpha 5$, $\alpha 6$, $\beta 3$, and $\beta 4$ subunits in formation of other nACh receptor types. Considerable evidence exists suggesting natural expression of functional nACh receptors in brain and spinal cord other than those containing just $\alpha 7$ or just $\alpha 4$ and $\beta 2$ subunits, but further work is needed to determine the subunit compositions and stoichiometries of these entities. From heterologous expression studies, it is known that unique, ligand binding pockets can be formed at interfaces between $\alpha 2$, $\alpha 3$, $\alpha 4$, or $\alpha 6$ subunits and $\beta 2$ or $\beta 4$ subunits. Presence of $\alpha 5$ or $\beta 3$ subunits has also been shown to influence ligand recognition of heterologously expressed nACh receptors containing other subunits. Heterologously expressed nACh receptors containing these subunits can be defined in a straightforward fashion, as knowledge about contributions of subunits to expressed nACh receptor warrants (e.g., $\alpha 6\beta 4$ -nACh receptors if composed only of $\alpha 6$ and $\beta 4$ subunits). Similarly, at least provisional definitions of naturally expressed nACh receptors can be generated as information about the subunits constituting those nACh receptor is progressively obtained (e.g., $\alpha 2^*$ -nACh receptors for receptors known to contain $\alpha 2$ subunits).

Acknowledgments. We thank J. Brek Eaton of the Barrow Neurological Institute for composing Fig. 1.

BIBLIOGRAPHY

- Albuquerque EX (1996) Properties of neuronal nicotinic acetylcholine receptors: Pharmacological characterization and modulation of synaptic function. *J Pharmacol Exp Ther* **280**:1117–1136.
- Barrantes FJ, ed (1998) *The Nicotinic Acetylcholine Receptor: Current Views and Future Trends*, Springer Verlag, Berlin/Heidelberg and Landes Publishing Co., Georgetown, TX.
- Changeux J-P, Devillers-Thiery A, Galzi J-L and Bertrand D (1992) The functional architecture of the acetylcholine receptor explored by affinity labeling and site-directed mutagenesis. *Q Rev Biophys* **25**:395–432.
- Clarke PBS, Quik M, Adlkofer F and Thurau K, eds (1995) *Effects of Nicotine on Biological Systems II*, Birkhauser Verlag, Basel.
- Gotti C, Fornasari D and Clementi F (1997) Human neuronal nicotinic receptors. *Prog Brain Res* **53**:199–237.

- Heinemann S, Boulter J, Connolly J, Deneris E, Duvoisin R, Hartley M, Hermans-Borgmeyer I, Hollman M, O'Shea-Greenfield A, Papke R, Rogers S and Patrick J (1989) The nicotinic receptor genes. *Clin Neuropharmacol* **14**:S45-S61.
- Karlin A and Akabas MH (1995) Toward a structural basis for the function of nicotinic acetylcholine receptors and their cousins. *Neuron* **15**:1231-1244.
- Le Novère N and Changeux J-P (1995-97) <http://www.pasteur.fr/units/neubiomol/LGIC.html>.
- Lindstrom J (1996) Neuronal nicotinic acetylcholine receptors, in *Ion Channels* (Narahashi T ed), vol 4, pp 377-450, Plenum Press, New York.
- Nordberg A, Fuxe K, Holmstedt B and Sundwall, eds (1989) Nicotinic receptors in the CNS: Their role in synaptic transmission. *Prog Brain Res* **79**:1-366.
- Numa S, Noda M, Takahashi H, Tanabe T, Toyosato M, Furutani Y and Kikuyotani S (1983) Molecular structure of the nicotinic acetylcholine receptor. *Cold Spring Harb Symp Quant Biol* **48**:57-70.
- Papke RL (1993) The kinetic properties of neuronal nicotinic receptors: Genetic basis of functional diversity. *Prog Neurobiol* **41**:509-531.
- Raftery M, Hunkapillar M, Strader C and Hood L (1980) Acetylcholine receptor, complex of homologous subunits. *Science* **208**:1454-1457.
- Sargent P (1993) The diversity of neuronal nicotinic acetylcholine receptors. *Annu Rev Neurosci* **16**:403-443.
- Williams M and Triggle DJ, eds-in-chief; Levin ED, Rose JE, Lippello P, Robinson J, eds (1996) Nicotine Addiction. *Drug Dev Res* **38**:135-304.